

Research

Open Access

## Outcomes of skin graft reconstructions with the use of Vacuum Assisted Closure (VAC®) dressing for irradiated extremity sarcoma defects

Alex Senchenkov\*<sup>1</sup>, Paul M Petty<sup>†2</sup>, James Knoetgen 3rd<sup>†2</sup>, Steven L Moran<sup>†3</sup>, Craig H Johnson<sup>†3</sup> and Ricky P Clay<sup>†2</sup>

Address: <sup>1</sup>Head and Neck Oncology Section, University of Cincinnati, 231 Albert B. Sabin Way, Cincinnati, OH, USA, <sup>2</sup>Plastic & Reconstructive Surgery, Mayo Clinic, 200 First Street SW, Rochester, MN 55905, USA and <sup>3</sup>Division of Plastic & Reconstructive Surgery and Department of Orthopedics Mayo Clinic, 200 First Street SW, Rochester, MN 55905, USA

Email: Alex Senchenkov\* - senchenkov@yahoo.com; Paul M Petty - petty.paul@mayo.edu; James Knoetgen - knoetgen.james@mayo.edu; Steven L Moran - moran.steven@mayo.edu; Craig H Johnson - johnson.craig@mayo.edu; Ricky P Clay - clay.ricky@mayo.edu

\* Corresponding author †Equal contributors

Published: 29 November 2007

Received: 25 August 2007

World Journal of Surgical Oncology 2007, 5:138 doi:10.1186/1477-7819-5-138

Accepted: 29 November 2007

This article is available from: <http://www.wjso.com/content/5/1/138>

© 2007 Senchenkov et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

**Background:** Flaps are currently the predominant method of reconstruction for irradiated wounds. The usefulness of split-thickness skin grafts (STSG) in this setting remains controversial. The purpose of this study is to examine the outcomes of STSGs in conjunction with VAC therapy used in the treatment of irradiated extremity wounds.

**Methods:** The records of 17 preoperatively radiated patients with extremity sarcomas reconstructed with STSGs in conjunction with VAC® therapy were reviewed regarding details of radiation treatment, wound closure, and outcomes.

**Results:** STSGs healed without complications (>95% of the graft take) in 12 (71%). Minor loss (6% – 20% surface) was noted in 3 patients (17.6%) and complete loss in 2 (11.7%). Two patients (11.7%) required flap reconstructions and 12 (88%) healed without further operative procedures.

**Conclusion:** Although flap coverage is an established treatment for radiated wounds, STSG in conjunction with liberal utilization of VAC therapy is an alternative for selected patients where acceptable soft tissue bed is preserved. Healing of the preoperatively radiated wounds can be achieved in the vast majority of such patients with minimal need for additional reconstructive operations.

### Background

Reconstructive surgeons are frequently confronted with irradiated post-ablative skin and soft tissue defects. Muscle and musculocutaneous flaps have been the traditional form of reconstruction in these patients, and little is known about outcomes of split-thickness skin grafts in the setting of preoperative radiation. In some cases, STSG

must be considered as the reconstructive option in patients with significant medical comorbidities, recurrence in the area of previous flap, or failed flap reconstruction that is not amenable to microvascular tissue transfer due to lack of recipient vessels. Historically, reported skin graft loss rates in preoperatively irradiated wounds varied from 30% – 100% [1-3].

Modern practice of reconstructive surgery is changing as evidenced by the improvement of surgical techniques, postoperative care, and especially wound care adjuncts. VAC® therapy may simplify reconstruction and improve the outcomes of skin grafts in cases of irradiated defects. The present study was undertaken to evaluate the outcomes of split-thickness skin grafts (STSGs) following oncologic resections in patients with musculoskeletal sarcomas who received preoperative radiation or were treated with locoregional radiation therapy in the past.

### Patients and methods

Retrospective review of the records was conducted to identify the patients who underwent STSG reconstruction of irradiated extremity defects in conjunction with Vacuum Assisted Closure (VAC®) therapy. Between January 1997 and December 2005, records of 19 such patients were identified and reviewed with permission of our institutional review board. All patients in this group had soft tissue sarcomas (Table 1). Prior to skin grafting, they were treated with external beam radiation to the tumor bed with the addition of intraoperative radiation or brachytherapy as dictated by treatment protocols.

