

RESEARCH ARTICLE

Open Access

Proteomic analysis of the extraembryonic tissues from cloned porcine fetus at day 35 of pregnancy

Yeoung-Gyu Ko^{1*}, Seongsoo Hwang¹, Sung Woo Kim¹, Hyun Kim¹, Hwan Hoo Seong¹, Jae-Hwan Kim¹, Yuno Song², Bo-Suck Yang³, Young Min Song³ and Jae-Hyeon Cho^{2*}

Abstract

Background: Somatic cell cloning by nuclear transfer (SCNT) in pig is clearly of great benefit for basic research and biomedical applications. Even though cloned offspring have been successfully produced in pig, SCNT is struggling with the low efficiency.

Results: In the present study, we investigated differentially expressed proteins of the extraembryonic tissue from pig SCNT fetus compared to control (normal) fetus. We obtained the extraembryonic tissue from embryos at day 35 of pregnancy and examined the protein expression profiles using two-dimensional electrophoresis (2-D) and Western blotting. The extraembryonic tissue of fetus in control pregnancy was compared to the extraembryonic tissue of SCNT fetus, which showed an abnormally small size and shape as well as exhibited abnormal placental morphology compared to control fetus. A proteomic analysis showed that the expression of 33 proteins was significantly increased or decreased in the extraembryonic tissue of SCNT fetus compared to control fetus. The differentially expressed proteins in the extraembryonic tissue of SCNT fetus included ATP or lipid binding proteins, antioxidant proteins, translation elongation factors, and transcription factors. Western blotting analysis indicated that antioxidant enzymes and anti-apoptotic proteins were down-regulated; however, the expression levels of apoptotic proteins, Bax and Hsp27, were increased in the extraembryonic tissue of SCNT fetus. Moreover, immunohistochemical analysis also showed that the expression of the catalase or GPX genes was decreased in the extraembryonic tissue with SCNT fetus compared to those with control fetus. In addition, we observed a significant decrease in DNA methyltransferase1 (Dnmt1) expression in SCNT extraembryonic tissue, and the expression levels of Dnmt3a and Dnmt3b were abnormally higher in SCNT fetus compared to control fetus. Moreover, a marked increase in the frequency of TUNEL-positive cells was observed in the extraembryonic tissue in SCNT fetus.

Conclusion: These results demonstrated that pig SCNT fetus showed abnormal protein expression in the extraembryonic tissue, and extensive apoptosis occurred in the extraembryonic tissue of the SCNT fetus due to an increase in apoptotic protein expression or a decrease in antioxidant protein expression.

Keywords: Extraembryonic tissue, Placenta, SCNT, Apoptosis, Gene expression

* Correspondence: kog4556@korea.kr; jaehcho@gnu.ac.kr

¹Animal Genetic Resources Station, National Institute of Animal Science, RDA, Namwon 590-832, Korea

²Institute of Agriculture and Life Science, College of Veterinary Medicine, Gyeongsang National University, Jiju Daero 501, Jinju, Gyeongsangnamdo 660-701, Korea

Full list of author information is available at the end of the article

Background

The success of somatic cell nuclear transfer (SCNT) in pigs is promising for a wide range of applications, such as genetically superior pig breed production, species resource preservation and xenotransplantation in humans [1,2]. The birth of cloned pigs is achieved by the nuclear transfer of the nuclei of somatic cells [3,4]. However, early embryonic mortality in SCNT embryos pregnant pigs is high during the first 30 days of pregnancy. The high rates of embryonic death of SCNT embryos are thought to be a result of inadequate nuclear reprogramming and extraembryonic tissue formation defects in the cloned embryo. Therefore, the majority of failed pregnancies appears to result from abnormal extraembryonic tissue development, such as reduced vascularization and enlarged placentomes [5-7].

In the genome of SCNT embryos, abnormal methylation patterns were detected in cloned embryos compared to embryos derived by in vitro fertilization [8]. Trophectoderm-localized methylation aberrancy was observed and is related to placental dysfunction closely observed in cloned animals [9]. The abnormal epigenetic reprogramming of trophoblasts in SCNT embryos suggests that the extraembryonic tissue of trophoblasts develops abnormally. Aberrant methylation in the trophectoderm of cloned blastocysts induces global gene dysregulation in extraembryonic tissues, and this type of gene dysregulation can potentially lead to the development of a dysfunctional placenta and have detrimental effects on fetal development [10,11].

Apoptosis is an important process during animal development and reproduction. During pregnancy, apoptosis is physiologically important for normal placental growth and development [12]. Apoptosis is triggered by many different cellular stimuli, including oxidative stress, oxidative damage and cytotoxicity, which induce the activation of the caspase-cascade signaling system. Oxidative stress contributes to ROS formation in SCNT fetus, and ROS levels gradually increase during early pregnancy, resulting in apoptotic cell death in SCNT fetus [13]. Koo and colleagues demonstrated that oxidative stress is a major cause of apoptosis in SCNT extraembryonic tissue and that the low birth rate of cloned animals is due to abnormal apoptosis in the extraembryonic tissue during early pregnancy [14].

In the present study, we investigated differences in gene expression profiles of the extraembryonic tissue of

SCNT and control fetus at early pregnancy (gestational day 35) using two-dimensional electrophoresis (2-D) analysis. In the proteomic analysis, 33 proteins were identified as being differentially regulated in the SCNT extraembryonic tissues. Among these proteins, 3 proteins were up-regulated, whereas the other 30 proteins were down-regulated. Proteomic analysis showed that the expression of apoptosis-related proteins, Bax and Hsp27 were expressed significantly higher in SCNT than that of the normal extraembryonic tissue, whereas the majority of the ATP binding proteins, antioxidant proteins, and metabolic-related proteins were down-regulated in SCNT extraembryonic tissues. The results of this study showed that the expression profile of the genes in extraembryonic tissue of SCNT fetus had increased apoptosis-related proteins and decreased oxidative stress proteins, causing abnormal placenta and fetal development and increased embryonic loss during early pregnancy.

