



Epidermolysis bullosa in Danish Hereford calves is caused by a deletion in *LAMC2* gene

Murgiano *et al.*

RESEARCH ARTICLE

Open Access

Epidermolysis bullosa in Danish Hereford calves is caused by a deletion in *LAMC2* gene

Leonardo Murgiano¹, Natalie Wiedemar¹, Vidhya Jagannathan¹, Louise K Isling², Cord Drögemüller¹ and Jørgen S Agerholm^{2,3*}

Abstract

Background: Heritable forms of epidermolysis bullosa (EB) constitute a heterogeneous group of skin disorders of genetic aetiology that are characterised by skin and mucous membrane blistering and ulceration in response to even minor trauma. Here we report the occurrence of EB in three Danish Hereford cattle from one herd.

Results: Two of the animals were necropsied and showed oral mucosal blistering, skin ulcerations and partly loss of horn on the claws. Lesions were histologically characterized by subepidermal blisters and ulcers. Analysis of the family tree indicated that inbreeding and the transmission of a single recessive mutation from a common ancestor could be causative. We performed whole genome sequencing of one affected calf and searched all coding DNA variants. Thereby, we detected a homozygous 2.4 kb deletion encompassing the first exon of the *LAMC2* gene, encoding for laminin gamma 2 protein. This loss of function mutation completely removes the start codon of this gene and is therefore predicted to be completely disruptive. The deletion co-segregates with the EB phenotype in the family and absent in normal cattle of various breeds. Verifying the homozygous private variants present in candidate genes allowed us to quickly identify the causative mutation and contribute to the final diagnosis of junctional EB in Hereford cattle.

Conclusions: Our investigation confirms the known role of laminin gamma 2 in EB aetiology and shows the importance of whole genome sequencing in the analysis of rare diseases in livestock.

Keywords: Cattle, Epidermolysis bullosa, Laminin gamma 2, Hereditary, Congenital, Skin

Background

Heritable forms of epidermolysis bullosa (EB) constitute a heterogeneous group of skin disorders of genetic aetiology that are characterised by skin and mucous membrane blistering and ulceration in response to even minor trauma. EB is classified into four major types based on the level of blister formation in the dermo-epidermal interface, i.e. within the epidermis, basement membrane zone or uppermost dermis. In EB simplex, blisters develop within the epidermis, while for junctional and dystrophic EB cleavage occurs in the lamina lucida or below the lamina densa, respectively. The fourth major type, Kindler

Syndrome, is characterized by blisters in the lamina lucida and below the lamina densa. In humans, many subtypes of which some have extracutaneous lesions have been identified [1-4].

In contrast to the situation in man, where more than 1000 mutations in at least 18 genes encoding structural proteins have been associated with EB and thousands of EB patients have been thoroughly diagnosed [3,4], rather few cases have been characterized to the molecular level in domestic animals. In cattle, EB simplex was associated with a mutation in *keratin 5* [5] and dystrophic EB was associated with *COL7A1* mutations in cattle [6] and dogs [7] while junctional EB has been diagnosed in sheep (*LAMC2* mutation) [8], horses (*LAMC2* and *LAMA3* mutations) [9-11], and dogs (*LAMA3* mutation) [12]. In addition to these, genetically uncharacterized EB cases in animals have been reported [13].

In addition to genetically characterized cases in cross-bred Holstein calves [5] and Rotes Höhenvieh cattle [6],

* Correspondence: jager@sund.ku.dk

²Department of Veterinary Disease Biology, Section for Veterinary Pathology, Faculty of Health and Medical Sciences, University of Copenhagen, Ridebanevej 3, DK-1870 Frederiksberg C, Denmark

³Department of Large Animal Sciences, Section for Veterinary Reproduction and Obstetrics, Faculty of Health and Medical Sciences, University of Copenhagen, Dyrslægevej 68, DK-1870 Frederiksberg C, Denmark
Full list of author information is available at the end of the article

sporadic [14-17] and multiple genetically linked cases within single herds have been reported [18-20]. In these cases, a presumptive diagnosis of EB was based on presence of congenital blistering of the skin and mucous membranes and histopathological detection of dermo-epidermal cleavage and in some cases supported by transmission electron microscopy (TEM) findings. Here we report the occurrence of EB in a Danish herd of Hereford cattle and its genetic characterization using positional cloning and whole genome sequencing.

Methods

Cases

The first case of EB, a female Hereford calf with a body weight of 30 kg, was born in March 2007 (case 1). The calf was euthanized four days old by intravenous injection of an overdose of pentobarbital sodium and submitted for necropsy. A second case of unregistered sex was stillborn in December 2007 (case 2). EB was diagnosed retrospectively based on the owner's description of lesions as the calf was destroyed. The third case was a male Hereford calf with a body weight of 48 kg that died immediately after parturition in July 2009 (case 3) and was submitted for necropsy. The herd consisted of four breeding females in 2007. None of the parents of affected calves had signs of a blistering skin disorder. The study was performed according to Danish legislation and the cases published with the consent of the owner.

Pathology

A complete necropsy was performed in both calves and specimens of skin and mucous membranes were sampled for histology. Samples were taken from within lesions, from the border between lesions and adjacent grossly normal tissue and from normal skin areas distant to lesions. Specimens were fixed in 10% neutral buffered formalin, processed by routine methods, embedded in paraffin, sectioned at 5 μ m, and stained by haematoxylin and eosin. Selected sections were stained with periodic acid-Schiff (PAS).

DNA samples and genotyping

Four-generation pedigrees of the cases were obtained from the Danish Cattle Database and analysed for inbreeding loops. Tissue samples were collected from the one of the available affected calves (case 1). In addition, blood samples were gathered from both parents of case 1 and the sire of case 3. DNA was extracted using standard methods. Genotyping of these animals was performed using the BovineHD BeadChip (illumina), including 777,961 evenly distributed single nucleotide polymorphisms (SNPs) and standard protocols as recommended by the manufacturer.