All patients had a split thickness skin graft placed on an irradiated recipient bed that otherwise was appropriate for grafting (exposed muscle, vascularized soft tissue, or granulation tissue). The decision to reconstruct with a STSG as opposed to a flap was made by the authors on a case-by-case basis with consideration of the patient's physiological status, oncologic situation, defect characteristics, and patient's and surgeon's preference. The patients that had exposed critical structures such as major nerves, blood vessels, tendons with stripped peritenon, cortical bone,

and avascular joint capsule were not suitable candidates for grafting, and no such patients were found in the Mayo Clinic database. Irradiated defects with the exposure of aforementioned structures were appropriately treated with flaps.

There were 5 patients with local muscle flaps, which were unequivocally exposed to a full radiation dose. These patients were subsequently reconstructed with STSGs. The patients who had a skin graft applied to an axial pattern muscle flap, which did not have a full exposure to radiation therapy, were not included in the study. Two patients who had skin grafts of irradiated recipient beds were excluded from the study because one had necrosis of the underlying irradiated muscle flap, and the other had a 40% of skin graft placed with epidermis facing the recipient bed (Figure 1). The remaining 17 patients comprised the study group.

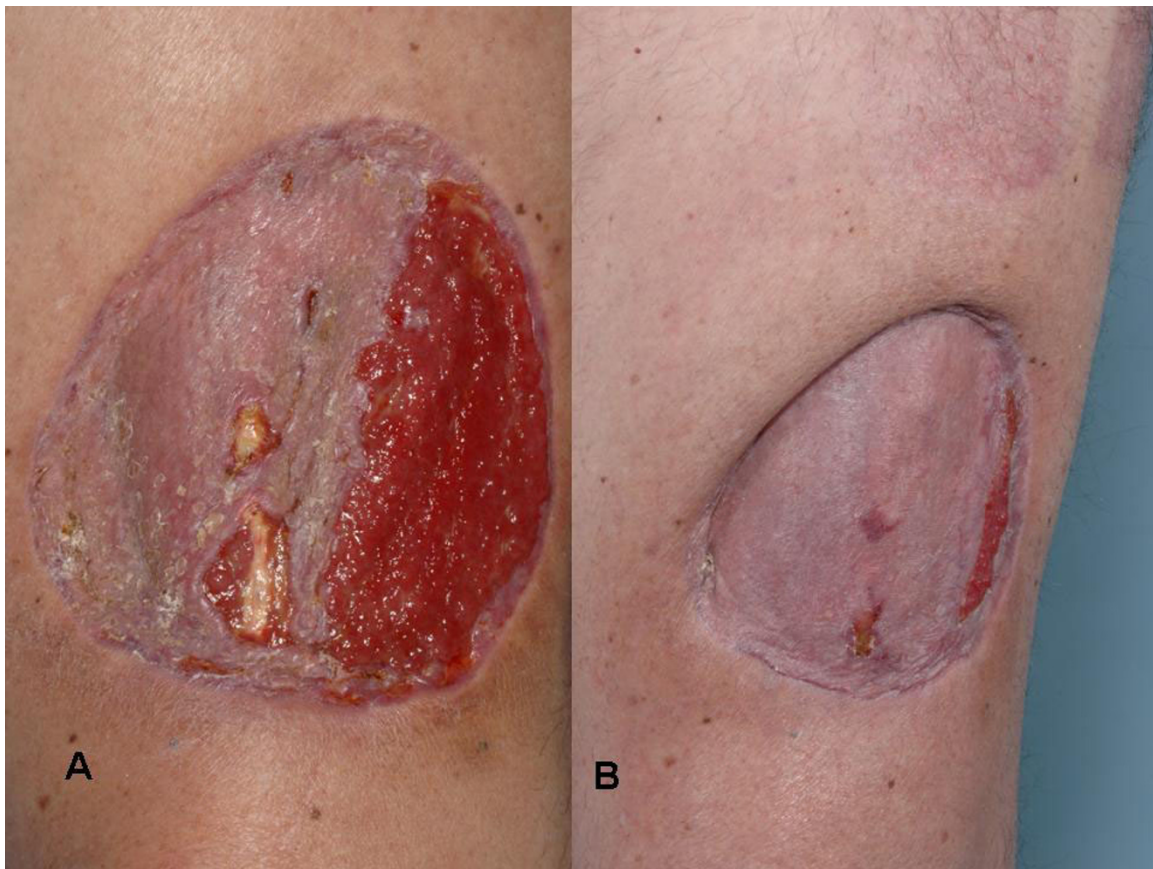
Skin grafting was performed as primary reconstruction at the time of tumor resection in 8 patients. Five patients had STSG performed 2 – 8 days following tumor resection as a delayed-primary reconstruction. During this delay, 3 patients underwent a 6-day course of brachytherapy when the brachytherapy afterloading catheters were covered with sterile VAC® dressing (Kinetic Concepts, Inc., San Antonio, TX). In the remaining 2 patients, the wound was temporarily covered with VAC® dressing and delayed-primary skin graft was performed within 2 and 8 days following tumor excision when the clear margins of resection were confirmed with permanent pathology. In these 13 patients who underwent primary or delayed-primary skin grafting, the time interval from completion of radiation therapy to skin graft reconstruction varied from 21 to 67 days, average 34 days.

