

Research article

Open Access

A polymorphism at codon 31 of gene p21 is not associated with primary open angle glaucoma in Caucasians

Thomas Ressiniotis^{1,2}, Philip G Griffiths¹, Sharon M Keers², Patrick F Chinnery*² and Michael Birch¹

Address: ¹Department of Ophthalmology, Royal Victoria Infirmary, Newcastle upon Tyne, UK and ²Department of Neurology, The Medical School, The University of Newcastle upon Tyne, UK

Email: Thomas Ressiniotis - tomres@doctors.org.uk; Philip G Griffiths - p.g.griffiths@ncl.ac.uk; Sharon M Keers - s.m.keers@ncl.ac.uk; Patrick F Chinnery* - p.f.chinnery@ncl.ac.uk; Michael Birch - birchmk@aol.com

* Corresponding author

Published: 04 April 2005

Received: 18 January 2005

BMC Ophthalmology 2005, 5:5 doi:10.1186/1471-2415-5-5

Accepted: 04 April 2005

This article is available from: <http://www.biomedcentral.com/1471-2415/5/5>

© 2005 Ressiniotis et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: Primary open angle glaucoma (POAG) is considered to be a neurodegenerative optic neuropathy, in which cell death occurs by apoptosis. *p21*, is an important protective component of the apoptotic pathway, regulating cellular arrest in the presence of DNA damage. An unstable or altered *p21* protein could modify the cellular response to genomic injury and abolish the effect of *p21*. A previous study on a Chinese cohort suggested that the *p21* codon 31 polymorphism may alter the state of apoptosis in glaucomatous optic neuropathy, failing to protect the ganglion cells. The aim of this study was to test the hypothesis that a *p21* codon 31 polymorphism is associated with POAG on a Caucasian cohort.

Methods: 140 POAG patients and a control group of 73 healthy individuals were included in the study. All the subjects were of Caucasian origin. Genomic DNA was amplified by polymerase chain reaction, followed by enzymatic restriction fragment length polymorphism technique (PCR-RFLP). Patients and controls were genotyped for a single nucleotide polymorphism (C/A transversion) in the third base of codon 31 of *p21*, which leads to a serine (Ser)/arginine (Arg) substitution.

Results: The distribution of the genotypes in the POAG patients showed 128 (91.4%) Ser homozygotes, 10 (7.1%) Ser/Arg heterozygotes and 2 (1.5%) Arg homozygotes. In the control cohort, there were 61 (83.6%) Ser homozygotes and 12 (16.4%) Ser/Arg heterozygotes. No Arg homozygotes were present amongst the control group. Both the allelic and genotypic frequencies of the Ser or Arg residues at codon 31 were not significantly different between POAG patients and controls (Fisher's exact test, $P = 0.20$ for alleles and $P = 0.0561$ for genotypes).

Conclusion: This study suggests that the *p21* codon 31 polymorphism does not contribute to the risk of POAG in the Caucasian population.

Background

Primary open angle glaucoma (POAG [MIM 137760]) is a multifactorial neurodegenerative disease, causing blind-

ness to approximately 70 million individuals worldwide [1,2]. The aetiology of POAG is poorly understood, with both genetic and environmental factors contributing to

the pathophysiology [3,4] Irrespectively of the causative factors leading to visual loss, most of which are still not understood, the final common pathway in POAG is retinal ganglion cell death, mediated by apoptosis, a genetically regulated process [5].

The apoptotic pathway consists of multiple interacting pathways, some of which are still unclear. It appears that, following DNA damage, cells can either proceed to apoptosis or enter a transient arrest cycle, allowing time for DNA repair. *p21* gene, also known as *WAF1* or *CIP1*, is a key component of this pathway. It can be up-regulated either by activated wild type p53, which acts as a transcription factor [6], or independently, by various factors such as TGF β , vitamin D, TPA and nerve growth factor. *p21* expression results in inhibition of the cyclin-dependent kinases (Cdks), that are essential for cell division. Consequently, cell cycle is arrested at the G1 phase, until genomic repair is established.

An unstable or altered p21 protein, therefore, could significantly affect the activity of Cdks, modifying the cellular response to genomic injury and abolishing the protective effect of p21. Such an outcome can derive from a single nucleotide polymorphism in the third base of codon 31 of the *p21* gene, following a C to A transverse change, which results in a Serine/Arginine amino acid substitution. This polymorphism probably encodes a probable DNA-binding zinc-finger domain, causing functional changes to the p21 protein.

There is evidence from a recent case-control study on a Chinese cohort that the Arg allele of the p21 codon 31 polymorphism is more common amongst POAG patients [7]. In this study, we tested the hypothesis of a possible association between the p21 codon 31 polymorphism and POAG on a Caucasian population.

Methods

Case selection

Having obtained ethical approval from our local research ethics committee, blood samples were analysed from an unrelated Caucasian cohort of 140 POAG patients and 73 controls from the north east of England.