Homozygosity mapping

PLINK software [21] was used to search for extended intervals of homozygosity with shared alleles as described previously [22]. Individuals and SNPs were selected using the commands `--keep`, and `--extract` while final files were generated through the `--merge` command. Homozygosity analysis was carried out on all cases using the commands `--cow`, `--homozyg` and `--homozyg-group`.

Whole genome re-sequencing and searching for variants

A fragment library with a 300 bp insert size was prepared and collected in a single lane of Illumina HiSeq2500 paired-end reads (2×100 bp); the fastq files were created using Casava 1.8. We obtained a total of 487,657,379 paired-end reads, which were then mapped to the cow reference genome UMD3.1/bosTau6 and aligned using Burrows-Wheeler Aligner (BWA) version 0.5.9-r16 [23] with default settings. The mapping showed that 403,122,849 reads had unique mapping positions. The SAM file generated by BWA was then converted to BAM and the reads sorted by chromosome using samtools [24]. Polymerase chain reaction (PCR) duplicates were marked using Picard tools [25]. We used the Genome Analysis Tool Kit (GATK version 2.4.9, [26]) to perform local realignment and to produce a cleaned BAM file. The genome data have been made freely available under accession no. PRJEB7527 at the European Nucleotide Archive [27].

Search for variants was then made with the unified genotyper module of GATK. The variant data for each sample was obtained in variant call format (version 4.0) as raw calls for all samples and sites flagged using the variant filtration module of GATK. Variant filtration was performed following best practice documentation of GATK version 4. The snpEFF software [28] together with the UMD3.1/bosTau Ensembl annotation was used to predict the functional effects of detected variants. The Delly package was used to detect larger deletions in cleaned BAM files [29]. Delly uses variation in pair-end reads distance and orientation to find deletions. Structural variation software that are based on coverage and orientation are unable to detect variations larger than the insert size as read mapping software usually requires the library insert size as an argument for aligning within range. Hence, in order to avoid missing large inserts, deletions and false positives all detected variants in the candidate region were also manually inspected by visual control of the BAM file using IGV browser [30].

Sanger sequencing

The *LAMC2* deletion was verified in the case and the available parents by re-sequencing of targeted PCR products using Sanger sequencing technology. PCR primers were designed using PRIMER3 [31]. PCR products were

run on 0.8% agarose gel, 0.5 µg/ml ethidium bromide. PCR products were amplified using flanking primers for the *LAMC2* exon 1 deletion (F) GGCCTATAGAGAGTGGCATGA, (R) CAAATGAAGCCCTTTGAGGA and a second Reverse primer exclusive for the region deleted in the mutants TTCCTTCCCTCACCATCATC with AmpliTaqGold360Mastermix (Life Technologies) and the products directly sequenced using the PCR primers on an ABI 3730 capillary sequencer (Life Technologies) after treatment with exonuclease I (N.E.B.) and rAPid alkaline phosphatase (Roche). Sequence data were analyzed using Sequencher 5.1 (GeneCodes).

Results and discussion

Phenotypes

Lesions in the skin were in principal similar in the two necropsied calves, but more widespread and severe in case 1 than in case 3 – probably reflecting the age difference (4 vs. 0 days of age). In case no. 3, skin lesions were restricted to bilateral absence of horn on the front limb dew claws and an area without horn affecting most of the hind limbs' lateral main digit (Figure 1a). The exposed dermis was hyperaemic but without exudation. The border was sharply demarcated and the adjacent epidermis seemed normal. The few days old case (no. 1) had skin lesions in the distal parts of all limbs. The horn wall was absent on all dew claws exposing a hyperaemic corium covered by crusts (Figure 1b) and the horn on all main digits was defective. The horn wall was totally absent in one hind limb digit and partly absent in the others and the exposed corium showed intense hyperaemia (Figure 1c).

The horn wall was loosened in both front limbs and separated from the corium in the coronet band with suppuration. Fistulas opened either in the coronet band or penetrated the sole. Skin ulcerations covered by crusts stretched proximally from the coronet band to the fetlock region in both hind limbs and were also found above the coronet band in the right front limb, in the left lateral metatarsal area and locally in the ventral part of the trunk.

Mucosal lesions were present in both necropsied cases. Case 3 had a large ulceration of the nasal plate with loosening of epidermis and stretching into the nostrils. In the oral cavity, both calves had extensive ulcerative lesions including a circular ulceration in the tongue around the anterior part of the torus (Figure 1d), and ulcerations in the palate, dental pad and adjacent area of the upper lip, gingiva, and cheeks.

Histopathological changes

Histopathological examination of the skin from the distal limbs of case 3, which died immediately after parturition, revealed an abrupt ulceration that was bordered by a hyperplastic epidermis. The skin proximal to the zone of epidermal hyperplasia was normal and without signs of dermoepidermal separation. The ulcerated area adjacent to the zone of epidermal hyperplasia showed acute mild suppurative inflammation, while more distant areas were dominated by an inflamed granulation tissue. Ulcerated areas were mostly without skin adnexa, i.e. glands and hair, although isolated hairs were rarely seen. Skin lesions of case 1 were dominated by ulcerations with superficial dermal necrosis, debris, and profound

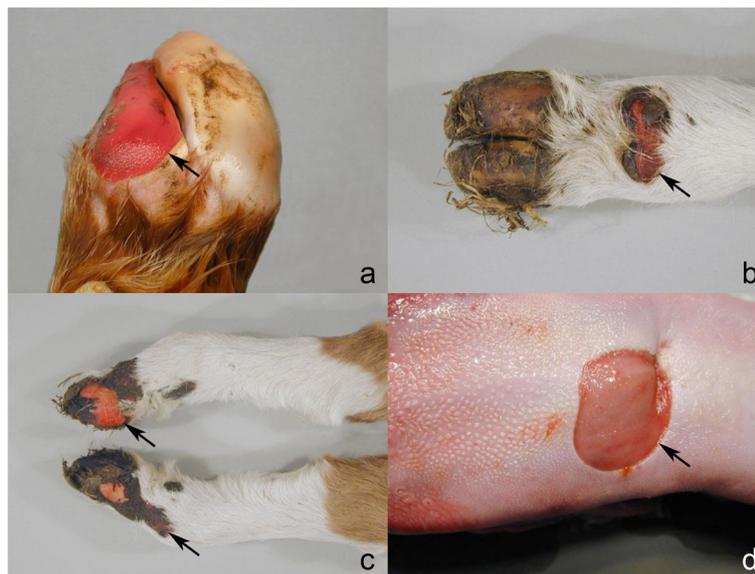


Figure 1 Gross lesions in Hereford calves with epidermolysis bullosa. **a)** Congenital absence of most of the hoof of a main digit (case 3); **b)** Loss of the horn of the dew claws with inflamed corium (case 1); **c)** Absence of part of the lateral aspect of the horn of the hind limb main digits exposing a hyperemic corium (case 1); **d)** Local absence of the lingual epithelium (case 1). The border of lesions is indicated by arrows.

