

RESEARCH ARTICLE

Open Access

Fractal-based analysis of optical coherence tomography data to quantify retinal tissue damage

Gábor Márk Somfai¹, Erika Tátrai¹, Lenke Laurik¹, Boglárka E Varga¹, Vera Ölvedy¹, William E Smiddy², Robert Tchitnga³, Anikó Somogyi⁴ and Delia Cabrera DeBuc^{2*}

Abstract

Background: The sensitivity of Optical Coherence Tomography (OCT) images to identify retinal tissue morphology characterized by early neural loss from normal healthy eyes is tested by calculating structural information and fractal dimension. OCT data from 74 healthy eyes and 43 eyes with type 1 diabetes mellitus with mild diabetic retinopathy (MDR) on biomicroscopy was analyzed using a custom-built algorithm (OCTRIMA) to measure locally the intraretinal layer thickness. A power spectrum method was used to calculate the fractal dimension in intraretinal regions of interest identified in the images. ANOVA followed by Newman-Keuls post-hoc analyses were used to test for differences between pathological and normal groups. A modified p value of <0.001 was considered statistically significant. Receiver operating characteristic (ROC) curves were constructed to describe the ability of each parameter to discriminate between eyes of pathological patients and normal healthy eyes.

Results: Fractal dimension was higher for all the layers (except the GCL + IPL and INL) in MDR eyes compared to normal healthy eyes. When comparing MDR with normal healthy eyes, the highest AUROC values estimated for the fractal dimension were observed for GCL + IPL and INL. The maximum discrimination value for fractal dimension of 0.96 (standard error = 0.025) for the GCL + IPL complex was obtained at a FD \leq 1.66 (cut off point, asymptotic 95% Confidence Interval: lower-upper bound = 0.905-1.002). Moreover, the highest AUROC values estimated for the thickness measurements were observed for the OPL, GCL + IPL and OS. Particularly, when comparing MDR eyes with control healthy eyes, we found that the fractal dimension of the GCL + IPL complex was significantly better at diagnosing early DR, compared to the standard thickness measurement.

Conclusions: Our results suggest that the GCL + IPL complex, OPL and OS are more susceptible to initial damage when comparing MDR with control healthy eyes. Fractal analysis provided a better sensitivity, offering a potential diagnostic predictor for detecting early neurodegeneration in the retina.

Keywords: Optical coherence tomography, Fractal analysis, Fractal dimension, Wavelet algorithm, Diabetic retinopathy, Ophthalmology

Background

Optical coherence tomography (OCT) is a real-time, non-invasive imaging modality that employs interferometry to detect backscattered near-infrared light to render two-dimensional (2D) or three-dimensional (3D) images of tissue. OCT is a powerful tool for retinal measurement [1]. Particularly, OCT has been used to measure volume and total thickness of the retina along with structural changes of the various cellular layers of the retina with the

aid of segmentation algorithms [2,3]. The role of OCT in the assessment and management of retinal diseases has become significant in understanding the vitreoretinal relationships and the internal architecture of the retinal structure. Particularly, structural information extracted from OCT retinal images has been used to characterize early neural loss in patients with diabetes and multiple sclerosis [4,5]. The most important retinal pathology caused by diabetes is diabetic retinopathy (DR), which is characterized by blood vessels damage.

OCT has also improved diagnosis and management of retinal diseases by reducing reliance on insensitive tests such as perimetry and subjective disc grading. Though

* Correspondence: dcabrera2@med.miami.edu

²Miller School of Medicine, Bascom Palmer Eye Institute, University of Miami, Miami, Florida 33136, USA

Full list of author information is available at the end of the article

thickness differences may characterize regions with early pathological signs from normal regions, differences in optical properties and texture descriptors of normal and abnormal retinal tissue may also provide additional information of disease development in pathological eyes. The appropriateness of texture to classify tissues in OCT images has been shown in previous studies [6]. We have also shown that diabetic retinopathy not only causes thinning of the inner retinal layers, but also reduces the amplitude of the back-reflected signal from these layers [7-9]. Therefore, predictors based on optical properties changes are also of interest. Differences in optical properties and roughness measures of normal and abnormal retinal tissue may provide additional information of disease development in pathological eyes.