The definition for POAG included characteristic cupping of the optic disc, open iridocorneal angle and typical glaucomatous visual field defects. An experienced glaucoma specialist clinically examined both patients and control groups (M.B.). Patients with intraocular pressure (IOP) higher than 30 mmHg at first presentation and secondary types of glaucoma (pseudoexfoliative, pigment dispersion syndrome, trauma or steroid induced) were excluded. The control group consisted of the spouses of our POAG patients, who were free from any coexisting ocular pathol-

ogy and had normal visual acuity, IOP, visual fields and optic discs.

Molecular genetic analysis

Purified genomic DNA was amplified by polymerase chain reaction (PCR) for exon 2 of the p21 gene. For each subject, 1 μ l of DNA was mixed with 1 unit Taq DNA polymerase (Promega, Madison, WI, USA), 10x Taq polymerase buffer (Promega, Madison, WI, USA), 2 mmol dNTP, 0.25 μ M of each oligonucleotide (primer) and H₂O to total volume of 30 μ l. The primers used were: Forward (5'-GTC AGA ACC GGC TGG GGA TG -3') and reverse (5'-CTC CTC CCA ACT CAT CCC GG -3'). Reactions were treated in a thermal cycle machine to incubation at 94°C for 5 min followed by 35 cycles of: 94°C for 30 sec, 57.2°C for 30 sec, 72°C for 30 sec and a final incubation of 72°C for 7 min.

For each sample, the amplified PCR product was digested with the restriction enzyme *Blp* I (New England Biolabs, Beverly, Massachusetts, USA). The *Blp* I digestion mixture contained 10 μ l PCR product, 6 units of enzyme, 3 μ l buffer (NEB Buffer 4) and H₂O to total volume of 20 μ l. The reactions were allowed to proceed for 12 hours at 37°C.

The resulting fragments were separated by electrophoresis on a 3% agarose gel and visualised by ethidium bromide staining with a digital camera. The Ser allele has a single *Blp* I restriction site (GCTNAGC), resulting in two fragments of 89 bp and 183 bp, where the Arg allele remains undigested, producing a single band of 272 bp.

The molecular genetic analysis was performed in the same laboratory by two investigators who were masked to the phenotype of the samples studied.

Fisher's exact test was used to compare the genotype and allele frequencies in cases and controls.

Results

Our cohort consisted of 140 POAG patients and 73 controls. The median age was 73 years for the POAG patients (range 51–87, S.D. = 8.01) and 78 years for the controls (range 68–90, S.D. = 4.4). Mean IOP was 20.8 mmHg for the patients (S.D. = 2.6) and 16.2 mmHg for the controls (S.D. = 3.4). Median cup/disc ratio was 0.8 and 0.3 for patients and controls respectively. The frequency distribution and the allelic frequencies of p21 codon 31 polymorphisms in POAG patients and healthy subjects are illustrated in Table 1 and 2 respectively. The allele frequencies follow Hardy-Weinberg proportions.

The distribution of the genotypes in the POAG patients was 128 (91.4%) Ser homozygotes, 10 (7.1%) Ser/Arg

Table 1: Frequency distribution of p21 codon 31 polymorphisms in POAG patients and healthy subjects. "Ser" and "Arg" represent encoding of Serine and Arginine, respectively, from the polymorphic site on exon 2.

	Ser/Ser	Ser/Arg	Arg/Arg	Total
POAG (n = 140)	128 (91.4%)	10 (7.1%)	2 (1.5%)	140
Controls (n = 73)	61 (83.6%)	12 (16.4%)	0 (0.0%)	73
Total	189 (88.8%)	22 (10.3%)	2 (0.9%)	213

Fisher's exact test: The two-tailed P value equals 0.0561. (Note: The frequencies of the Arg homozygotes were excluded from the analysis, as they were very low in the POAG group and 0 in the control group)

Table 2: Allelic frequencies in POAG patients and healthy subjects.

	Ser	Arg	Total
POAG	266 (95%)	14 (5%)	280
Controls	134 (91.8%)	12 (8.2%)	146
Total	400	26	426

Fisher's exact test: The two-tailed P value equals 0.20.

heterozygotes and 2 (1.5%) Arg homozygotes. In the control cohort, there were 61 (83.6%) Ser homozygotes and 12 (16.4%) Ser/Arg heterozygotes. No Arg homozygotes were present amongst the control group. We therefore pooled the Arg/Arg groups with the Ser/Arg group for the genotype analysis. Both the allelic and genotypic frequencies of the Ser or Arg residues at codon 31 were not significantly different between POAG patients and controls (Fisher's exact test, $P = 0.20$ for alleles and $P = 0.0561$ for genotypes).