suppurative inflammation. The epidermis adjacent to the ulcerative area was necrotic and separated from the dermis and the dermis had diffuse suppurative inflammation. This zone continued into areas with subepidermal blisters with a purulent content (Figure 2a) and in more distant areas by subepidermal blisters with just a few neutrophils and decreasing degrees of dermal inflammation (Figure 2b). Remnants of skin adnexa were present in the ulcerated areas.

Lesions of the dental pad and upper lip of case 3 were characterised by an abrupt transition from normal epithelium to an ulcer with peripheral acute inflammation and distant granulation tissue formation with sparse inflammation. Beneath ulcerated lesions, profoundly located merocrine sweat glands were present. In case 1, dental pad and nasal plate lesions were characterised by severe fibrinonecrotising and suppurative inflammation of the denuded

dermis and bordered by areas with subepidermal blisters. In the tongue, lesions corresponded to those observed in the dental pad of cases 1 and 3, respectively. PAS staining revealed a basement membrane apparently located at the bottom of some blisters, while a distinct basement membrane was not present in others. An EB type/subtype was not established as appropriate materials were not available. Tissues were autolysed due to prolonged time between the death of the calves and necropsy and cryopreserved specimens for immunofluorescence antigen mapping were not sampled. Typing of EB is severely compromised if optimal specimens for immunofluorescence antigen mapping are not sampled and diagnostic based on formalin fixed tissues and suboptimal TEM examinations may be misleading [1,32] in microscopic typing of EB.

The calves had a severe congenital blistering disorder affecting the skin and mucous membranes, and the claws and dewclaws had either total or partial loss of horn. Histologically, blisters were present in the areas of the skin affected in EB although the precise localization of the spitting plane could not be determined. In combination, these findings are consistent with EB and the cases share many features with established or suspected cases of EB in cattle [6,16-20].

Pedigree analysis and mapping

The close familiar relationship between cases strongly indicated a genetic aetiology; we therefore decided to analyse the pedigree data to infer an inheritance mechanism. Pedigree data were not complete, but the available information regarding ancestors allowed us to draw a genealogical diagram (Figure 3). Analysis of the diagram indicated that inbreeding and the transmission of a recessive mutant allele from a common ancestor; either cow IV/A or Sire IV/B (Figure 3) could be the founder or the distributor of the responsible mutation. These animals had been mated and produced a son (III/A), who was bred to his own mother (IV/A). This inbreeding loop produced case 1; III/A was also bred to his sister (III/B) and produced case 2 and got a son (II/C) with a cow of unknown descent (III/C). The son II/C was mated to his mother (III/B), who gave birth to case 3. III/C and IV/A could share a common ancestor - presence of other common ancestors could not be excluded due to incomplete pedigree data. These data suggested a monogenic recessive inheritance mechanism; therefore, we hypothesized a simple Mendelian recessive inheritance was the most likely explanation for the condition. We initiated a positional cloning study to unravel the underlying genetics. We assumed that the affected calves were expected to be identical by descent (IBD) for the causative mutation and flanking chromosomal segments. We initially genotyped 777,961 evenly spaced SNPs one family trio (case 1 plus its parents). We searched for

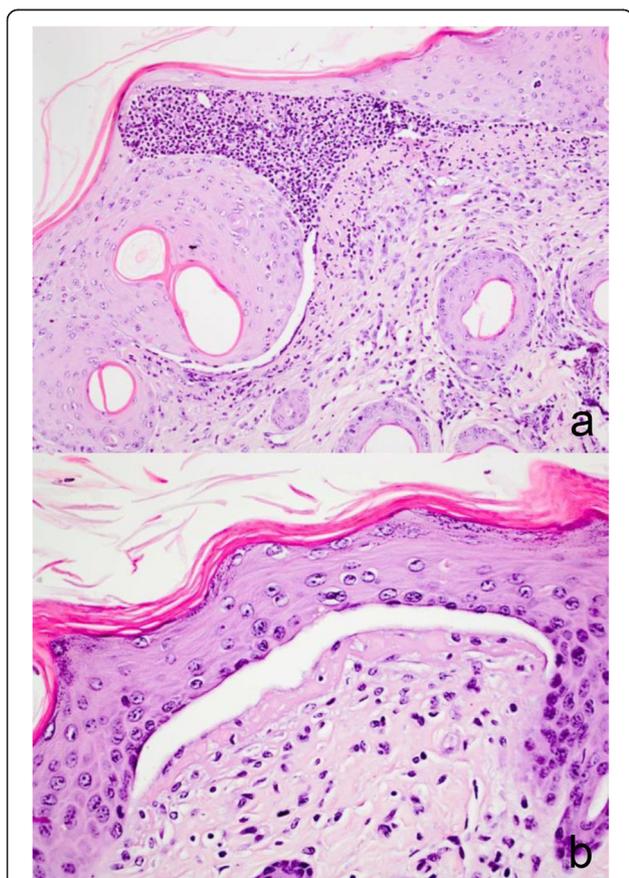
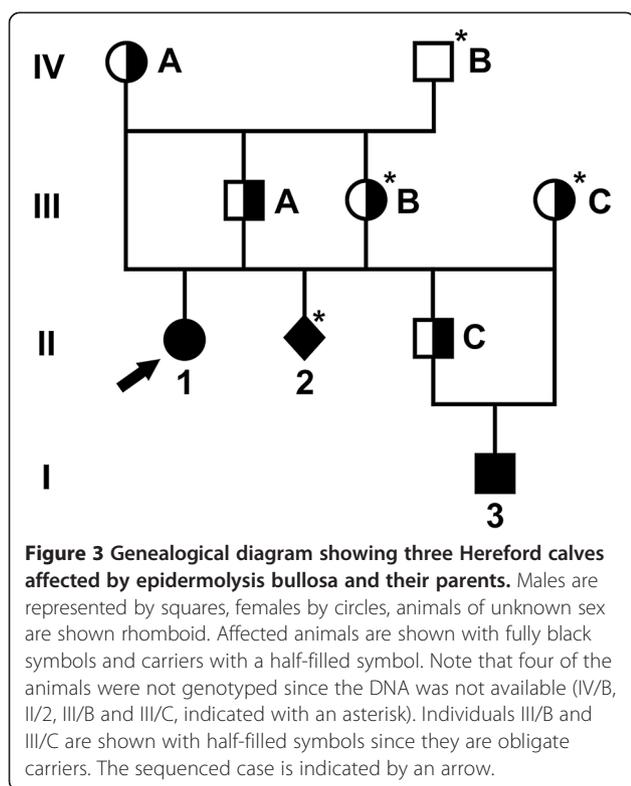