The fractal analysis of biological structures has been a continuous area under discussion ever since Mandelbrot's famous essay [10]. Fractal analysis techniques are common tools in physics and image processing. Fractals are objects that show self-similarity at different magnifications. One of the advantages of fractal analysis is the ability to quantify the irregularity and complexity of objects with a measurable value, which is called the fractal dimension [10]. The fractal dimension is a measure of the roughness of a fractal structure. Higher values indicate rougher surface. Fractal dimension is regarded as local property of the system. Fractal analysis has also been used for the description of texture in medical images [11]. Texture can be defined as the spatial distribution of intensity values in an image. In our particular case, texture can be defined as the spatial distribution of intensity values in an OCT image, where the intensity at each pixel is the back-reflection of the incident light. The back-reflected light contains information of the retinal structure such as the directionality, function and dysfunction (in the case of pathological retina) of the cellular layers. In ophthalmology, a major interest has been focused on the fractal properties of the retinal vasculature especially for diagnosis purpose. Most of the studies have used differences in the fractal dimension as a discriminant factor to detect and diagnose eye disease [12-15]. In general, a global measure characterizing the whole branching pattern of the retinal vascular network has been used as a single parameter in these previous studies. However, the global analysis of the vascular network features may overlook the very early changes in the structure and, therefore, not be sensitive to the early manifestation of the particular disease. Up to now, fractal-based analysis of OCT data has been used to quantify photoreceptor rearrangement and vision restoration, identify early glaucomatous damage in the retinal nerve fiber layer and as an index for capillary integrity of pathological disorders [16-18]. However, it has not been implemented to differentiate normal healthy eyes from pathological eyes with early neural loss in multiple

intraretinal layers (e.g. in DR and multiple sclerosis) using a local approach through segmentation of the various cellular layers of the retina and characterization of texture-based features on OCT intensity images.

In this study, the sensitivity of OCT images to identify retinal tissue morphology characterized by early neural loss in diabetes from normal healthy eyes is tested by calculating structural information and fractal dimension of the various cellular layers of the retina. Particularly, we found that fractal analysis provided a better sensitivity, offering a potential diagnostic predictor for detecting early neurodegeneration in the diabetic retina.

Methods

In this study, we evaluated the diagnostic power of a novel method based on the fractal analysis of OCT-derived retinal tissue layer properties in discriminating normal healthy eyes from diabetic eyes with early neural loss. Although texture measures of the retinal tissue are not standardized measures for detecting significant intraretinal changes, texture-based measures were obtained from OCT intensity images and used in the fractal dimension analysis. In addition, the fractal analysis' diagnostic outcome was compared with the standard approach that uses structural information extracted from OCT images. Specifically, we calculated fractal dimension and thickness using features measured locally for each intraretinal layer and evaluated their suitability to quantify retinal tissue damage.

Study population

The study was approved by the Institutional Review Board in each institution involved in the study (University of Miami, Miami, FL, USA and Semmelweis University, Budapest, Hungary). The research adhered to the tenets set forth in the declaration of Helsinki and written informed consent was obtained from each subject. In this prospective study, enrollment was offered to type 1 diabetic patients referred to the comprehensive ophthalmology clinic that had diabetic retinopathy up to ETDRS level 35 and without macular edema, as well as type 1 diabetic patients with no retinopathy [19]. Patients with proliferative disease, clinically significant macular edema (CSME), and anatomic abnormalities that might confound evaluation of macular architecture, such as glaucoma, vitreoretinal traction and epiretinal membranes were excluded. Healthy controls were selected if best-corrected visual acuity was at least 20/25, a history of any current ocular or systematic disease was lacking, and the macula appeared normal when examined with contact lens biomicroscopy. Patients with medical conditions that might affect visual function, receiving treatments with medications that might affect retinal thickness (e.g. chloroquine or niacin containing anticholesterol agents), recent

cataract surgery, previous vitrectomy, or unstable blood sugars were excluded.

Once enrolled a comprehensive eye examination was performed including slit lamp examination, measurement of intraocular pressure (using Goldmann tonometer), and fundus biomicroscopy. OCT imaging and 2 standard field stereoscopic fundus photos (SFPs) were obtained in all patients. The SFPs were classified by independent graders according to the criteria of proposed international clinical diabetic retinopathy and diabetic macular edema disease severity scales based on the ETDRS protocol [20,21]. The graders were unaware of the OCT findings and clinical data. In addition, a hemoglobin A1c level test was required at this visit for diabetic patients.

OCT data analysis and measurement of fractal dimension

The appropriateness of texture to classify tissues in OCT images has been shown in previous studies [6]. By analyzing the spatial arrangement of intensities in an image or selected region of interest (ROI), the image irregularities can be measured. Because the apparent reflectivity measured by OCT is a combination of the actual reflectivity and the scattering and absorption characteristics of the overlying media, the reflectivity measured by OCT may be affected by abnormalities in the retinal tissue. Consequently, structure disorder in the retinal tissue can be assessed when the fractal dimension is calculated using the intensity or reflectivity profile along the direction of depth in OCT images. Therefore, the fractal dimension was analyzed for each intraretinal layer segmented on OCT images and used as an indicator of retinal structure disorder or roughness measure.