Discussion

Apoptosis and cell-cycle arrest are regulated through various interacting pathways [8]. DNA damage leads to activation of transcription factors, such as the *p53* gene, which can induce apoptosis by up-regulating the protein Bax and down-regulating the protein Bcl-2. Alternatively, DNA insult can cause activation of the *p21* gene, a tumour suppressor gene, either directly or through transactivation by wild type *p53*. *p21* is a cyclin – dependent kinase inhibitor resulting in cell-cycle arrest by inhibiting the G1 to S and S to G2 phases [9]. Cancer research has revealed that mutations of *p21* are very rare and that single nucleotide polymorphisms are more likely to have a functional effect [10]. The Ser/Arg polymorphism at codon 31 is located in a highly conserved region of the *p21* [11] and has been associated with cancer of the lungs, breast [12], bladder [13], and colorectal tumours [14].

A recent study on a Chinese population showed an association between the Arg form of the *p21* codon 31 polymorphism and POAG, suggesting that this allele may alter the state of apoptosis in glaucomatous optic neuropathy, failing to protect the ganglion cells [7].

In our cohort, we have not detected any statistically significant difference of the Ser and Arg allelic frequencies between the POAG group and the healthy individuals. This difference may reflect sampling bias, as the Chinese cohorts were smaller (58 POAG patients and 59 control subjects), or it could be attributed to ethnic disparity. The Arg allele is considerably more common in Chinese, with reported frequency of 0.50. Our findings correlate well with previous studies on Swedish and French populations, which identified a low Arg allele frequency of 0.05 [15].

However, we cannot exclude the possibility of *p21* codon 31 polymorphism being associated with POAG in other ethnic groups or the likelihood of an association between POAG and another *p21* polymorphism. This might be a worthwhile future strategy to elucidate the issue.

References

1. Quigley HA, Vitale S: **Models of open-angle glaucoma prevalence and incidence in the United States.** *Invest Ophthalmol Vis Sci* 1997, **38**:83-91.
2. Coleman AL: **Glaucoma.** *Lancet* 1999, **354**:1803-1810.
3. Lichter PR: **Genetic clues to glaucoma's secrets. The L Edward Jackson Memorial Lecture. Part 2.** *Am J Ophthalmol* 1994, **117**:706-727.
4. WuDunn D: **Genetic basis of glaucoma.** *Curr Opin Ophthalmol* 2002, **13**:55-60.
5. Nickells RW: **Apoptosis of retinal ganglion cells in glaucoma: an update of the molecular pathways involved in cell death.** *Surv Ophthalmol* 1999, **43 Suppl 1**:S151-61.
6. Levine AJ: **p53, the cellular gatekeeper for growth and division.** *Cell* 1997, **88**:323-331.
7. Tsai FJ, Lin HJ, Chen WC, Tsai CH, Tsai SW: **A codon 31ser-arg polymorphism of the WAF-1/CIP-1/p21/tumour suppressor gene in Chinese primary open-angle glaucoma.** *Acta Ophthalmol Scand* 2004, **82**:76-80.
8. Cox LS: **Multiple pathways control cell growth and transformation: overlapping and independent activities of p53 and p21Cip1/WAF1/Sdi1.** *J Pathol* 1997, **183**:134-140.
9. el-Deiry WS, Harper JW, O'Connor PM, Velculescu VE, Canman CE, Jackman J, Pietenpol JA, Burrell M, Hill DE, Wang Y, et al.: **WAF1/CIP1 is induced in p53-mediated G1 arrest and apoptosis.** *Cancer Res* 1994, **54**:1169-1174.
10. Terry LA, Boyd J, Alcorta D, Lyon T, Solomon G, Hannon G, Berchuck A, Beach D, Barrett JC: **Mutational analysis of the p21/WAF1/CIP1/Sdi1 coding region in human tumor cell lines.** *Mol Carcinog* 1996, **16**:221-228.
11. Chedid M, Michieli P, Lengel C, Huppi K, Givol D: **A single nucleotide substitution at codon 31 (Ser/Arg) defines a polymorphism in a highly conserved region of the p53-inducible gene WAF1/CIP1.** *Oncogene* 1994, **9**:3021-3024.
12. Keshava C, Frye BL, Wolff MS, McCanlies EC, Weston A: **Waf-1 (p21) and p53 polymorphisms in breast cancer.** *Cancer Epidemiol Biomarkers Prev* 2002, **11**:127-130.
13. Chen WC, Wu HC, Hsu CD, Chen HY, Tsai FJ: **p21 gene codon 31 polymorphism is associated with bladder cancer.** *Urol Oncol* 2002, **7**:63-66.
14. Li YJ, Laurent-Puig P, Salmon RJ, Thomas G, Hamelin R: **Polymorphisms and probable lack of mutation in the WAF1-CIP1 gene in colorectal cancer.** *Oncogene* 1995, **10**:599-601.

15. Birgander R, Sjalander A, Saha N, Spitsyn V, Beckman L, Beckman G: **The codon 31 polymorphism of the p53-inducible gene p21 shows distinct differences between major ethnic groups.** *Hum Hered* 1996, **46**:148-154.

Pre-publication history

The pre-publication history for this paper can be accessed here:

<http://www.biomedcentral.com/1471-2415/5/5/prepub>

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