Results

Morphology of the fetus and extraembryonic tissue

A total of 1500 nuclear transferred embryos at the 1- to 2-cell stage were surgically transferred to the oviducts of 18 recipients. Eight recipients (8 pregnancy/18 recipient) were confirmed to be pregnant at day 35 after the transfer of the embryo. As shown in Table 1, the number of recovered SCNT-derived fetus (44.4% pregnancy) at gestational day 35 is significantly lower than the number of fetus recovered from the control (100% pregnancy) (Table 1). The control conceptuses (normal) were obtained from recipients that were impregnated by natural mating. The size and shape of fetus and extraembryonic tissues were observed in the microscopic analysis. SCNT extraembryonic tissue from day 35 of pregnancy was smaller in size and more abnormal in appearance compared to the control extraembryonic tissue (Figure 1). Abnormal blood vessel development was observed in the extraembryonic tissue of SCNT fetus. Interestingly, the sizes of the allantoic and the amniotic sac of both SCNT fetus and control fetus were similar. The fetal weight of the SCNT groups (2.71 ± 0.4) was significantly smaller than the weight of the control groups (3.34 ± 0.26) (Table 1). The number of recovered fetuses from the control group (11.33 ± 1.33) was higher than the number recovered from the SCNT group (4.67 ± 2.33) (Table 1). The number of abnormal fetus with SCNT group (2.33 ± 0.33) was significantly higher than the control

Table 1 Fetal body weight and the number of fetuses recovered at day 35 of pregnancy

Animal group	Recipients	Pregnancy (%)	No. fetuses recovered		Body weight (g)
			Normal	Abnormal	
Control	10	100	11.33 ± 1.33	1 ± 0	3.34 ± 0.26
SCNT	18	44.4	4.67 ± 2.33	2.33 ± 0.33	2.71 ± 0.4

Control, natural mating; SCNT, SCNT fetus. The number mean of normally recovered fetuses from the one SCNT-recipient gilt was 4.67 ± 2.33 . The ratios of control fetal weight were significantly higher than those of the SCNT fetus. Data were expressed as mean \pm SD.

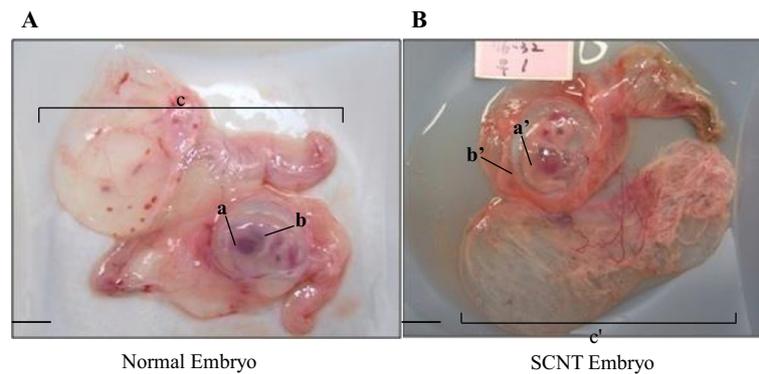


Figure 1 Morphology of fetus and extraembryonic tissues in the control and SCNT-derived fetus from 35 day of pregnancy. (A) Control fetus and extraembryonic tissue from 35 day of pregnancy. **(B)** The SCNT fetus and extraembryonic tissue from 35 day of pregnancy. a and a', embryo; b and b', amniotic sac; c and c' extraembryonic tissue. The weight of fetus was measured after removal of all extraembryonic tissues including chorioallantois, amniotic and allantoic fluid, and umbilical cord. Bar, 10 mm.

group (1 ± 0) (Table 1). Together, these results demonstrated that the extraembryonic tissue in the SCNT fetus showed an abnormally small size and shape, and displayed abnormal morphology of the extraembryonic tissue compared with control (natural mating).

2-D gel analysis of the extraembryonic tissue from control and SCNT fetus

To examine protein expression differences between control and SCNT fetus, we performed proteomics analysis using the extraembryonic tissue of SCNT and control

fetus on day 35 of pregnancy. Figure 2 shows the profile of extraembryonic proteins from SCNT and control fetus using 2-D with a nonlinear IPG gradient of pH 4–10 and 10 ~ 16% gradient gels in the second dimension. A representative 2-D image of proteins in the control and SCNT extraembryonic tissue is shown in Figure 2, and 33 spots were identified as shown in Tables 2 and 3. After protein identification with mass spectrophotometry and protein database search, 3 proteins were up-regulated (2.0- to 4.5-fold increases) and 30 proteins were down-regulated (2.0- to 5.5-fold decreases) in the SCNT extraembryonic

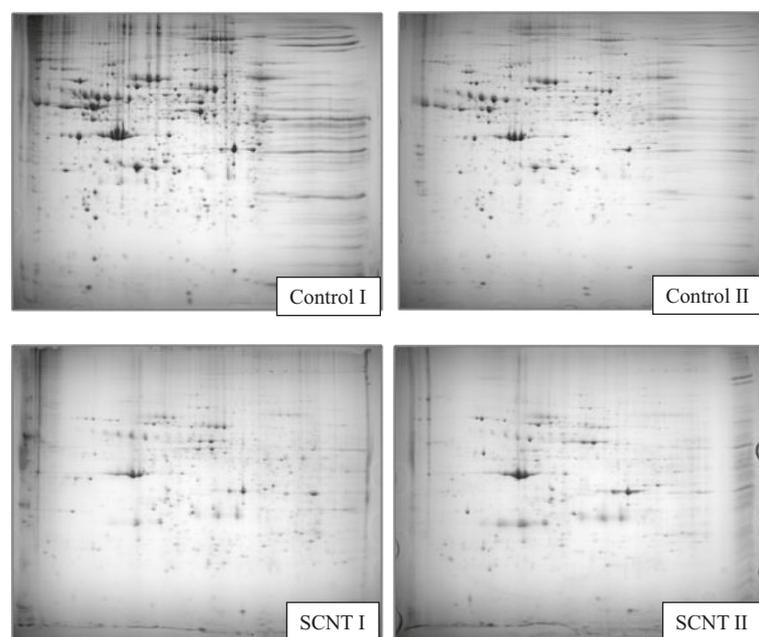


Figure 2 Two-dimensional protein separation of the control and SCNT extraembryonic tissue as visualized by the silver staining. Proteins were isolated from the control and SCNT extraembryonic tissue, and 1000 μ g of total protein were loaded to the 2-D gel. The first dimension was 24-cm pH 4–10 nonlinear IPG, and the second dimension was 10 ~ 16% gradient gels (Control I and II, control extraembryonic tissues; SCNT I and II, SCNT extraembryonic tissues).