A method based on the power spectrum was used to calculate the fractal dimension in OCT images [22]. Since the average power spectrum of an image obeys a power law scaling, the fractal dimension was calculated from the power law detected in the graph of the power spectrum as a function of the frequency in the Fourier transform of the OCT image (gray scale). In this particular case, when the graph is plotted in a log-log scale the curve is approximately similar to a straight line and the dimension is provided by the slope of the line. The fast Fourier transform (FFT) was applied to the OCT reflectivity's profiles (see Figure 1) to obtain the power spectrum as follows:

$$P(\omega) \sim \omega^{-\beta} \quad (1)$$

Where $P(\omega)$ is the power spectrum with the frequency ω . β is the spectral exponent of the reflectivity profile. The equation (1) can be converted into:

$$\ln(P(\omega)) \sim -\beta \ln(\omega) \quad (2)$$

The fractal dimension is linked to the power-law exponent β by the following relationship [22]:

$$FD = \frac{5-\beta}{2} \quad (3)$$

Therefore, the fractal dimension was evaluated from the slope β of a least-square regression line fit (polynomial regression of degree 1) to the data points in log-log plot of power spectrum. The fractal dimension was calculated for the reflectivity profile within each intraretinal layer for each A-scan (see Figure 1). The mean value of the fractal dimension was calculated by averaging the fractal dimension measurements across all A-scans in each macular region of each intraretinal layer. MATLAB software (The Mathworks, Natick, MA) was used to perform the fractal dimension analysis using a custom-built algorithm.

All Stratus OCT study cases were obtained using the "macular thickness" map protocol. This protocol consists of six radial scan lines centered on the fovea, each having a 6 mm transverse length. Macular radial line scans of the retina for each case were exported to disc with the export feature available in the Stratus OCT device and analyzed using a custom-built software (OCTRIMA) that facilitates the automatic segmentation of 7 cellular layers of the retina on OCT images based on their optical densities (see Figure 2). These retinal layers are the retinal nerve fiber layer (RNFL), the ganglion cell and inner plexiform layer complex (GCL + IPL), the inner nuclear layer (INL), the outer plexiform layer (OPL), the outer nuclear layer and inner photoreceptor segment (ONL + IS), outer photoreceptor segment (OS) and retinal pigment epithelium (RPE) [3]. Details of the methodology, such as segmentation, speckle noise removal and semiautomatic correction of discontinuities in each detected boundary after automated segmentation, along with manual error correction using direct visual evaluation of the detected boundaries, have been described in detail elsewhere [3-5,7-9,23-26].

Each OCT image used in this study was composed of 512 A-scans. Lateral coordinates of the blood vessel shadows were first extracted by using a blood vessel shadowgram technique and removed in each OCT image before calculating parameters related to reflectivity values [27].

In brief, we used the image gradient to detect edges such as the boundaries of blood vessel shadows for the shadowgram technique. With a proper threshold, locations of blood vessel shadows can be found in OCT images [27]. As the incident light perpendicularly penetrates into the retinal tissue, the direction of the blood vessel shadows' boundaries are vertical in OCT images which was employed to detect the lateral coordinates of

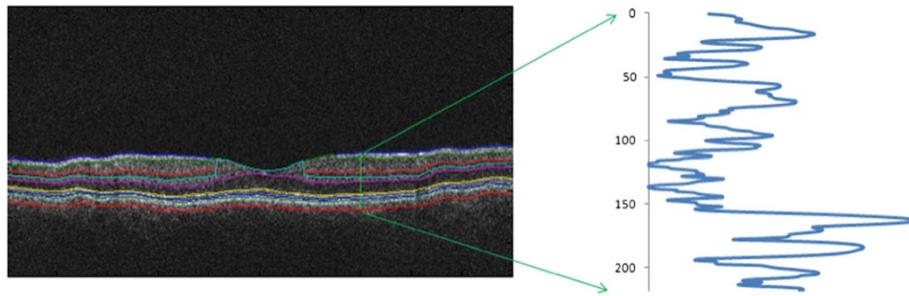


Figure 1 Reflectivity profile used to calculate the fractal dimension. The fractal dimension was calculated for the reflectivity profile within each intraretinal layer for each A-scan.

the blood vessel shadows [27]. The algorithm flowchart is shown on Figure 3 while Figure 4 shows an example of the use of the shadowgram technique.