Four patients underwent skin grafting for closure of preoperatively radiated complicated wounds. These wounds were treated with serial débridements and frequent dressing changes, and VAC® therapy. STSGs were used for reconstruction when the wound filled with healthy granulation tissue and was judged amenable to grafting.

All patients underwent reconstruction with 0.012–0.015-inch split-thickness skin grafts that were applied directly on the radiated recipient bed. All skin grafts in this series were meshed 1.5:1 ratio and secured in place with either staples or chromic suture. Xeroform gauze® (Tyco Healthcare Group, Mansfield, MA) or Furacin® (Shire US, Inc. Newport, KY) ointment on N-Terface® Interpositional Surface Material (Winfield Laboratories, Inc. Richardson, TX) were placed directly onto the skin graft prior to the VAC® dressing application. Inpatient VAC® therapy at 75 mm Hg in continuous mode was instituted to secure the split-thickness skin graft in place until postoperative day 5.

**Table 1: Histologic characteristics and distribution of primary tumors**

<b>Histology</b>	
Malignant fibrous histiocytoma	6
Fibrosarcoma	3
Liposarcoma	3
Angiosarcoma	1
Leiomyosarcoma	1
Synovial	1
Chondrosarcoma soft tissues	1
Fibromyxosarcoma	1
<b>Tumor location</b>	
Thigh	8
Lower leg	6
Upper arm	2
Forearm	1
<b>Total patients</b>	<b>17</b>



**Figure 1**

Forty percent graft surface failure of the irradiated defect due to upside down application of the skin graft. Notice prominence of granulation tissue in the irradiated wound in the area of failed skin graft in response to 4 days of VAC therapy (A). Interval healing progress after 7 weeks of local wound care (B).

During this time, the patients with lower extremity wounds were kept on bed rest. On the fifth postoperative day, the VAC<sup>®</sup> dressing was taken down. Following discharge, the patients performed Xeroform gauze<sup>®</sup> dressing changes once or twice a day for 4 – 6 weeks. All patients had the wounds re-examined postoperatively as a part of their oncologic follow up. Skin graft take was judged by gross inspection, and this information was extracted from the medical records. The end point of the study was complete healing of the wound with stable skin coverage.

## Results

### **Patients and oncologic treatments**

Skin grafts were performed on 17 patients (9 men and 8 women, age 42 to 82, mean 65). The sizes of skin grafts varied from 23 cm<sup>2</sup> to 240 cm<sup>2</sup>, mean 118 cm<sup>2</sup>. All patients had histologically confirmed high-grade (grade 3 or 4) soft tissue sarcomas (Table 1). Two patients had diabetes mellitus, one was a smoker, and none were on steroids. The usual radiation dose was from 50 to 62 Gy with

the exception of one patient with a recurrent tumor who received a total of 100-Gy to his recipient bed prior to skin graft reconstruction. On average, the patients received a cumulative dose of 59.3 Gy, ranging from 50 Gy 100 Gy.

### **Reconstructive settings**

Thirteen patients underwent skin grafting under sterile conditions in the setting of either immediate (primary) or delayed-primary reconstruction. In 4 patients, skin grafts were performed for tertiary intention closure of complicated wounds following surgical site infection and breakdown of primary closure (3 patients) and flap necrosis (1 patient). Thirteen patients had a STSG placed directly on a defect, and 4 patients had it applied to irradiated local muscles flaps (tibialis anterior – 2 and rectus femoris – 2).