Table 2 Up-regulated proteins in the SCNT extraembryonic tissue

Spot no.	Protein name	NCBI accession no.	Species	PMF (MS)			Experimental		Ideal	
				Matched peptides	Measured peptides	Sequence coverage (%)	Molecular mass (kDa)	pI	Molecular mass (kDa)	pI
2401	peroxisome proliferative activated receptor gamma	AAN15206.1	Ovis aries	8	16	23	54.95	6.2	46.63	4.60
5110	heat shock 27 kDa protein 1	NP_001007519.1	Sus scrofa	11	13	43	22.98	6.2	28.02	5.80
5118	Elongation factor 1-gamma (EF-1-gamma) (eEF-1B gamma)	spQ29387	Sus scrofa	7	8	18	49.95	6.2	13.90	5.72

After protein identification with mass spectrophotometry and protein database search, 3 proteins were up-regulated (2.0-to 4.5-fold increases) in the SCNT extraembryonic tissue.

tissue (Tables 2 and 3). Tables 2 and 3 summarizes the primary properties of the 33 proteins that were differentially regulated in SCNT and control fetus. The proteins are classified into six groups based on function: (1) lipid binding proteins (2/33, 6%), (2) antioxidant proteins (3/33, 9%), (3) translation elongation factors (3/33, 9%), (4) ATP binding proteins (6/33, 18%), (5) transcription factors (2/36, 6%), and (6) others (15/36, 51%). The majority of the ATP binding proteins, antioxidant proteins, and metabolic-related proteins were down-regulated, whereas the expression levels of peroxisome proliferative activated receptor gamma (PPAR γ), Hsp27, and elongation factor 1-gamma (EF1- γ) were up-regulated in SCNT extraembryonic tissue.

Immunohistochemical analysis in control and SCNT extraembryonic tissues

To determine the effect of the abnormal development of extraembryonic tissue in SCNT fetus, we performed immunohistochemical analysis of control and SCNT extraembryonic tissue. As shown in Figure 3, histological analysis of SCNT extraembryonic tissues showed clusters of terminal villi with apparent syncytial bridging and sprouting. Adhesion occurred between the maternal and fetal epithelium with the formation of microvilli resulting in an interdigitation of both tissues. The expression of antioxidant proteins GPX and catalase revealed extensive positive staining in the extraembryonic tissues of control fetus; however, SCNT-derived extraembryonic tissue exhibited significantly lower expression levels of GPX and catalase (Figure 3A and B).

Western blot analysis in control and SCNT extraembryonic tissues

To further investigate whether the developmental defects in the extraembryonic tissue of SCNT fetus affect the expression of antioxidation-related proteins or apoptotic marker proteins, such as GPX, catalase, Bax, Bcl2, and Hsp27, we performed western blot analysis using control and SCNT extraembryonic tissue. The expression levels of catalase or GPX related to oxidative stress were decreased in SCNT extraembryonic tissues (Figure 3C). The expression of Hsp27 was up regulated more than two-fold in

SCNT extraembryonic tissue compared with control tissue, and these data are consistent with 2-D analysis. The expression of Bax, an apoptotic marker protein, was up-regulated, and the expression of the anti-apoptotic Bcl-2 gene decreased in the SCNT extraembryonic tissue (Figure 4A). Next, we investigated the expression of DNA methyltransferases 1, 3a and 3b (Dnmt 1, 3a and 3b) in control and SCNT extraembryonic tissue. We found that that Dnmt1 was highly expressed in the extraembryonic tissue of control fetus, whereas the activation of the protein is restricted in the SCNT extraembryonic tissue (Figure 4B). In contrast, the expression levels of Dnmt3a and 3b were highly detectable in the SCNT extraembryonic tissue, while they were low in control fetus (Figure 4B). These results suggest that a relatively low level of Dnmt1 activity exists during the development of SCNT fetus.

TUNEL analysis of control and SCNT extraembryonic tissue

To determine whether the high expression of apoptotic or antioxidant proteins is due to increased apoptosis in SCNT-derived extraembryonic tissue, the TUNEL assay was performed with the extraembryonic tissues from control and SCNT fetus. A small portion of the TUNEL-positive cells was observed in the extraembryonic tissue of control fetus (Figure 4C); however, marked apoptosis was observed in SCNT extraembryonic tissue (Figure 4C). The apoptotic cells in the SCNT extraembryonic tissue were trophoblast cells, which play an important role during pregnancy in the growth and development of the placenta. These results demonstrate that inefficient development of the extraembryonic tissue in SCNT fetus caused abnormal levels of apoptosis in SCNT-derived extraembryonic tissue.