Mean reflectivity values per intraretinal layer were normalized to the RPE reflectance and used in the analyses. Mean thickness values per intraretinal layer were obtained by calculating the mean distance between the boundaries comprising each layer. The mean values were calculated per intraretinal layer across the six radial OCT scans. We have previously shown the high repeatability and reproducibility of OCTRIMA measurements [23,24]. Figure 5 shows a flowchart of the overall methodology. One-way ANOVA was performed followed by Newman-Keuls post-hoc analyses to test for differences between pathological and normal groups. A modified p value of <0.001 was considered statistically significant. Receiver operating characteristic (ROC) curves were constructed to describe the ability of each quantitative parameter to discriminate between eyes of pathological patients and normal healthy eyes. The parameters of interest were the thickness and fractal dimension of each intraretinal layer. Several discriminative diagnostic characteristics of the ROC curve were analyzed. These included the c -statistic (the concordance index, which is the area

under the ROC curve used to compare diagnostic power), the sensitivity, specificity, and the positive likelihood ratio (PLR, sensitivity/ $1 -$ specificity). The positive likelihood ratio (PLR) combines the sensitivity and specificity at the threshold value by dividing the proportion of true positives by the proportion of false positives. The PLR statistic indicates how likely it is that a case will have an abnormal test compared with a control. The AUROC calculations and statistical analyses were performed using the software package SPSS version 16 (SPSS Inc, Chicago, Illinois).

Results and discussion

A total of 117 eligible eyes (702 B-scans) were analyzed, which included a total of 74 healthy eyes (34 ± 12 years, 52 female, 22 male), and 43 eyes with mild diabetic retinopathy (MDR, 43 ± 17 years, 21 female, 22 male). The demographic and clinical characteristics of the study population are summarized in Table 1.

Table 2 shows the thickness and fractal dimension results as well as the outcome of the ROC and statistical analyses. Figure 6. shows a graphic interpretation of the predictive value of the parameters analyzed. The thickness of the GCL + IPL complex, OPL and OS were statistically significantly smaller (8%, 13% & 10%, respectively) in the

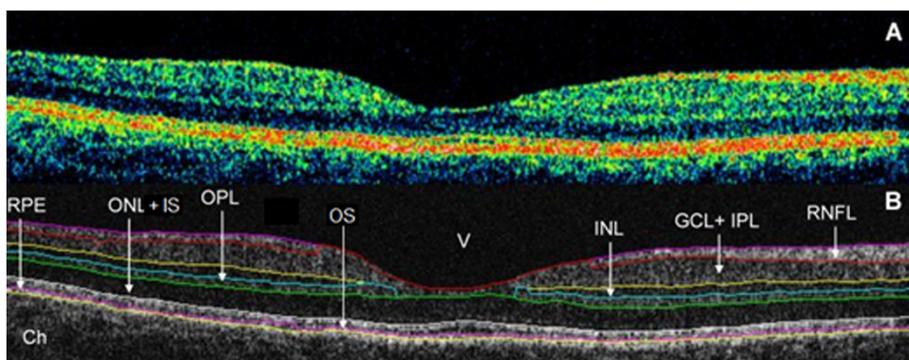
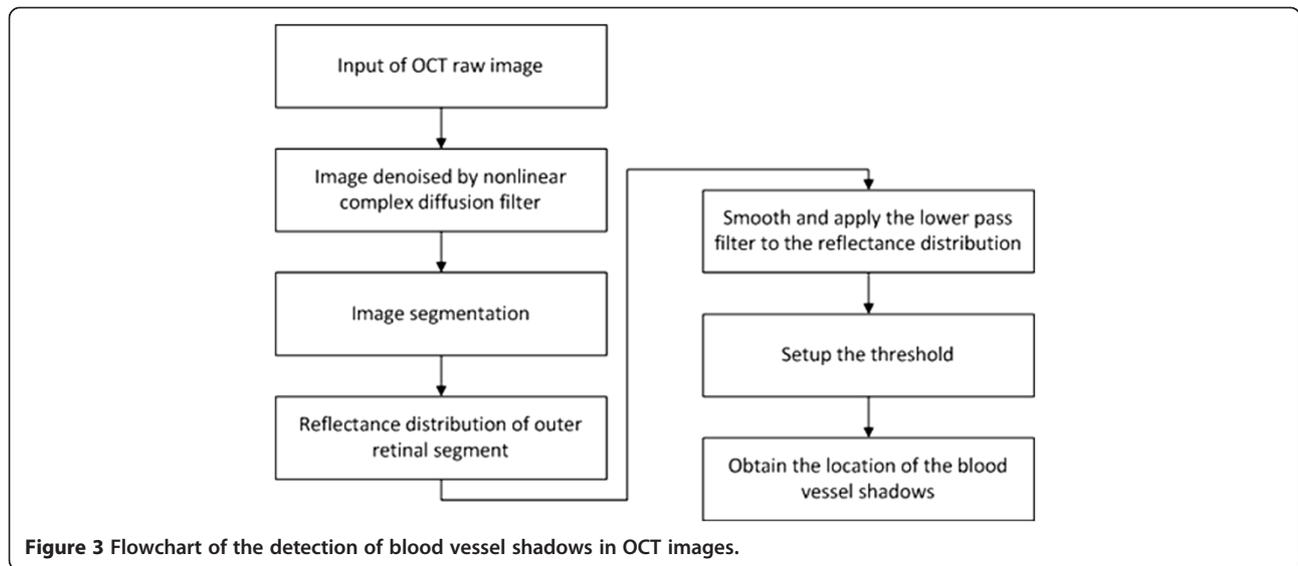