### **Graft healing**

Twelve skin grafts (71%) had greater than 95% graft take and healed completely by primary intention; 3 patients (18%) lost between 6% and 20% of the graft surface; and

2 patients lost their entire graft. Three patients healed by secondary intention: 2 patients with 10% and 20% skin graft loss healed with dressing changes only and 1 patient with complete graft loss healed with wound VAC® therapy. Overall, 15 of 17 patients (88%) healed without further operative intervention, and only 2 patients required reoperation for tertiary intention closure of the defect. One patient with 10% graft loss with exposed tendons of the forearm required a free hemi-latissimus dorsi muscle flap, and the other patient with complete graft loss was salvaged with medial gastrocnemius muscle flaps (Table 2).

**VAC therapy and grafting results**

VAC® dressing was utilized in all patients to secure the STSG during the early postoperative period. Additionally, VAC® therapy was used for temporary sterile closure of open wounds (5 patients) that allowed delivering brachytherapy (3 patients) and obtaining permanent margins (2 patients) without committing to definitive reconstruction. Four patients with complicated irradiated wounds (3 wound infections and 1 flap loss) were managed with serial débridements to achieve control of the wound. Until granulation tissue build up was attained, these wounds were treated with VAC® therapy for 16 to 88 days, average 48 days. All 4 patients were skin grafted and successfully healed. Time intervals between skin grafting and completion of preoperative external beam radiation in these patients were 61, 93, 123, and 584 days; they had had open wounds for 51, 46, 82, and 16 days, respectively.

One patient deserves a special mention. This patient was originally treated with 45 Gy external beam radiation for primary synovial sarcoma of the lower leg, and the tumor was excised and closed primarily. The patient developed a local recurrence 2 years later and underwent wide-local excision with 35 Gy brachytherapy over the open wound. The defect was reconstructed with a STSG that initially healed, but was lost after receiving a 20-Gy course of additional postoperative external beam radiation therapy to the operated site. At that point, the wound was managed with VAC® therapy for 16 days until it filled with granulation tissue and then was successfully skin grafted after a cumulative radiation dose of 100 Gy.

**Discussion**

Split-thickness skin grafts can be used in conjunction with VAC® therapy for reconstruction of irradiated defects with acceptable results following preoperative radiation therapy of soft tissue sarcomas or remote exposure to radiation.

DNA damage is a hallmark of radiation injury to the cell that occurs during radiation therapy. This renders certain susceptible cells, particularly rapidly dividing tumor cells, reproductively incompetent and leads to cell program cell death, apoptosis. While many cells die, those which survive and continue to function have considerably impaired functions and proliferative capacity. This results in compromised wound healing, susceptibility to infections, and marked increase of postoperative wound morbidity [4-6]. These changes however occur in phases. Acute irradiation of the tissues over a short time leads to the initial increase in vascularity that peaks in the second week and then gradually decreases during 4<sup>th</sup> through 6<sup>th</sup> weeks as the wound passes the period of subacute inflammation. After the 8<sup>th</sup> week, vascular density in irradiated wounds becomes lower than it is in controls [7]. These microcirculatory changes are similar to those following radiation therapy and are related to endarteritis obliterans, fibrosis, disseminated thrombosis of the small vessels and chronic ischemia of the tissues [5,8]. Reconstruction is best carried out at the same time or within 4 – 6 weeks of resection, before chronic fibrous reaction sets in [8,9].

Surgery in an irradiated field requires sound clinical judgment. Every plastic surgery technique has been applied to reconstruction of radiated wounds, but there is no simple algorithm in deciding an optimal reconstructive strategy. Although analysis of anatomic characteristics of the defect is guided by general reconstructive principles, the decisions should be made with considerations of extent of radiation damage to the tissues, plans for adjuvant treatments and postoperative surveillance, and functional demands of the patient [2]. Operability and wound healing of an oncologic patient may be affected by malnutrition, anemia, immunosuppression, blood transfusions, chemotherapy, and often age-related medical conditions [10]. Surgical procedures in an irradiated field emphasize