Figure 2 Microscopic lesions in Hereford calves with epidermolysis bullosa. **a)** Subepidermal blister with a purulent content and inflammation of the superficial dermis (case 1, hematoxylin and eosin, obj x 20); **b)** Subepidermal blister with just a few neutrophils, slight suppurative inflammation in the superficial dermis that is also covered by an eosinophilic material. The clear space may be artificial due to autolysis and tissue shrinkage during processing (case 1, hematoxylin and eosin, obj x 40).



extended regions of homozygosity and compared the homozygous region between the case and the parents. Interestingly, in the genotyped affected animal about 37% of the genome is homozygous as expected in a consanguineous son-mother mating (Figure 3). We excluded any homozygous regions already present in the parents. Thereby, we found 40 genome regions greater than 1 Mb that fulfilled these criteria (Additional file 1).

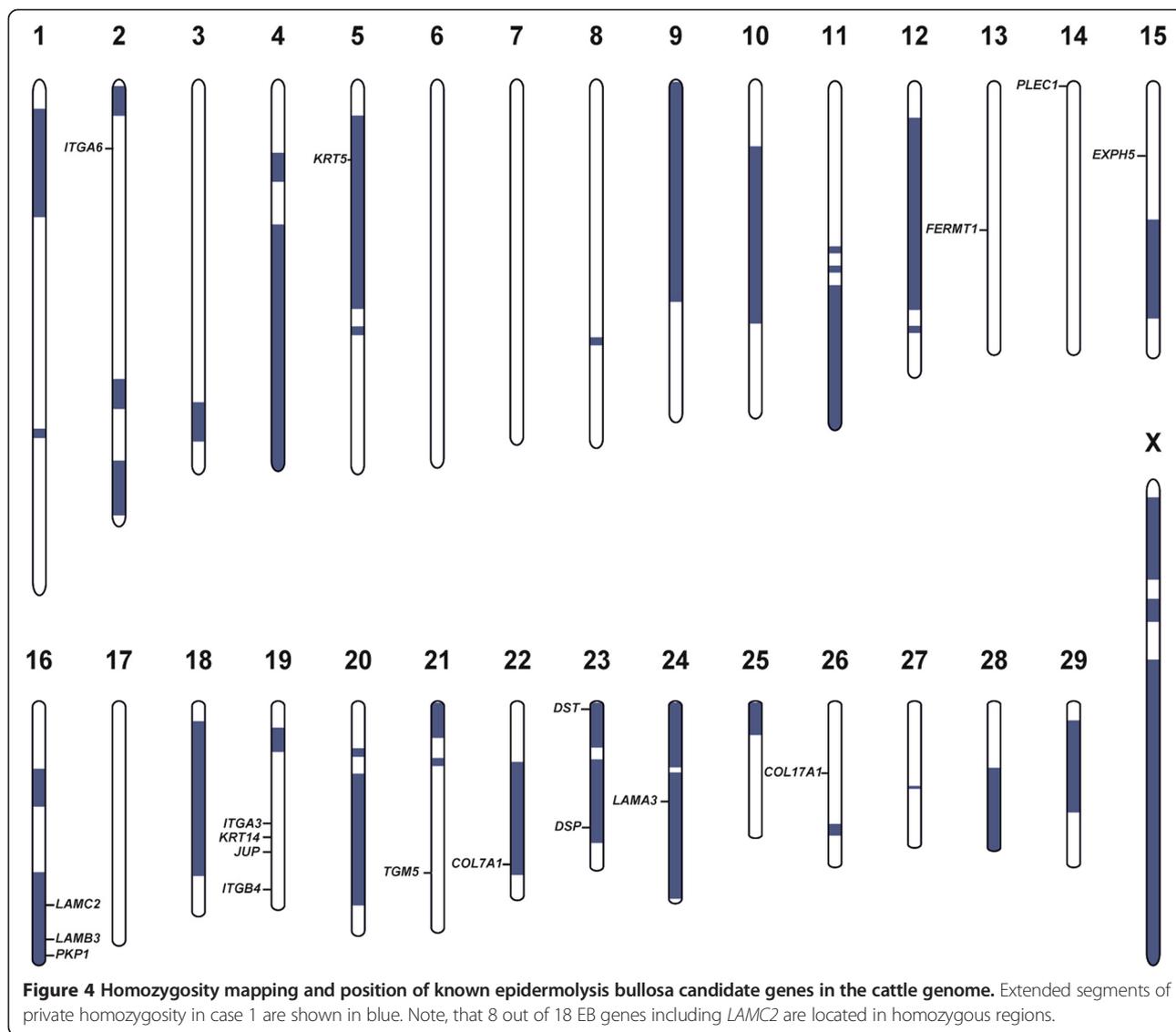
In the light of the few reports of EB causing mutations in livestock we hypothesized that a causative variant might affect one of the known EB candidate genes. Eight out of 18 EB candidates mapped in the identified homozygous regions (Figure 4). We sequenced the whole genome of case 1 at 17.5× coverage of the genome and proceeded to screen the candidate genes present in the mapped homozygous intervals for possible variants. This allowed us to identify 118,014 single nucleotides and short insertion/deletion variants within the whole exome. From this point, we decided to use a candidate gene based approach. We carefully checked for all the variants present in the coding sequence of the 8 remaining EB candidate genes, which were located in the previously identified homozygous candidate regions (Figure 4). We found no homozygous private variant in all EB genes in the sequenced affected animal after comparison with available data of 40 sequenced control cattle genomes (Additional file 2), which were sequenced in the course of other