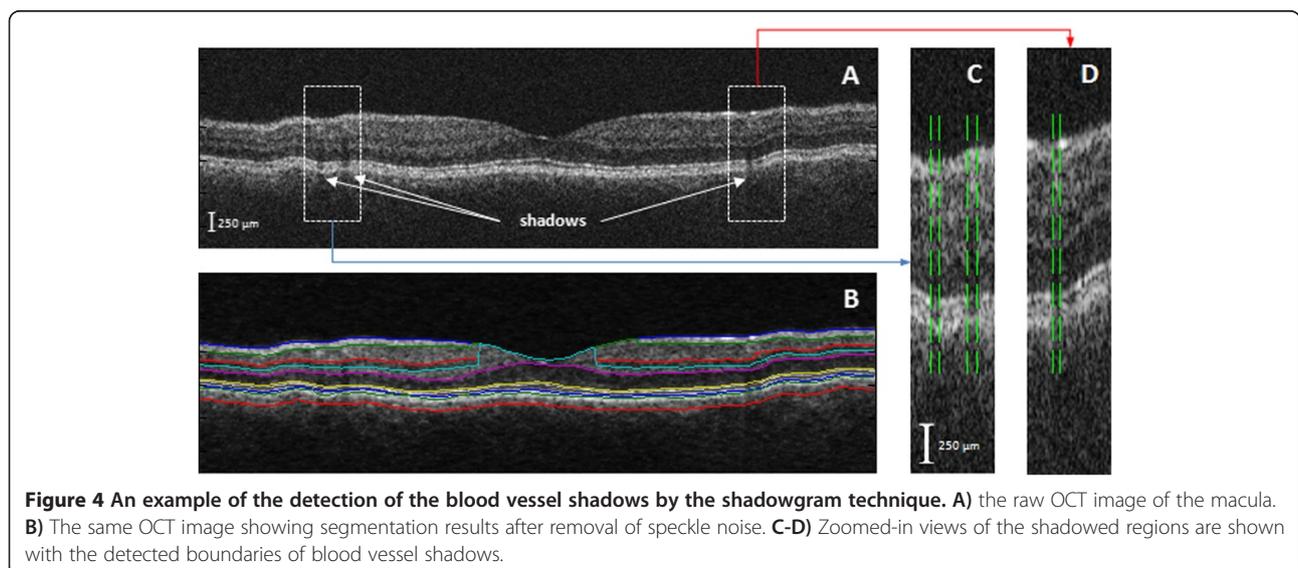
Figure 2 Macular image segmentation results using OCTRIMA. (A) The image of a healthy macula scanned by Stratus OCT. **(B)** The same OCT scan processed with OCTRIMA. Abbreviations: Ch, choroid; GCL + IPL, ganglion cell layer and inner plexiform layer complex; INL, inner nuclear layer; ONL + IS, combined outer nuclear layer and inner segment of photoreceptors; OS, outer segment of photoreceptors; OPL, outer plexiform layer; RNFL, retinal nerve fiber layer; RPE, retinal pigment epithelial layer; V, vitreous.