**Table 2: Outcomes of split-thickness skin grafts with VAC utilization in 19 consecutive patients with irradiated extremity wounds**

Graft take (%)	Graft (n)	Healing of grafted wound (intention)		
		Primary	Secondary	Tertiary (salvage procedures)
95–100	12 (70.6%)	12	-	-
80–94	3 (17.6%)	-	2	1* (free flap)
0%	2(11.7)	-	1**	1 (Gastroc flap)

\* Exposed tendons  
 \*\*VAC therapy only

sterility to prevent bacterial contamination since these wounds are prone to infection. Meticulous operative technique calls for atraumatic tissue handling, tension-free closure, and obliteration of all dead spaces [6]. Wide, histologically-controlled, negative margins must be assured and, if local control of the tumor is in question, the tumor bed should be readily accessible to surveillance.

Pedicled or free muscular and musculocutaneous flaps allow bringing well-vascularized distant tissue from outside the radiation field that is resilient to both postoperative external beam radiation therapy and brachytherapy. They have an established track record in post-radiation reconstruction because they provide stable coverage, enhance wound healing, and decrease risk of wound breakdowns and infections [3,11]. Microvascular tissue transfer allows the greatest versatility in reconstruction of three-dimensional irradiated defects especially in the head and neck region. They can be safely anastomosed with irradiated vessels of the recipient site [12,13]. Distant non-irradiated pedicle flaps are a great asset, but within the radiated field they can be treacherous, and their use in chest wall reconstruction resulted in 32% wound complication rate and 14% total flap loss [14]. One of five irradiated local muscle flaps was lost in the present study, leading to the complete failure of the skin graft and an amputation of the extremity.

Split-thickness skin grafting is simple to perform and has a low morbidity. If final margins are found positive, prompt re-excision can be performed without the need of excision of the flap along with all the tissue planes of dissection and surgical drain tracks that were intraoperatively seeded with tumor. It facilitates oncologic surveillance of the tumors with a high rate of local recurrence, such as many soft tissue sarcomas. Concerns have been voiced regarding the use of STSGs in the setting of preoperative and postoperative radiation therapy, and this issue remains controversial. Animal data suggested that STSGs were vulnerable to adjuvant radiation and tolerated doses within only 25 Gy limit [15]. On the other hand, 90% of patients retained stability of their wound coverage with STSG following adjuvant radiation ( $59 \pm 0.9$  Gy) in the study from Memorial Sloan-Kettering Cancer Center [16].

There is no agreement about the use of STSGs for reconstruction of irradiated wounds. High failure rates without clear correlation with the radiation doses were historically reported in the literature [1-3] and no clear data on skin graft outcomes in irradiated wounds have been published to date. Because skin graft take is largely dependent on inosculation and neovascularization, failure of the STSG in an irradiated field was ascribed to vascular phenomena [9,17]. Changes in the vascular bed and fibrosis associated with the ensuing chronic phase of radiation insult nega-

tively influence the result of reconstructive operations. While in mild-to-moderate radiation impairment of the tissues, skin grafting could be considered [6], the success of skin grafts in an irradiated field is unpredictable. Rudolph reported 100% skin graft loss requiring reoperation regardless whether or not irradiated wounds were excised [3].

Laboratory and clinical studies have shown that the VAC<sup>®</sup> therapy increases wound blood flow, granulation tissue formation, and decreases accumulation of fluid and bacteria [18]. Recently, conformational changes in the cytoskeleton of the cells in response to application of micromechanical forces, i.e. stretch of the wound surface by the irregularity of the VAC sponge, was postulated to be an important factor in VAC-augmented wound healing [19]. Similar mechanism of stretch-induced cell proliferation is thought to be the driving force in tissue expansion [20,18,21] and distraction osteogenesis [22,23]. Initially applied for treatment of the chronic wounds, it was found to be useful in the management of acute wounds as well. VAC<sup>®</sup> therapy has been shown to hasten wound closure and the formation of granulation tissue in a variety of settings [24-26].