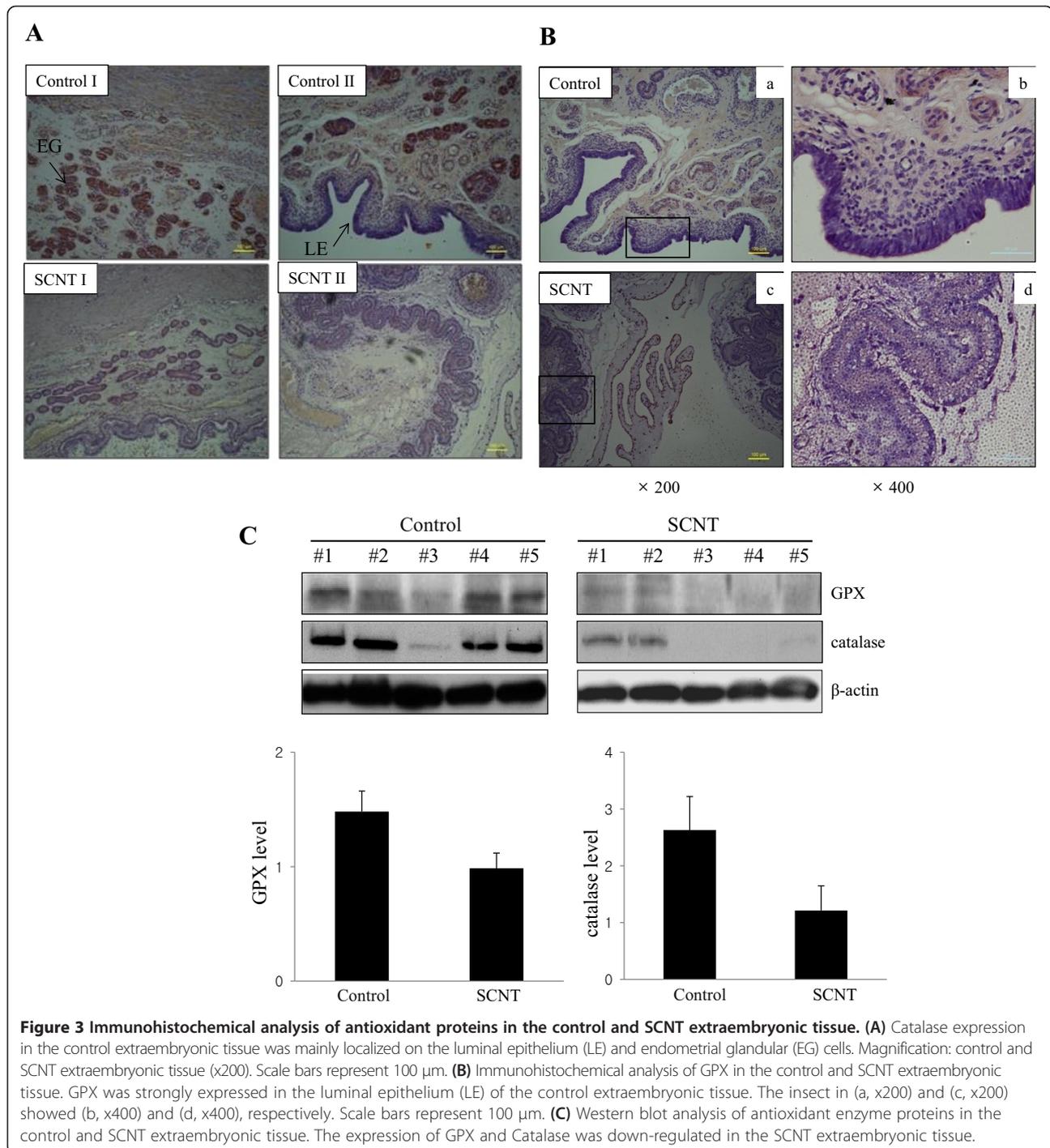
Discussion

In the present study, we investigated differences in the protein expression profiles between control and SCNT extraembryonic tissues at early pregnancy at gestational day 35. Our data showed that proteomics analysis of the extraembryonic tissue from SCNT fetus revealed that 33 proteins were differentially regulated compared to their expression in control fetus. We found that the gross differences in the protein expression patterns in the SCNT

Table 3 Down-regulated proteins in the SCNT extraembryonic tissue

Spot no.	Protein name	NCBI accession no.	Species	PMF (MS)			Experimental		Ideal	
				Matched peptides	Measured peptides	Sequence coverage (%)	Molecular mass (kDa)	pI	Molecular mass (kDa)	pI
2106	apolipoprotein A-I	AAA30992.1	<i>Sus scrofa</i>	24	24	61	30.31	5.4	26.42	4.67
1203	translational elongation factor 1 delta	NP_001009449.1	<i>Ovis aries</i>	9	9	27	30.97	5.4	38.83	4.43
1204	calpain, small subunit 1	NP_999483.1	<i>Sus scrofa</i>	8	14	21	28.16	5.0	29.96	4.45
1312	thioredoxin-like 1	NP_004777.1	<i>Homo sapiens</i>	13	14	48	32.63	4.8	39.48	4.57
1409	cytokeratin 19	NP_001015600.1	<i>Bos taurus</i>	14	24	33	43.87	4.9	47.00	4.52
1511	beta 5-tubulin	BAD08435.1	<i>Sus scrofa</i>	28	28	43	50.11	4.8	63.04	4.47
1601	gamma-aminobutyric acid (GABA) A receptor, alpha 2	1411303A	<i>Bos taurus</i>	10	13	23	51.46	9.3	67.05	4.32
1808	90-kDa heat shock protein alpha	NP_999138.1	<i>Sus scrofa</i>	26	28	38	85.11	4.9	112.70	4.53
1811	heat shock protein 90 beta	BAB20776.1	<i>Equus caballus</i>	28	42	35	82.29	5.0	112.11	4.58
2107	eukaryotic translation initiation factor 5A	NP_001003658.1	<i>Bos taurus</i>	8	12	42	17.04	5.1	13.78	4.69
2108	Peroxiredoxin-2 (Thioredoxin peroxidase 1) (Thioredoxin-dependent peroxide reductase 1) (Thiol-specific antioxidant protein) (TSA)	spP52552	<i>Sus scrofa</i>	6	6	34	13.82	4.7	23.67	4.72
2201	epsilon1-COP	BAA94967.1	<i>Bos taurus</i>	14	17	32	34.69	5.0	36.64	4.60
2203	cytochrome b	AAR10359.1	<i>Phoca caspica</i>	5	9	15	40.42	6.6	31.74	4.65
2204	Rho GDP dissociation inhibitor (GDI) beta	NP_786991.1	<i>Bos taurus</i>	5	6	30	22.83	5.1	29.40	4.67
2205	carbonic anhydrase III	NP_001008688.1	<i>Sus scrofa</i>	12	23	56	29.68	7.9	31.26	4.69
2206	pro alpha 1(I) collagen	CAC38832.1	<i>Bos taurus</i>	7	11	20	32.14	5.5	38.27	4.70
2301	tropomodulin 3 (ubiquitous)	NP_999459.1	<i>Sus scrofa</i>	13	15	41	39.74	5.0	45.66	4.67
2305	apolipoprotein E precursor	NP_999473.1	<i>Sus scrofa</i>	15	19	26	36.64	5.6	39.77	4.85
2610	Vimentin	spP02543	<i>Sus scrofa</i>	13	21	59	31.03	6.5	64.64	4.74
2611	Tubulin alpha-1A chain (Tubulin alpha-1 chain) (Alpha-tubulin 1)	spP02550	<i>Sus scrofa</i>	16	25	43	50.80	4.9	67.27	4.75
2709	6A3-5 protein	AAT75226.1	<i>Sus scrofa</i>	13	17	6	188.33	6.1	86.97	4.83
2815	valosin-containing protein	NP_999445.1	<i>Sus scrofa</i>	19	22	23	89.96	5.1	119.25	4.89
3206	apolipoprotein E precursor	NP_999473.1	<i>Sus scrofa</i>	23	25	44	36.64	5.6	38.43	5.00
3412	Eukaryotic initiation factor 4A-I (ATP-dependent RNA helicase eIF4A-1) (eIF4A-I) (eIF-4A-I)	spP29562	<i>Oryctolagus cuniculus</i>	29	31	59	45.50	5.3	50.39	5.11
3703	Structural maintenance of chromosomes protein 2 (Chromosome-associated protein E) (XCAP-E homolog) (FGF-inducible protein 16)	spQ8CG48	<i>Mus musculus</i>	13	14	15	134.77	8.7	98.12	4.94
3801	transglutaminase 2	NP_803473.1	<i>Bos taurus</i>	10	17	20	78.29	5.1	103.17	4.93
3805	valosin-containing protein	1303334A	<i>Sus scrofa</i>	34	38	43	89.97	5.1	118.87	4.95
5105	retinol-binding protein 4, plasma	spP27485	<i>Sus scrofa</i>	12	12	60	23.39	5.4	23.05	5.61
5117	stathmin-1	NP_001009582.1	<i>Sus scrofa</i>	5	5	29	17.28	5.8	13.97	5.52
6701	albumin	CAA30970.1	<i>Sus scrofa</i>	24	25	41	71.39	5.9	82.15	6.12

The 30 differentially down-regulated proteins (2.0- to 5.5-fold decreases) were identified according to protein identification with mass spectrophotometry and protein database search.



fetus, including genes involved in lipid binding proteins, translation elongation factors, ATP binding proteins, transcription factors, and metabolic regulation genes, were associated with placental abnormalities [15].