MDR eyes compared to normal healthy eyes (see Table 2). The thickness in other layers (except in the ONL + IS and RPE) showed a tendency towards thinning without reaching significance as compared to normal healthy eyes. Fractal dimension values were bigger for all the layers (except the GCL + IPL and INL) in MDR eyes compared to normal healthy eyes. When comparing MDR with normal healthy eyes, the highest AUROC values estimated for the fractal dimension were observed for GCL + IPL and INL (see Table 2). Moreover, the highest AUROC values estimated for the thickness measurements were observed for the OPL, GCL + IPL and OS. Particularly, compared to the standard thickness measurement, we found that the fractal dimension of the GCL + IPL complex might be a much better indicator for early DR diagnosis when comparing MDR eyes with control healthy eyes. (see Figure 6).

The maximum discrimination value for fractal dimension of 0.96 (standard error = 0.025) for the GCL + IPL complex was obtained at a FD ≤ 1.66 (cut off point, asymptotic 95% CI: lower-upper bound = 0.905-1.002). Therefore, there is a 96% probability the diabetic subject will have an abnormal GCL + IPL structure (i.e. disordered structure compared to normal healthy subjects). The ≤ 1.66 threshold coincides with the mean $\pm 2SD$ for the OCT measurements. At this value, the sensitivity for the GCL + IPL complex is 98% with a specificity of 88%. The positive likelihood ratio for GCL + IPL complex is 15.53, which increase the probability of early retinopathy development about 70%.

Our results suggest that the RNFL and GCL + IPL complex, OPL and OS are more susceptible to initial damage when comparing MDR with control healthy