VAC<sup>®</sup> dressings are successfully used for securing skin grafts [24,25,27], especially in wounds with exudative, irregular, or mobile recipient beds and in difficult anatomic locations [25,28-30]. The manufacturer guidelines recommend continuous mode negative pressure of 75 mm Hg to 125 mm Hg [31]. We traditionally used 75 mm Hg for skin graft application although 125 mmHg negative pressure has also been used by other authors [25,27,32,33]. The VAC<sup>®</sup> stabilizes the skin graft and conforms well to the shape of recipient bed, removes fluid, decreases bacterial counts, and provides a secured dressing [27]. Improved graft survival and reduced need for repeat skin grafting were noted in one retrospective study [32].

The function of the VAC<sup>®</sup> technique in irradiated wounds is largely unknown and clinical experience is very limited [34]. In the present study, VAC<sup>®</sup> technique was used in the vast majority of patients in four types of settings: coverage of afterloading catheters during brachytherapy, optimization of the wound prior to grafting, securing the split-thickness skin graft, and secondary closure of skin graft losses.

Immediate flap reconstruction has been traditionally advocated for coverage of afterloading brachytherapy catheters [35,36]. Utilization of the VAC<sup>®</sup> in lieu of immediate flap reconstruction in sarcoma patients requiring brachytherapy provided rapid and simple temporary coverage with good stability of the catheters. It allows sparing

the flap from radiation insult and performing an elective delayed reconstruction after completion of brachytherapy [30]. Patients who underwent flap reconstructions were not included in this series. However, further study is indicated concerning the effects of temporary VAC® coverage on brachytherapy effectiveness. Theoretically, the increased blood flow and oxygenation may minimize postoperative tissue ischemia and improve effectiveness of brachytherapy.

VAC® therapy was also used for preoperative optimization of irradiated wounds to allow skin grafting of a viable, granulating bed as well as secondary closure of the skin graft breakdowns. Radiation primarily impairs small vessels, decreases their size and density [17], which causes local hypoxemia, decreased bacterial clearance, and impaired regeneration, but the VAC® counteracts all these effects. It quadruples local blood flow, promotes delivery of oxygen and nutrients, decreases bacterial counts, increases size and density of capillaries, and promotes the growth of granulation tissue [18,24,37,38]. Successful skin graft take was achieved in one patient who received a cumulative local dose of 100 Gy as a result of VAC® utilization for optimization of the wound and graft application.

The present study retrospectively analyzed a group of extremity sarcoma patients who received compatible doses of preoperative radiation and had a close follow up. These data give insight into the clinical behavior of STSGs following standard doses of preoperative radiation therapy commonly used in soft tissue sarcoma treatment. Despite small breakdowns of the skin grafts that eventually healed and are common in non-irradiated grafts, only 2 patients (11.7%) had complete graft loss. Eighty-eight percent of patients in this study eventually healed without requiring further operative procedures. The data demonstrate that the reconstruction of the suitable irradiated soft tissue wounds with STSG in conjunction with VAC® therapy may be considered as an acceptable reconstructive approach in the setting of questionable margins, high risk of recurrence, and poor physiological condition of the patient.

## Conclusion

Split-thickness skin grafting provides a simple one-stage reconstructive option for skin and soft tissue defects, but its use in the irradiated wound is controversial. In the present study of skin-grafting of post-ablative defects in the preoperatively radiated soft tissue sarcoma patients, we found that with meticulous surgical technique and utilization of VAC® therapy complete or partial skin graft take and complete healing of the wound without reoperation was achieved in 88% of cases. Complete (>95%) skin graft take was observed in 71% of cases, partial (80%–94%)

take in 18%, and complete loss of the graft in 12%. Only 12% of skin-grafted wounds required closure with an additional operation. Split-thickness skin graft reconstruction of irradiated skin defects can be performed following preoperative radiation therapy of soft tissue sarcoma patients in conjunction with VAC® therapy with acceptable skin graft take rate and minimal morbidity to the patients.

## Abbreviations

STSG: split-thickness skin graft

VAC: vacuum-assisted closure

Gastroc flap: gastrocnemius muscle flap

## Competing interests

The author(s) declare that they have no competing interests.

## Authors' contributions

AS: conception and design, acquisition of data, analysis and interpretation of data, drafting the manuscript, revising it critically for important intellectual content

PMP, JK, SLM, CHJ and RCP: conception and design, analysis and interpretation of data, revising the manuscript critically for important intellectual content

## Acknowledgements

The authors would like to thank Kimberly A. Garrett for her assistance in preparation of the manuscript.