ongoing projects in our group (variants exclusive of the sequenced animal after this filtering step are reported in Additional file 3).

Using these controls genomes we went on searching for larger deletions and found a total of 349 private deletions occurring only in the genome of the affected calf (Additional file 4). Interestingly, among these deletions the only variant detected overlapping with an annotated coding region found was a 2,433 bp deletion on chromosome 16 (g.65,704,617_65,707,049del) affecting an EB candidate gene. This homozygous deletion encompasses the region more than 900 bp upstream and 1.1 kb downstream of the first exon of the annotated transcript *ENSBTAT00000061289*. This annotated bovine transcript corresponds to the human *LAMC2* gene encoding the laminin gamma 2 protein. The variant causes the complete deletion of the entire *LAMC2* exon 1 containing the start codon and the first 79 coding bases of the transcript (Figure 5). We genotyped the available family members (dam of case 1, IV/A; sire of case 1, III/A; sire of case 3, II/C) and found the mutation present in heterozygosity in these animals (Figure 5). This confirms the assumed recessive inheritance of the EB mutation within this cattle family. The mutant allele was absent in normal controls. A homozygous mutation completely removing the start codon of the evolutionary conserved *LAMC2* wildtype transcript is highly likely disruptive and almost certainly negates completely the presence of a functional *LAMC2* protein. Alternatively, the possible usage of a second start codon located approximately 900 nucleotides further downstream is predicted to lead to a truncated protein lacking about 25% of the wildtype *LAMC2* including three conserved domains. Therefore we speculate that this mutant protein will, if really expressed, probably not compensate the physiological function of the wild type protein. The *LAMC2* loss of function mutation affects a well-known candidate gene associated with junctional EB in humans [33-40] and domestic animals like sheep and horse [8,9]. For this reason, we concluded that the observed EB type in this cattle family was caused by the detected *LAMC2* deletion.

Clinical and molecular characterization of EB

Diagnostic of EB in animals, including classification in major types, has been done in several studies based on histology and TEM. However, routine histological processing of skin is not recommended by human pathologists for EB diagnostic due to the difficulties in distinguishing at the light microscopy level between several types of EB [4]. Furthermore, correct sampling and processing of specimens is crucial to avoid artifactual blistering. Also fresh blisters should be induced by gently rubbing the skin rather than sampling older blisters as this may lead to a wrong diagnosis [41]. TEM examination has been used in