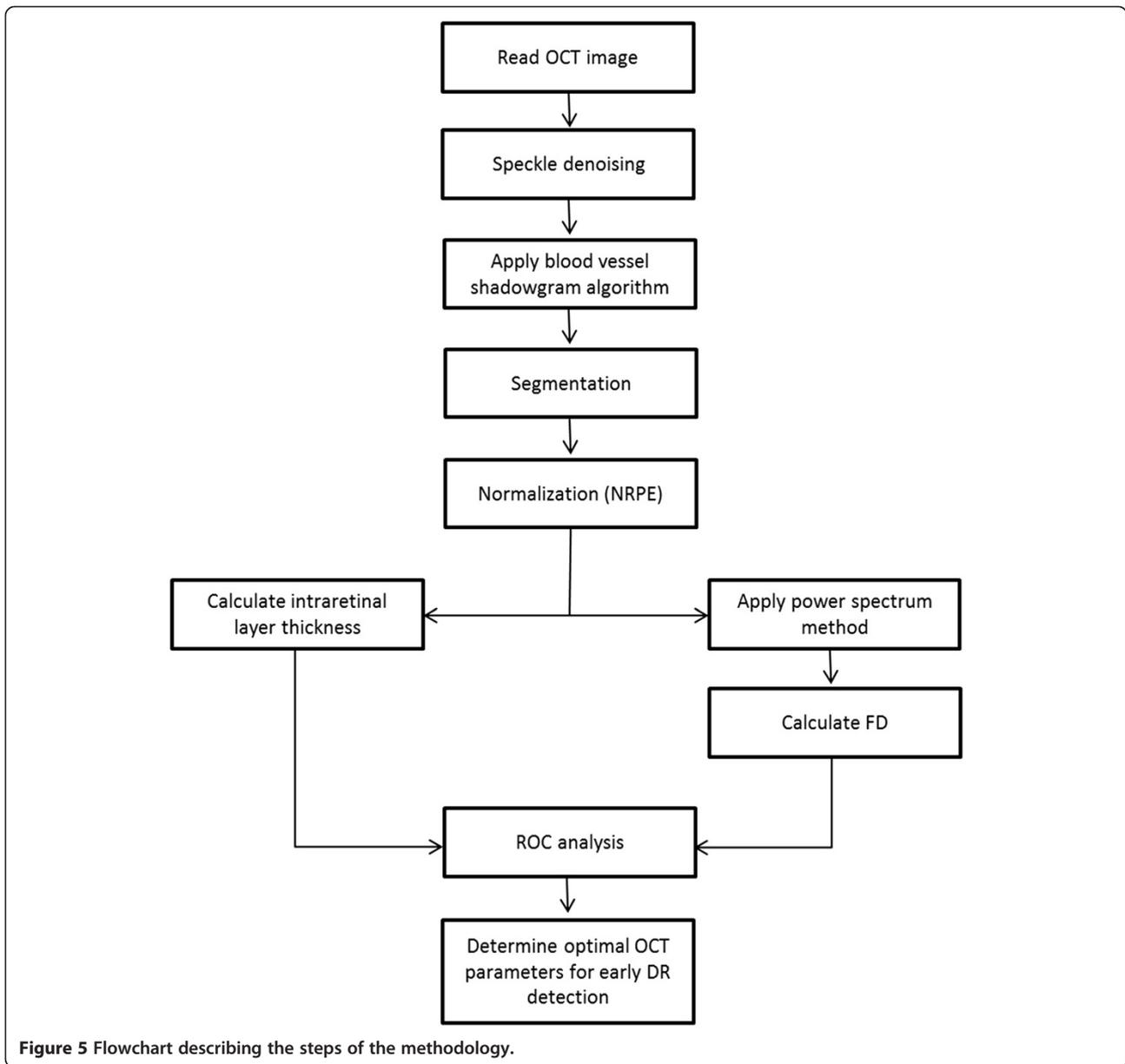


Table 1 Descriptive statistics of the study participants

Characteristic	Controls	MDR
Number of participants	41	29
Number of eyes	74	43
Age (years, mean ± SD)	34 ± 12	43 ± 17
Female, N (% total eyes)	52 (70%)	21 (49%)
Race (% Caucasian)	100	91
Hemoglobin A1c level (%)	-	8.51 ± 1.76
DM duration (years, mean ± SD)	-	22 ± 10
IOP (mmHg, mean ± SD)	14.5 ± 1.23	15.09 ± 1.56
BCVA	1.0 ± 0.00	0.97 ± 0.06
Total macular thickness (µm ± SD)	324.36 ± 10.27	297.40 ± 21.79

Abbreviations: SD standard deviation, BCVA best corrected visual acuity.

eyes. Particularly, the trend observed for the thickness (thinning) of the RNFL and GCL + IPL complex in MDR eyes might be associated with pathological metabolic changes in the retina and may reflect neurodegenerative changes in the diabetic retina. These findings also have possible implications for the early detection of macular damage in diabetes. Interestingly, our results showed for the first time that the thickness of the OPL in MDR eyes was significantly reduced compared with similar measures in normal healthy eyes. Interestingly, a significant decrease in fractal dimension was only observed for the GCL + IPL complex of MDR eyes compared to controls. This result is in agreement with previous reports showing a significant reduction of the fractal dimension during induced apoptosis

Table 2 Distribution statistics of thickness and fractal dimension

Intraretinal layer	Thickness (microns)					
	Healthy (mean ± SD)	MDR (mean ± SD)	AUROC (mean ± SE)	Asymptotic 95% confidence interval (Lower-upper bound)	Cutoff point	Positive likelihood ratio
RNFL	42.02 ± 2.11	41.38 ± 2.93	0.598 ± 0.059	0.483 - 0.713	41.03	1.51
GCL + IPL	78.30 ± 4.09	71.80 ± 8.22 [‡]	0.756 ± 0.05	0.657 - 0.855	75.88	2.90
INL	35.02 ± 1.60	35.05 ± 2.76	0.508 ± 0.061	0.388 - 0.627	34.86	1.33
OPL	41.30 ± 2.49	36.07 ± 3.45 [‡]	0.878 ± 0.041	0.797 - 0.958	38.24	6.48
ONL + IS	86.41 ± 5.21	88.39 ± 8.21	0.394 ± 0.055	0.285 - 0.503	86.86	1.07
OS	16.27 ± 3.06	14.40 ± 2.20 [‡]	0.688 ± 0.049	0.591 - 0.784	14.59	8.22
RPE	12.71 ± 1.32	12.76 ± 1.09	0.481 ± 0.054	0.375 - 0.588	12.40	3.52

Intraretinal layer	Fractal dimension					
	Healthy (mean ± SD)	MDR (mean ± SD)	AUROC (mean ± SE)	Asymptotic 95% confidence interval (Lower-upper bound)	Cutoff point	Positive likelihood ratio
RNFL	1.74 ± 0.04	1.78 ± 0.10 [‡]	0.393 ± 0.056	0.284 ± 0.503	1.74	1.02
GCL + IPL	1.68 ± 0.01	1.58 ± 0.05 [‡]	0.953 ± 0.025	0.905 - 1.002	1.66	15.53
INL	1.78 ± 0.01	1.76 ± 0.03 [‡]	0.785 ± 0.053	0.680 - 0.890	1.77	3.02
OPL	1.51 ± 0.01	1.56 ± 0.04 [‡]	0.111 ± 0.041	0.031 - 0.190	1.52	1.02
ONL + IS	1.78 ± 0.03	1.79 ± 0.04	0.336 ± 0.055	0.228 - 0.444	1.78	2.96
OS	1.70 ± 0.02	1.73 ± 0.04 [‡]	0.268 ± 0.047	0.177 - 0.359	1.71	1.00
RPE	1.68 ± 0.01	1.68 ± 0.01	0.433 ± 0.056	0.323 - 0.543	1.68	1.09

Note that mean ± SD, mean ± SE for groups ([‡] p < 0.001, SD: standard deviation, SE: standard error), AUROC, cutoff point, confidence interval and positive likelihood ratio values are also included for each variable analyzed. The Fractal Dimension of the GCL+IPL layer had the highest discrimination value (shown in bold).

throughout early apoptotic phases in breast cancer cells [28].