## References

1. Kurul S, Dincer M, Kizir A, Uzunismail A, Darendeliler E: **Plastic surgery in irradiated areas: analysis of 200 consecutive cases.** *Eur J Surg Oncol* 1997, **23**:48-53.
2. Stotter A, McLean NR, Fallowfield ME, Breach NM, Westbury G: **Reconstruction after excision of soft tissue sarcomas of the limbs and trunk.** *Br J Surg* 1988, **75**:774-778.
3. Rudolph R: **Complications of surgery for radiotherapy skin damage.** *Plast Reconstr Surg* 1982, **70**:179-185.
4. Ariyan S, Marfuggi RA, Harder G, Goodie MM: **An experimental model to determine the effects of adjuvant therapy on the incidence of postoperative wound infection: I. Evaluating preoperative radiation therapy.** *Plast Reconstr Surg* 1980, **65**:328-337.
5. Ariyan S, Krizek TJ: **Radiation effects: biologic and surgical considerations.** In *Plastic Surgery Volume 1*. Edited by: McCarthy JG. Philadelphia: W. B. Saunders Company; 1990:831-848.
6. Miller MJ, Janjan NA: **Treatment of injuries from radiation therapy.** In *Reconstructive plastic surgery for cancer* Edited by: Kroll SS. Philadelphia: Mosby – Year Book, Inc; 1996:17-36.
7. Ueda M, Torii S, Kaneda T, Oka T: **Revascularization of autogenous skin grafts placed on irradiated tissue.** *J Oral Maxillofac Surg* 1982, **40**:477-481.
8. Young CM, Hopewell JW: **Surgical management of irradiated skin in the pig.** *Br J Radiol Suppl* 1986, **19**:125-128.
9. Ueda M, Torii S, Oka T: **An experimental study of skin autografts on irradiated tissue.** *J Oral Maxillofac Surg* 1982, **40**:74-77.
10. Chmell MJ, Schwartz HS: **Analysis of variables affecting wound healing after musculoskeletal sarcoma resections.** *J Surg Oncol* 1996, **61**:185-189.