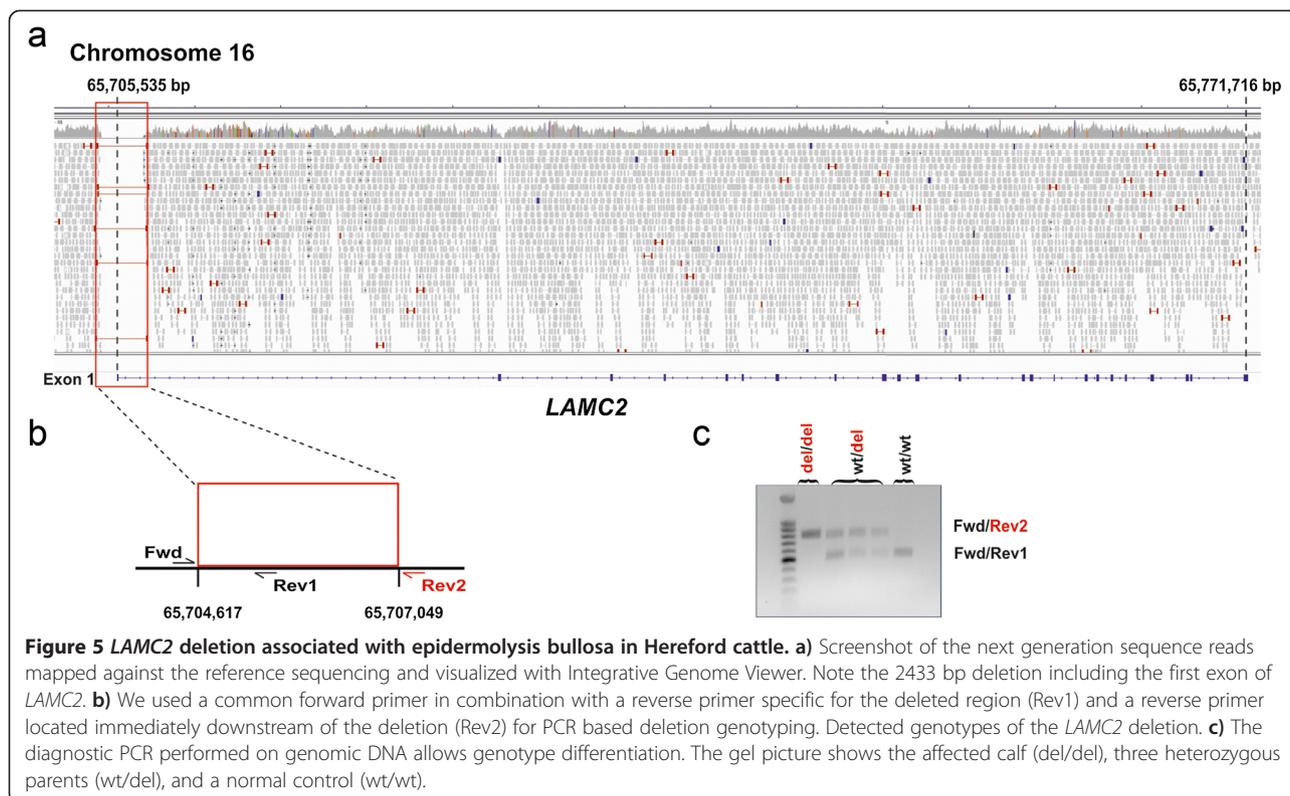


several studies on EB in animals but require samples without autolysis. This may be difficult to achieve as animals often die or are euthanized on the farm by humane grounds before appropriate sampling is possible as for the cases reported here. Furthermore, processing of skin biopsy specimens for TEM and interpretation of EB ultrastructural changes require extensive experience and expertise that is available only in a very few recommended reference laboratories worldwide. Otherwise findings may be quite misleading and should be interpreted with caution [4]. Immunofluorescence mapping for antigens associated with EB in cryopreserved skin specimens taken from fresh spontaneous or traction-force induced blisters is the recommended diagnostic method, especially if samples are shipped to a reference laboratory [4]. The recommendations by experts in EB in

man [4] shows that there are many pitfalls in non-molecular diagnostic of EB that are obstacles in veterinary diagnostic, especially due to suboptimal materials and lack of experience with the highly specialised EB diagnostic.

Congenital localised absence of skin (CLAS) and mucosal epithelium was apparent in the calf that died immediately after parturition (case 3). CLAS has been reported as a manifestation of EB in man, e.g. Bart's Syndrome [42-44] and although CLAS and mucosal epithelium defects may be due inadequate development, CLAS in association with EB merely reflects intrauterine loss of tissue due to foetal development of EB [43].

Congenital blistering disorders in the mucous membranes, muzzle and skin, especially the distal part of the limbs, have been reported in cattle through decades and



usually referred to as “epitheliogenesis imperfecta” (EI) as proposed by Hadley [45]. Many of these cases have occurred in familial patterns associated with inbreeding and likely transmission of an autosomal recessive mutant allele from a founder animal [44-46] and many cattle breeds have been affected [15,45-51]. However, EI probably comprises of two distinct entities, namely congenital cutaneous aplasia (CCA) and EB and taking the current knowledge on the genetic background and variation even within subtypes of EB in man into consideration, the reported cases are likely to have different etiopathogeneses. Investigation of CCA has demonstrated that this is not a blistering disease and it is striking that cases resembling EI (except CCA cases) have turned out to be EB if investigated to the molecular level [5,6,9,10,52]. Furthermore, it is worth remembering that EI was introduced to veterinary medicine based on gross lesions only [44]. It is therefore proposed that EI is no longer used as a disease entity but replaced by either CCA or EB as also suggested by others [6,53].

EB is a rare disease known in man and many animal species. However, in livestock subtypes with recessive inheritance have the potential to become of significance if the defective allele is present in important sires used for artificial breeding through several years as seem for other genetic diseases in cattle [54-56]. It is therefore

important that cases are diagnosed to subtype level when the first cases are recognised to enable rapid development of genetic test that allows screening of the sire population. However, subtyping of EB is a task for clinicians and pathologists, but it starts with clinical examination and sampling and processing of adequate materials for microscopic and genomic analysis [41]. The quick identification of an highly disruptive mutation in a known candidate gene allowed us to unambiguously identify and classify the EB cases in Hereford and thus (I) allow for possible screening of the mutation in the Danish Hereford population as preventive measure (II) giving a new animal model for EB caused by a mutation in laminin gamma genes.

Conclusions

The study reports for the first time the occurrence of EB in Hereford cattle. Our investigation confirms the role of a recessively inherited *LAMC2* loss of function mutation in EB aetiology. Next-generation sequencing offers a powerful tool for understanding the genetic background of rare diseases in domestic animals with an available reference genome sequence. Verifying the causative mutation allowed us to confirm to the diagnosis of EB and allows breeders to eliminate this genetic defect from the population.

Additional files

Additional file 1: Private regions of homozygosity detected in epidermolysis bullosa case 1. Only homozygous regions exclusive for the case are shown.

Additional file 2: List of all the control cattle genomes and their breed.

Additional file 3: List of all the private homozygous variants of the sequenced epidermolysis bullosa case 1.

Additional file 4: Private larger deletions detected in epidermolysis bullosa case 1. Deletion in encompassing *LAMC2* exon 1 is highlighted.

Abbreviations

Bp: Base pairs; CCA: Congenital cutaneous aplasia; CLAS: Congenital localised absence of skin; EB: Epidermolysis bullosa; El: Epitheliogenesis imperfect; kb: Kilo bytes; PAS: Periodic acid-Schiff; PCR: Polymerase chain reaction; SNP: Single nucleotide polymorphism; TEM: Transmission electron microscopy.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

JSA necropsied case 1, evaluated pedigree information, did the histopathological analyses and drafted the manuscript. LKI necropsied case 3. LM and NW performed the genetic investigations and drafted the manuscript. VJ analysed the whole genome sequencing data. CD supervised the genetic project and edited the manuscript. All authors participated in discussion of results and critically revised the manuscript. All authors approved the final version of the manuscript.

Acknowledgements

The authors acknowledge Mr Ole R Kjærulf, Roskilde, Denmark for submission of the cases. The study was funded by the Danish Surveillance Programme for Genetic Diseases in Cattle. The authors wish to thank Michèle Ackermann and Muriel Fragnière for their precious technical assistance. We would like to express our appreciation to the University of Bern for the use of the Next Generation Sequencing Platform in performing the whole genome re-sequencing experiment and the Vital-IT high-performance computing center of the Swiss Institute of Bioinformatics for performing computationally intensive tasks (<http://www.vital-it.ch/>).