Pig extraembryonic tissue contains the trophoblast, a major component of the placenta that induces blastocyst attachment into the uterine wall, and regulates the development of both the fetal and maternal compartments of

the placenta [16]. The result from the failure to develop a normal trophoblast was believed to be the primary cause of early fetus lethality and defects. In the present study, our results showed that the size and shape of the fetus and amniotic sac of control and SCNT fetus were similar. However, the extraembryonic tissue derived from SCNT fetus was abnormally small, and it did not develop to the same degree as control fetus. SCNT fetus in mice showed

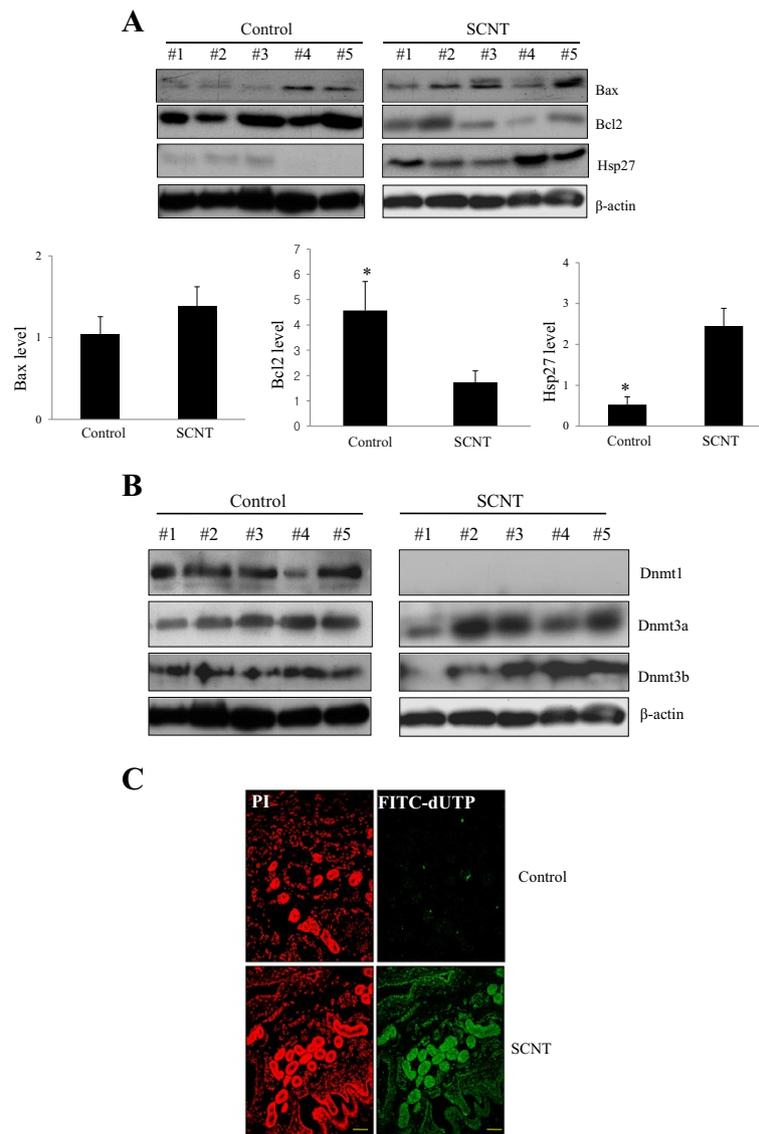


Figure 4 Up-regulated apoptotic marker proteins in the SCNT extraembryonic tissue. (A) The expression of Bax and Hsp27 were up-regulated and the Bcl2 expression was down-regulated in the SCNT extraembryonic tissue. Actin was used as a loading control. **(B)** Dnmt1 was highly expressed in the control extraembryonic tissue while the expression of Dnmt3a and 3b Dnmt3a and 3b were highly detectable in the SCNT extraembryonic tissue. **(C)** TUNEL assays in the control and SCNT extraembryonic tissue. Dewaxed paraffin sections from the control and SCNT extraembryonic tissue were subjected to the TUNEL assay. Fragmented DNAs were detected by a TUNEL assay using a fluorescence microscope with either propidium iodide (PI) (left) or FITC (FITC-dUTP) (right). In normal extraembryonic tissue, apoptotic cells were rarely identified while apoptotic cells significantly increased in SCNT extraembryonic tissue. Magnification: control and SCNT extraembryonic tissue (x200).

a large placenta size that was associated with hypertrophy of the basal layer, spongiotrophoblasts, giant trophoblasts, and glycogen cells [17]. These results obtained from pig SCNT fetus do not correspond to results obtained from mouse SCNT placentas, suggesting that these differences in placenta size are the result of different cell types of trophoblasts or implantation types in the uterus. These results strongly indicate that the extraembryonic tissue in SCNT fetus fails to develop normally, which causes subsequent

abnormalities in implantation and the development of the placenta.

During the normal development of SCNT fetus, epigenetic modifications in the somatic nucleus occur during preimplantation development of the SCNT fetus, with the silencing of differentiation-associated genes and the activation of genes critical for fetus development. Recently, many studies demonstrate that global gene expression is similar in SCNT and control fetus; however,

dysregulated gene expression is observed in the extraembryonic tissues of SCNTs [18]. Most SCNT fetus display abnormalities in their DNA methylation patterns and abnormal expression of several imprinted and nonimprinted genes, suggesting a relationship between these aberrations and abnormalities in the development of SCNT fetus [19,20]. Our data showed that the expression of Dnmt1 significantly decreased in SCNT extraembryonic tissue compared with extraembryonic tissue of a control fetus, whereas Dnmt3a and Dnmt3b were highly expressed in SCNT extraembryonic tissue. These results indicated that the expression of Dnmt1 is restricted in SCNT fetus and tightly controlled compared with control fetus, reflecting the occurrence in aberrant epigenetic reprogramming and aberrant protein expression in SCNT-derived extraembryonic tissue. Several studies demonstrated that cloned fetus frequently possess aberrant DNA methylation changes [21], and genome-wide aberrant demethylation was observed in cloned bovine fetus during preimplantation development [22]. Therefore, our data suggest that changes in DNA methylation may frequently occur in SCNT extraembryonic tissue, and it is possible that the cloned fetus are reprogrammed incompletely in their extraembryonic tissue and subsequently show abnormalities in protein expression.