11. Evans GRD: **Radiation effects.** In *Plastic surgery: indications, operations, and outcomes Volume 1*. Edited by: Achauer BM, Eriksson E. St. Louis: Mosby, Inc; 2000:409-424.
12. Bengtson BP, Schusterman MA, Baldwin BJ, Miller MJ, Reece GP, Kroll SS, Robb GL, Goepfert H: **Influence of prior radiotherapy on the development of postoperative complications and success of free tissue transfers in head and neck cancer reconstruction.** *Am J Surg* 1993, **166**:326-330.
13. Knoetgen J 3rd, Choudry U, Finical SJ, Johnson CH: **Head and neck reconstruction with a second free flap following resection of a recurrent malignancy.** *Ann Plast Surg* 2005, **55**:378-383.
14. Arnold PG, Lovich SF, Pairolero PC: **Muscle flaps in irradiated wounds: an account of 100 consecutive cases.** *Plast Reconstr Surg* 1994, **93**:324-327. discussion 328-329.
15. Tadjalli HE, Evans GR, Gurlek A, Beller TC, Ang KK, Stephens LC: **Skin graft survival after external beam irradiation.** *Plast Reconstr Surg* 1999, **103**:1902-1908.
16. Bui DT, Chunilal A, Mehrara BJ, Disa JJ, Alektiar KM, Cordeiro PG: **Outcome of split-thickness skin grafts after external beam radiotherapy.** *Ann Plast Surg* 2004, **52**:551-556. discussion 557.
17. Schultze-Mosgau S, Rodel F, Radespiel-Troger M, Worl J, Grabenbauer GG, Neukam FW: **Vascularization of the area between free grafts and irradiated graft beds in the neck in rats.** *Br J Oral Maxillofac Surg* 2002, **40**:37-44.
18. Morykwas MJ, Argenta LC, Shelton-Brown EI, McGuirt W: **Vacuum-assisted closure: a new method for wound control and treatment: animal studies and basic foundation.** *Ann Plast Surg* 1997, **38**:553-562.
19. Saxena V, Hwang CW, Huang S, Eichbaum Q, Ingber D, Orgill DP: **Vacuum-assisted closure: microdeformations of wounds and cell proliferation.** *Plast Reconstr Surg* 2004, **114**:1086-1096. discussion 1097-1088.
20. Olenius M, Dalsgaard CJ, Wickman M: **Mitotic activity in expanded human skin.** *Plast Reconstr Surg* 1993, **91**:213-216.
21. De Filippo RE, Atala A: **Stretch and growth: the molecular and physiologic influences of tissue expansion.** *Plast Reconstr Surg* 2002, **109**:2450-2462.
22. Ilizarov GA: **The tension-stress effect on the genesis and growth of tissues. Part I. The influence of stability of fixation and soft-tissue preservation.** *Clin Orthop Relat Res* 1989:249-281.
23. Ilizarov GA: **The tension-stress effect on the genesis and growth of tissues: Part II. The influence of the rate and frequency of distraction.** *Clin Orthop Relat Res* 1989:263-285.
24. Argenta LC, Morykwas MJ: **Vacuum-assisted closure: a new method for wound control and treatment: clinical experience.** *Ann Plast Surg* 1997, **38**:563-576. discussion 577.
25. Schneider AM, Morykwas MJ, Argenta LC: **A new and reliable method of securing skin grafts to the difficult recipient bed.** *Plast Reconstr Surg* 1998, **102**:1195-1198.
26. Armstrong DG, Lavery LA: **Negative pressure wound therapy after partial diabetic foot amputation: a multicentre, randomised controlled trial.** *Lancet* 2006, **366**:1704-1710.
27. Molnar JA, DeFranzo AJ, Marks MW: **Single-stage approach to skin grafting the exposed skull.** *Plast Reconstr Surg* 2000, **105**:174-177.
28. Hallberg H, Holmstrom H: **Vaginal construction with skin grafts and vacuum-assisted closure.** *Scand J Plast Reconstr Surg Hand Surg* 2003, **37**:97-101.
29. Weinfeld AB, Kelley P, Yuksel E, Tiwari P, Hsu P, Choo J, Hollier LH: **Circumferential negative-pressure dressing (VAC) to bolster skin grafts in the reconstruction of the penile shaft and scrotum.** *Ann Plast Surg* 2005, **54**:178-183.
30. Senchenkov A, Knoetgen J, Chrouser KL, Nehra A: **Application of vacuum-assisted closure dressing in penile skin graft reconstruction.** *Urology* 2006, **67**:416-419.
31. V.A.C.® *Therapy Clinical Guidelines: A reference source for clinicians* KCI Licensing, Inc; 2007.
32. Scherer LA, Shiver S, Chang M, Meredith JW, Owings JT: **The vacuum assisted closure device: a method of securing skin grafts and improving graft survival.** *Arch Surg* 2002, **137**:930-933. discussion 933-934.
33. Andrews BT, Smith RB, Goldstein DP, Funk GF: **Management of complicated head and neck wounds with vacuum-assisted closure system.** *Head Neck* 2006, **28**:974-981.
34. Schimp VL, Worley C, Brunello S, Levenback CC, Wolf JK, Sun CC, Bodurka DC, Ramirez PT: **Vacuum-assisted closure in the treatment of gynecologic oncology wound failures.** *Gynecol Oncol* 2004, **92**:586-591.
35. Duman H, Evans GR, Reece G, Kroll SS, Miller M, Langstein H, Chang D, Butler C, Robb G: **Brachytherapy: reconstructive options and the role of plastic surgery.** *Ann Plast Surg* 2000, **45**:477-480.
36. Lee HY, Cordeiro PG, Mehrara BJ, Singer S, Alektiar KM, Hu QY, Disa JJ: **Reconstruction after soft tissue sarcoma resection in the setting of brachytherapy: a 10-year experience.** *Ann Plast Surg* 2004, **52**:486-491. discussion 492.
37. Wackenfors A, Gustafsson R, Sjogren J, Algotsson L, Ingemansson R, Malmstro M: **Blood flow responses in the peristernal thoracic wall during vacuum-assisted closure therapy.** *Ann Thorac Surg* 2005, **79**:1724-1730. discussion 1730-1721.
38. Chen SZ, Li J, Li XY, Xu LS: **Effects of vacuum-assisted closure on wound microcirculation: an experimental study.** *Asian J Surg* 2005, **28**:211-217.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