Author details

¹Institute of Genetics, Vetsuisse Faculty, University of Bern, Bremgartenstrasse 109a, CH-3001 Bern, Switzerland. ²Department of Veterinary Disease Biology, Section for Veterinary Pathology, Faculty of Health and Medical Sciences, University of Copenhagen, Ridebanevej 3, DK-1870 Frederiksberg C, Denmark. ³Department of Large Animal Sciences, Section for Veterinary Reproduction and Obstetrics, Faculty of Health and Medical Sciences, University of Copenhagen, Dyrhavevej 68, DK-1870 Frederiksberg C, Denmark.

Received: 3 November 2014 Accepted: 22 January 2015

Published online: 07 February 2015

References

1. Fine JD. Inherited epidermolysis bullosa: recent basic and clinical advances. *Curr Opin Pediatr*. 2010;22:453–8.
2. Ginn PE, Mansell JEKL, Rakich PM. Skin and appendages. In: Maxie MG, editor. *Jubb, Kennedy and Palmer's pathology of domestic animals*, vol. 1. 5th ed. Edinburgh: Saunders-Elsevier; 2007.
3. Fine JD, Bruckner-Tuderman L, Eady RA, Bauer EA, Bauer JW, Has C, et al. Inherited epidermolysis bullosa: updated recommendations on diagnosis and classification. *J Am Acad Dermatol*. 2014;70:1103–26.
4. Has C, Bruckner-Tuderman L. The genetics of skin fragility. *Annu Rev Genomics Hum Genet*. 2014;15:245–68.
5. Ford CA, Stanfield AM, Spelman RJ, Smits B, Ankersmidt-Udy AE, Cottier K, et al. A mutation in bovine keratin 5 causing epidermolysis bullosa simplex, transmitted by a mosaic sire. *J Invest Dermatol*. 2005;124:1170–6.
6. Menoud A, Welle M, Tetens J, Lichtner P, Drögemüller C. A COL7A1 mutation causes dystrophic epidermolysis bullosa in Rotes Höhenvieh cattle. *PLoS One*. 2012;7:e38823.
7. Baldeschi C, Gache Y, Rattenholl A, Bouillé P, Danos O, Ortonne JP, et al. Genetic correction of canine dystrophic epidermolysis bullosa mediated by retroviral vectors. *Hum Mol Genet*. 2003;12:1897–905.
8. Mömke S, Kerkmann A, Wöhlke A, Ostmeier M, Hewicker-Trautwein M, Ganter M, et al. A frameshift mutation within *LAMC2* is responsible for Herlitz type junctional epidermolysis bullosa (HJEB) in black headed mutton sheep. *PLoS One*. 2011;6:e18943.
9. Milenkovic D, Chaffaux S, Taourit S, Guérin G. A mutation in the *LAMC2* gene causes the Herlitz junctional epidermolysis bullosa (H-JEB) in two French draft horse breeds. *Genet Sel Evol*. 2003;35:249–56.
10. Graves KT, Henney PJ, Ennis RB. Partial deletion of the *LAMA3* gene is responsible for hereditary junctional epidermolysis bullosa in the American Saddlebred Horse. *Anim Genet*. 2009;40:35–41.
11. Spirito F, Charlesworth A, Linder K, Ortonne JP, Baird J, Meneguzzi G. Animal models for skin blistering conditions: absence of laminin 5 causes hereditary junctional mechanobullous disease in the Belgian horse. *J Invest Dermatol*. 2002;119:684–91.
12. Capt A, Spirito F, Guaguere E, Spadafora A, Ortonne JP, Meneguzzi G. Inherited junctional epidermolysis bullosa in the German Pointer: establishment of a large animal model. *J Invest Dermatol*. 2005;124:530–5.
13. Bruckner-Tuderman L, McGrath JA, Robinson EC, Uitto J. Animal models of epidermolysis bullosa: update 2010. *J Invest Dermatol*. 2010;130:1485–8.
14. Deprez P, Maenhout T, De Cock H, Wullepit J, Charlier G, Muylle E, et al. Epidermolysis bullosa in a calf: a case report [in Dutch]. *Vlaams Diergeneeskd Tijdschr*. 1993;62:155–9.
15. Agerholm JS. Congenital generalized epidermolysis bullosa in a calf. *Zentralbl Veterinarmed A*. 1994;41:139–42.
16. Stocker H, Lott G, Straumann U, Rüschi P. Epidermolysis bullosa in a calf [in German]. *Tierarztl Prax*. 1995;23:123–6.
17. Medeiros GX, Franklin Riet-Correa F, Armien AG, Dantas AFM, de Galiza GJN, Simões SVD. Junctional epidermolysis bullosa in a calf. *J Vet Diagn Invest*. 2012;24:231–4.
18. Thompson KG, Crandell RA, Rugeley WW, Sutherland RJ. A mechanobullous disease with sub-basilar separation in Brangus calves. *Vet Pathol*. 1985;22:283–5.
19. Bassett H. A congenital bovine epidermolysis resembling epidermolysis bullosa simplex of man. *Vet Rec*. 1987;121:8–11.
20. Foster AP, Skuse AM, Higgins RJ, Barrett DC, Philbey AW, Thomson JR, et al. Epidermolysis bullosa in calves in the United Kingdom. *J Comp Pathol*. 2010;142:336–40.
21. Purcell S1, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, et al. PLINK: a toolset for whole-genome association and population-based linkage analysis. *Am J Hum Genet*. 2007;81:559–75.
22. Murgiano L, Jagannathan V, Benazzi C, Bolcato M, Brunetti B, Muscatello LV, et al. Deletion in the *EVC2* gene causes chondrodysplastic dwarfism in Tyrolean Grey Cattle. *PLoS One*. 2014;9:e94861.
23. Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics*. 2009;25:1754–60.
24. Homepage Samtools. [<http://samtools.sourceforge.net>]
25. Picard. [<http://sourceforge.net/projects/picard/>]
26. McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis K, Kernysky A, et al. The genome analysis toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res*. 2010;20:1297–303.
27. Homepage European Nucleotide Archive. [<http://www.ebi.ac.uk/ena/data/view/PRJEB7527>. Accessed 14 Juli 2014].
28. Cingolani P, Platts A, Wang L, Coon M, Nguyen T, Wang L, et al. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of *Drosophila melanogaster* strain w1118; iso-2; iso-3. *Fly*. 2012;6:80–92.
29. Rausch T, Zichner T, Schlattl A, Stütz AM, Benes V, Korbel JO. DELLY: structural variant discovery by integrated paired-end and split-read analysis. *Bioinformatics*. 2012;28:i333–9.
30. Thorvaldsdóttir H, Robinson JT, Mesirov JP. Integrative Genomics Viewer (IGV): high-performance genomics data visualization and exploration. *Brief Bioinform*. 2013;14:178–92.
31. Untergrasser A, Cutcutache I, Koressaar T, Ye J, Faircloth BC, Remm M, et al. Primer3 - new capabilities and interfaces. *Nucleic Acids Res*. 2012;40:e115.