Oxidative stress results from the endogenous or exogenous generation of reactive oxygen species (ROS), which damages macromolecules and causes apoptosis [23]. The decrease in the formation of ROS is derived by Superoxide dismutase, Glutathione Peroxidase (GPX), and catalase, which converts H_2O_2 to O_2 and H_2O [23]. In the present study, several antioxidant proteins were significantly down-regulated in the extraembryonic tissue of pig SCNT fetus. Our data showed that the catalase and GPX proteins were down-regulated in SCNT extraembryonic tissue via Western blotting analysis and immunohistochemistry. A recent study showed that increased oxygen demand leads to an increased rate of ROS production during embryonic development and oxidative stress, which contributes to ROS formation in the extraembryonic tissue of the placenta [24]. Moreover, SCNT extraembryonic tissue increased the expression of Hsp27, which induces apoptotic signaling by interacting with key components of the apoptosis pathway and activates the caspase signaling cascade [25]. The expression level of the antiapoptotic gene Bcl-2 was lower in SCNT extraembryonic tissue compared with control tissue, whereas the expression of the apoptotic marker gene Bax increased in the extraembryonic tissue of SCNT fetus. Taken together, these findings indicate that extraembryonic tissue from SCNT fetus induced apoptotic signaling, which was due to the upregulated expression of apoptosis-related proteins. Furthermore, our data showed that a small portion of the cells was TUNEL-positive in the extraembryonic tissue of control fetus,

whereas a large portion of the cells was TUNEL-positive in SCNT extraembryonic tissue, indicating that apoptosis occurred in SCNT fetus. Together, our data suggested that oxidative stress or apoptosis-related proteins activate apoptotic signaling, and apoptosis occurred in the pig SCNT extraembryonic tissue. These results demonstrated that abnormal apoptosis in the extraembryonic tissue of SCNT fetus during early embryonic development leads to the early loss of cloned pig fetus.

Conclusion

In the proteomic analysis using the extraembryonic tissue on the 35 day of pregnancy from the SCNT fetus, 33 proteins were identified as differentially regulated proteins. Among the up-regulated proteins, Hsp27 and Bax were found, which are closely related to the process of apoptosis. However, the expression levels of the catalase and GPX genes were down-regulated in SCNT fetus, suggesting that ROS accumulates in SCNT extraembryonic tissue and causes apoptosis. Therefore, we suggest that abnormal protein expression and apoptosis in the extraembryonic tissue of SCNT fetus is a primary reason for the high rates of embryonic death in pig SCNT fetus during early pregnancy.

Methods

Animal ethics

The treatment of the pigs used in this research abided the guidelines of the National Institute of Animal Science, Rural Development Administration, Suwon, South Korea, and was approved by the Institutional Animal Care and Use Committee (approval number: 2010-009, D-grade).

Production of cloned embryos

Ovaries were obtained from prepubertal crossbred gilts at a local slaughterhouse and transported to the laboratory at 30-35°C. COCs that had several layers of cumulus cells were selected and washed three times in maturation medium. For the maturation culture, approximately 50-100 COCs were transferred to 500 μ l of maturation medium (TCM-199, Gibco-BRL, Grand Island, NY, USA) covered with mineral oil in a four-well dish. Oocytes were matured at 38.5°C under 5% CO_2 in air for 40 to 44 hours. Primary culture of fetal fibroblasts was established from the ear skin tissue of the crossbred pig by routine cell culture technology [8,26]. The ear tissues were cut into small pieces and dispersed by exposure to 0.25% trypsin-0.02% EDTA (Gibco-BRL). Following digestion, the cells were centrifuged to remove debris and cultured in DMEM (Gibco-BRL) supplemented with 10% fetal bovine serum and 75 μ g/ml of antibiotics. Fibroblast cells were cultured and passed (2 to 8 passages) and then used as donor cells for nuclear transfer. After maturation, cumulus cells were removed from oocytes by vortexing the COCs in PBS

supplemented with 0.1% PVA and 0.1% hyaluronidase for 4 min. Oocytes were enucleated by aspirating the first polar body and metaphase-II (MII) plate in a small amount of surrounding cytoplasm with a glass pipette. All micromanipulation procedures were performed in TCM-199 supplemented with 3 mg/ml BSA and 5 μ l/ml cytochalasin B. Enucleation was confirmed by staining the oocytes with 10 μ g/ml Hoechst 33342 for 15–20 min at 39°C. After the enucleation, the oocytes were held in TCM-199 supplemented with 3 mg/ml BSA until injection of donor cells. Reconstructed oocytes were then placed between 0.2 mm diameter wire electrodes (1 mm apart) in a fusion chamber overlaid with 0.3 M mannitol solution supplemented with 0.1 mM MgSO₄, 1.0 mM CaCl₂, and 0.5 mM HEPES. For the fusion, two DC pulses of 1.2 kV/cm were applied for 30 μ s using a BTX Electro Cell Manipulator 2001 (BTX, San Diego, CA, USA). After the fusion treatment, the reconstructed oocytes were cultured in TCM-199 supplemented with 3 mg/ml BSA for 1 hr, and fusion was determined. The basic culture medium used was PZM-3. SCNT embryos were cultured in PZM-3 for 1 or 2 days prior to embryo transfer.

Extraembryonic tissues

Recipients were prepared as described previously [26]. The embryos produced by SCNT were cultured for 1 or 2 days. A total of 150 embryos that were morphologically normal at the 1–2 cell stages were selected for transfer. Embryos were then transferred into the oviducts of the recipient gilt approximately 48 h after human chorionic gonadotropin (hCG) injection. The pregnancy status of recipients was determined by ultrasound between days 30 and 33 after embryo transfer. The extraembryonic tissues including chorioallantois, umbilical cord, and placenta were obtained from the uterus of pregnant gilts at day 35 of gestation after transfer of SCNT embryos or natural mating (normal embryos). The size and shape of extraembryonic tissues were observed in the microscopic analysis. Three extraembryonic tissues surrounding each fetus which were recovered from three recipients were subjected to subsequent experiments.

2-D gel analysis and protein identification

2-D and spot analysis were performed as previously described [8]. From control and SCNT extraembryonic tissues were solubilized in lysis buffer containing 7 M urea, 2 M thiourea, 4% w/v CHAPS, 40 mM DTT, and 0.5% Pharmalyte at pH 4–7. Insoluble material was removed by centrifugation. Total proteins (1000 μ g) for analytical runs were transferred into IPG strips (17 cm, pH 4–7, BioRad) holder channels (BioRad, CA, USA). 2-D separated protein mixtures by IEF in the first dimension and SDS-PAGE in the second dimension. The resulting gels provided a high-resolution separation of a complex mixture

of proteins. The gels were stained with silver staining to visualize proteins. Silver staining kit (Amersham Biosciences) was used for 2-D gels. Three or four independent gels were performed in triplicate. The gels were fixed in 40% ethanol and 10% acetic acid for 30 min and sensitized in ethanol glutaraldehyde (25%, w/v), sodium thiosulfate (5%, w/v), and sodium acetate for 30 min followed by three washes with water for 15 min each. Then the gels were immersed in silver nitrate (2.5%, w/v) and formaldehyde (37%, w/v) for 20 min, developed with sodium carbonate and formaldehyde (37%, w/v) for 5 min, and stopped in EDTA-Na₂-2H₂O. Gels were scanned and Image analysis was performed using Progenesis SameSpots software (Nonlinear Dynamics, Newcastle, UK). Using this software, the differentially expressed spots were identified by automatic matching of the detected protein spots. Those spots differing significantly ($p < 0.05$, ANOVA test) in their intensities with a fold-change ≥ 2 were used for further analysis. Target spots, identified using PDQuest software (BioRad), were excised from the gel, destained, and subjected to in-gel digestion with bovine trypsin (Promega, USA). Briefly, trypsin digestion reactions were terminated with TFA (final concentration 10%). Peptides were concentrated and desalted using ZipTips (m-c18, Millipore, Etten-Leur, The Netherlands) and eluted directly onto the MALDI target in 1 mL of a saturated solution of CHCA in 50% ACN. Peptides were analyzed using a Voyager DE-STR MALDI-TOF mass spectrometer (Applied Biosystems, Framingham, MA, USA) in reflection mode, at a 20 kV accelerating voltage.

Immunohistochemical analysis

Tissues were fixed in 4% w/v paraformaldehyde in 0.01 M PBS (pH 7.4), washed in PBS, dehydrated in a series of ethanol washes (70, 90, and 100%) and embedded in paraffin. Sections were rehydrated (xylene for 5 min; 100, 95, 70% ethanol, 2 min each) and washed in distilled water prior to TUNEL staining. For TUNEL staining, sections were incubated for 15 min with proteinase K (20 mg/mL) at room temperature, washed with PBS, and then incubated in 2% H₂O₂ for 5 min to block the endogenous peroxidase activity. Sections were rewashed three times with PBS and incubated for 60 min at 37°C in a moist chamber with TUNEL mixtures (0.3 U/mL calf thymus terminal deoxynucleotidyl transferase, 7 pmol/mL biotin dUTP, and 1 mM cobalt chloride in 16 reaction buffer in distilled water). After washing in four PBS baths of 5 min each at room temperature, the sections were saturated in 2% BSA for 10 min at RT. Sections were treated for 30 min at 37°C in a moist chamber with a 1:20 dilution of ExtraAvidin peroxidase antibody. After three PBS washes, detection was performed with DAB (1.24 mg DAB, 25 mL 3% NiCl₂, 152 mL 1 M Tris-HCl (pH 7.5) in 2 mL distilled water). Slides were mounted in crystal

mount (Biomed, Foster City, CA). Immunohistochemical staining was performed with an ABC Kit (Oncogene Science, MA, USA), according to the manuscript's instruction. Sections were washed in water and mounted with a cover slip.

Western blot analysis

To further examine the differentially expressed genes between the control and SCNT extraembryonic tissues, we performed Western blot analysis using control and SCNT extraembryonic tissue. For Western blot analysis, 50 µg of soluble protein were loaded onto a 10% SDS/polyacrylamide gel and separated by electrophoresis, followed by electrotransfer to a PVDF membrane (Immobilon-P, Millipore) using a semidry blotting apparatus and a low ionic discontinuous buffer system. The membrane was incubated in TBST containing 5% skim milk and 0.05% Tween-20 and hybridized with primary antibodies. GPX, Catalase, Bax, Bcl2, and Hsp27 antibody were obtained from Cell Signaling Technology (MA, USA), Dnmt1, 3a, and 3b were from Santa Cruz Technology (Santa Cruz, CA, USA), and the monoclonal β-actin antibody was from Millipore (Darmstadt, Germany). HRP-labeled mouse anti-rabbit IgG were from Jackson ImmunoResearch. The Chemiluminescence kit was from Pierce (Rockford, IL). After incubation with horseradish-peroxidase-conjugated secondary antibody at room temperature, immunoreactive proteins were detected using a chemiluminescent ECL assay kit (Amersham Pharmacia, UK) according to the manufacturer's instructions.

Statistical analysis

Data are expressed as means ± SD. Significant difference among the treatment means were determined using ANOVA and Duncan's multiple range test at $p < 0.05$.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

YGK, SH, HK, HHS, and SWK performed the experiments. JHK, YS, BSY, YMS, and JHC conceived the idea, designed the experiments, and interpreted the experimental results. YGK and JHC wrote the manuscript. All authors contributed to manuscript preparations and approved the final manuscript.

Acknowledgements

This work was supported by grants from the Agenda Program (No.PJ0086322013), Rural Development Administration, Republic of Korea.

Author details

¹Animal Genetic Resources Station, National Institute of Animal Science, RDA, Namwon 590-832, Korea. ²Institute of Agriculture and Life Science, College of Veterinary Medicine, Gyeongsang National University, Jiju Daero 501, Jinju, Gyeongsangnamdo 660-701, Korea. ³Department of Animal Science & Biotechnology, Gyeongnam National University of Science and Technology, Jinju 660-758, Korea.