32. Ostmeier M, Kerkmann A, Frase R, Ganter M, Distl O, Hewicker-Trautwein M. Inherited junctional epidermolysis bullosa (Herlitz type) in German black-headed mutton sheep. *J Comp Pathol.* 2012;146:338–47.
33. Aumailley M, Bruckner-Tuderman L, Carter WG, Deutzmann R, Edgar D, Ekblom P, et al. A simplified laminin nomenclature. *Matrix Biol.* 2005;24:326–32.
34. Castiglia D, Posteraro P, Spirito F, Pinola M, Angelo C, Puddu P, et al. Novel mutations in the LAMC2 gene in non-Herlitz junctional epidermolysis bullosa: effects on laminin-5 assembly, secretion, and deposition. *J Invest Dermatol.* 2001;117:731–9.
35. Hartwig B, Borm B, Schneider H, Arin MJ, Kirfel G, Herzog V. Laminin-5-deficient human keratinocytes: defective adhesion results in a saltatory and inefficient mode of migration. *Exp Cell Res.* 2007;313:1575–87.
36. Mühle C, Jiang QJ, Charlesworth A, Bruckner-Tuderman L, Meneguzzi G, Schneider H. Novel and recurrent mutations in the laminin-5 genes causing lethal junctional epidermolysis bullosa: molecular basis and clinical course of Herlitz disease. *Hum Genet.* 2005;116:33–42.
37. Nakano A, Chao SC, Pulkkinen L, Murrell D, Bruckner-Tuderman L, Pfindner E, et al. Laminin 5 mutations in junctional epidermolysis bullosa: molecular basis of Herlitz vs. non-Herlitz phenotypes. *Hum Genet.* 2002;110:41–51.
38. Pulkkinen L, Uitto J. Mutation analysis and molecular genetics of epidermolysis bullosa. *Matrix Biol.* 1999;18:29–42.
39. Schneider H, Mühle C, Pacho F. Biological function of laminin-5 and pathogenic impact of its deficiency. *Eur J Cell Biol.* 2007;86:701–17.
40. Varki R, Sadowski S, Pfindner E, Uitto J. Epidermolysis bullosa. I. Molecular genetics of the junctional and hemidesmosomal variants. *J Med Genet.* 2006;43:641–52.
41. Intong LR, Murrell DF. How to take skin biopsies for epidermolysis bullosa. *Dermatol Clin.* 2010;28:197–200. vii.
42. Butler DF, Berger TG, James WD, Smith TL, Stanely JR, Rodman OG. Bart's syndrome: microscopic, ultrastructural, and immunofluorescent mapping features. *Pediatr Dermatol.* 1986;3:113–8.
43. Kanzler MH, Smoller B, Woodley DT. Congenital localized absence of the skin as a manifestation of epidermolysis bullosa. *Arch Dermatol.* 1992;128:1087–90.
44. Bart BJ, Lussky RC. Bart syndrome with associated anomalies. *Am J Perinatol.* 2005;22:365–9.
45. Hadley FB. Congenital epithelial defects of calves. *J Hered.* 1927;18:487–95.
46. Leipold HW, Mills JH, Huston K. Epitheliogenesis imperfecta in Holstein-Friesian calves. *Can Vet J.* 1973;14:114–8.
47. Dyrendahl S. Epitheliogenesis imperfecta in the Swedish Red and White breed [in Swedish]. *Nord Vet Med.* 1956;8:953–8.
48. Straub OC. Epitheliogenesis imperfecta in a calf [in German]. *Vet Med Nachricht.* 1969;3:189–93.
49. Yeruhama I, Goshen T, Lahav D, Perl S. Simultaneous occurrence of epitheliogenesis imperfecta with syndactyly in a calf and a lamb. *Aust Vet J.* 2005;83:149–50.
50. Jayasekara MU, Leipold HW. Epitheliogenesis imperfecta imperfecta in Shorthorn and Angus cattle. *Zentralbl Veterinarmed A.* 1979;26:497–501.
51. Hutt FB, Frost JN. Hereditary epithelial defects in Ayrshire cattle. *J Hered.* 1948;39:131–7.
52. Lieto LD, Swerczek TW, Cothran EG. Equine epitheliogenesis imperfecta in two american saddlebred foals is a lamina lucida defect. *Vet Pathol.* 2002;39:576–80.
53. Benoit-Biancamano MO, Drolet R, D'Allaire S. Aplasia cutis congenita (epitheliogenesis imperfecta) in swine: observations from a large breeding herd. *J Vet Diagn Invest.* 2006;18:573–9.
54. Agerholm JS. Inherited disorders in Danish cattle. *APMIS.* 2007;115 Suppl 122:1–76.
55. Murgiano L, Testoni S, Drögemüller C, Bolcato M, Gentile A. Frequency of bovine congenital pseudomyotonia carriers in selected Italian Chianina sires. *Vet J.* 2013;195:238–40.
56. Murgiano L, Drögemüller C, Sbarra F, Bolcato M, Gentile A. Prevalence of paunch calf syndrome carriers in Italian Romagnola cattle. *Vet J.* 2014;200:459–61.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