Received: 11 May 2014 Accepted: 18 November 2014
Published: 29 November 2014

References

1. Rogatcheva MM, Rund LA, Swanson KS, Marron BM, Beaver JE, Counter CM, Schook LB: **Creating porcine biomedical models through recombineering.** *Comp Funct Genomics* 2004, **5**:262–267.
2. Fujimura T, Takahagi Y, Shigeheisa T, Nagashima H, Miyagawa S, Shirakura R, Murakami H: **Production of alpha 1,3-galactosyltransferase gene-deficient pigs by somatic cell nuclear transfer: a novel selection method for gal alpha 1,3-Gal antigen-deficient cells.** *Mol Reprod Dev* 2008, **75**:1372–1378.
3. Onishi A, Iwamoto M, Akita T, Mikawa S, Takeda K, Awata T, Hanada H, Perry AC: **Pig cloning by microinjection of fetal fibroblast nuclei.** *Science* 2000, **289**:1188–1190.
4. Polejaeva IA, Chen SH, Vaught TD, Page RL, Mullins J, Ball S, Dai Y, Boone J, Walker S, Ayares DL, Colman A, Campbell KH: **Cloned pigs produced by nuclear transfer from adult somatic cells.** *Nature* 2000, **7**:86–90.
5. Hashizume K, Ishiwata H, Kizaki K, Yamada O, Takahashi T, Imai K, Patel OV, Akagi S, Shimizu M, Takahashi S, Katsuma S, Shiojima S, Hirasawa A, Tsujimoto G, Todoroki I, Izaika Y: **Implantation and placental development in somatic cell clone recipient cows.** *Clon Stem Cell* 2002, **4**:197–209.
6. Hill JR, Burghardt RC, Jones K, Long CR, Looney CR, Shin T, Spencer TE, Thompson JA, Winger QA, Westhusin ME: **Evidence for placental abnormality as the major cause of mortality in first-trimester somatic cell cloned bovine fetuses.** *Biol Reprod* 2000, **63**:1787–1794.
7. Hill JR, Edwards JF, Sawyer N, Blackwell C, Cibelli JB: **Placental anomalies in a viable cloned calf.** *Cloning* 2001, **3**:83–88.
8. Chae JI, Yu K, Cho SK, Kim JH, Koo DB, Lee KK, Han YM: **Aberrant expression of developmentally important signaling molecules in cloned porcine extraembryonic tissues.** *Proteomics* 2008, **8**:2724–2734.
9. Bauersachs S, Ulbrich SE, Zakhartchenko V, Minten M, Reichenbach M, Reichenbach HD, Blum H, Spencer TE, Wolf E: **The endometrium responds differently to cloned versus fertilized embryos.** *Proc Natl Acad Sci U S A* 2009, **106**:5681–5686.
10. Humpherys D, Eggan K, Akutsu H, Friedman A, Hochedlinger K, Yanagimachi R, Lander ES, Golub TR, Jaenisch R: **Abnormal gene expression in cloned mice derived from embryonic stem cell and cumulus cell nuclei.** *Proc Natl Acad Sci U S A* 2002, **99**:12889–12894.
11. Cho SK, Kim JH, Park JY, Choi YJ, Bang JI, Hwang KC, Cho EJ, Sohn SH, Uhm SJ, Koo DB, Lee KK, Kim T, Kim JH: **Serial cloning of pigs by somatic cell nuclear transfer: restoration of phenotypic normality during serial cloning.** *Dev Dyn* 2007, **236**:3369–3382.
12. Smith SC, Baker PN, Symonds EM: **Placental apoptosis in normal human pregnancy.** *Am J Obstet Gynecol* 1997, **177**:57–65.
13. Wisdom SJ, Wilson R, McKillop JH, Walker JJ: **Antioxidant systems in normal pregnancy and in pregnancy-induced hypertension.** *Am J Obstet Gynecol* 1991, **165**:1701–1704.
14. Chae JI, Lee KS, Kim DJ, Han YM, Lee DS, Lee KK, Koo DB: **Abnormal gene expression in extraembryonic tissue from cloned porcine embryos.** *Theriogenology* 2009, **71**:323–333.
15. Constant F, Guillomot M, Heyman Y, Vignon X, Laigre P, Servely JL, Renard JP, Chavatte-Palmer P: **Large offspring or large placenta syndrome? Morphometric analysis of late gestation bovine placentomes from somatic nuclear transfer pregnancies complicated by hydrallantois.** *Biol Reprod* 2006, **75**:122–130.
16. Strickland S, Richards WG: **Invasion of the trophoblasts.** *Cell* 1992, **71**:355–357.
17. Tanaka S, Oda M, Toyoshima Y, Wakayama T, Tanaka M, Yoshida N, Hattori N, Ohgane J, Yanagimachi R, Shiota K: **Placentomegaly in cloned mouse concepti caused by expansion of the spongiotrophoblast layer.** *Biol Reprod* 2001, **65**:1813–1821.
18. Chavatte-Palmer P, Camous S, Jammes H, Le Cleac'h N, Guillomot M, Leed RSF: **Placental perturbations induce the developmental abnormalities often observed in bovine somatic cell nuclear transfer.** *Placenta* 2012, **26**:99–104.
19. Beyhan Z, Ross PJ, Lager AE, Kocabas AM, Cunniff K, Rosa GJ, Cibelli JB: **Transcriptional reprogramming of somatic cell nuclei during preimplantation development of cloned bovine embryos.** *Dev Biol* 2007, **305**:637–649.
20. Nolen LD, Gao S, Han Z, Mann MR, Chung Y, Otte AP, Bartolomei MS, Latham KE: **X chromosome reactivation and regulation in cloned embryos.** *Dev Biol* 2005, **279**:525–540.
21. Dean W, Santos F, Stojkovic M, Zakhartchenko V, Walter J, Wolf E, Reik W: **Conservation of methylation reprogramming in mammalian development: aberrant reprogramming in cloned embryos.** *Proc Natl Acad Sci U S A* 2001, **98**:13734–13738.

22. Kang YK, Koo DB, Park JS, Choi YH, Chung AS, Lee KK, Han YM: **Aberrant methylation of donor genome in cloned bovine embryos.** *Nat Genet* 2001, **28**:173–177.
23. Hussain SP, Amstad P, He P, Robles A, Lupold S, Kaneko I, Ichimiya M, Sengupta S, Mechanic L, Okamura S, Hofseth LJ, Moake M, Nagashima M, Forrester KS, Harris CC: **p53-induced up-regulation of MnSOD and GPx but not catalase increases oxidative stress and apoptosis.** *Cancer Res* 2004, **64**:2350–2356.
24. Chae JI, Cho SK, Seo JW, Yoon TS, Lee KS, Kim JH, Lee KK, Han YM, Yu K: **Proteomic analysis of the extraembryonic tissue from cloned porcine embryos.** *Mol Cell Proteomics* 2006, **5**:1559–1566.
25. Concannon CG, Gorman AM, Samali A: **On the role of Hsp27 in regulating apoptosis.** *Apoptosis* 2003, **8**:61–70.
26. Park MR, Im GS, Kim SW, Hwang S, Park JH, Kim H, Do YJ, Park SB, Yang BS, Song YM, Cho JH, Ko YG: **Aberrant gene expression patterns in extraembryonic tissue from cloned porcine embryos.** *Res Vet Sci* 2013, **94**:531–538.

doi:10.1186/1756-0500-7-861

Cite this article as: Ko et al.: Proteomic analysis of the extraembryonic tissues from cloned porcine fetus at day 35 of pregnancy. *BMC Research Notes* 2014 **7**:861.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