There are limitations to the present study, some of which might be improved in subsequent investigations. First, improved validation of the current methodology demands a larger patient population for analysis. Second, although the TD-OCT technology provides lower image resolution compared to advanced OCT technologies, the

six retinal layers were reliably assessed and were the standard when this study was initiated in 2007. However, better results might be expected with advanced OCT imaging technologies mentioned earlier and should be the standard for future studies [23,24]. Third, although texture measures of the retinal tissue are not standardized measures for detecting significant intraretinal changes, texture-based measures can be obtained from OCT

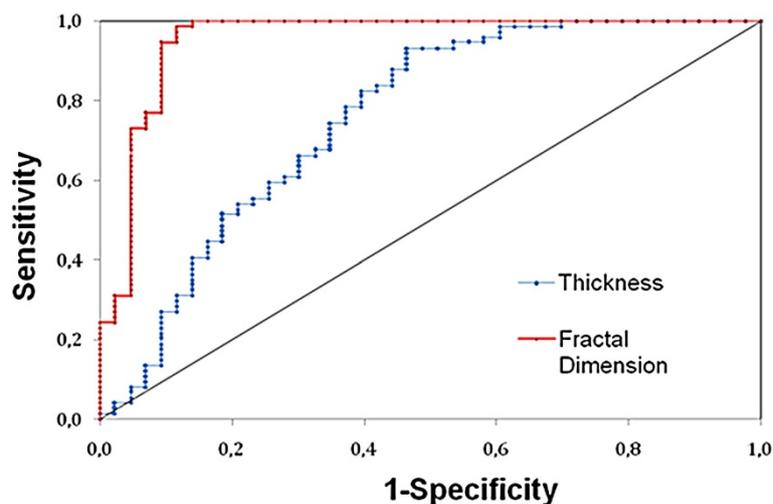


Figure 6 ROC curve showing the results of the sensitivity and specificity test. The GCL + IPL complex was used for classifying diabetic retinal tissue with early neural loss based on fractal dimension in OCT images. The AUROC is calculated to be 0.96.

intensity images. Therefore, as reported by previous studies comparing results between TD-OCT and SD-OCT devices, we expect the trends reported here to be replicated by advanced OCT devices [29,30]. Fourth, FD of the GCL + IPL was not always discriminative. For example, the discrimination power of the FD parameter of the GCL + IPL failed to classify Hispanic subjects. Our study population was overwhelmingly white and Caucasian (see Table 1). Our model using the FD of the GCL + IPL as a risk predictor for early retinopathy showed a good discrimination with high sensitivity and specificity for Caucasian participants. However, a homogenous population does not allow us to probe into the whole scope of the variability in DR risk. Our normal healthy (MDR) subjects were 100 (91)% white/Caucasians (see Table 1). Therefore, our model, which ignored ethnicity and race, could still discriminate well in a population made up entirely of white and Caucasian people, since in these cases ethnicity and/or race is not relevant to their risk relative to one another. In a population of mixed ethnicity, it would discriminate less well the larger the minority group. As a result, calibration and reclassification tasks at specific thresholds reflecting race/ethnicity variability should be in place when using a heterogeneous population in future studies. Fifth, the MDR group was not age-matched to controls in our study. Although aging is known to be associated with loss of complexity in organ structures of the human body due to functional loss, [31,32] earlier works did not find any correlation between aging and FD of the retinal vasculature [33,34]. These earlier studies were based on the box-counting method to calculate the FD, which is not the best technique to estimate FD in the presence of segmentation errors due to background noise [35]. However, recent evidence supporting rarefaction of the retinal vasculature has been reported by Zulfazal et al. [36]. However, the possibility of intraretinal changes as a secondary effect to aging cannot be discarded when comparing MDR with healthy eyes. Therefore, additional work is needed to include more subjects with a broader age range, to fully appreciate the effects on FD from this aging factor using OCT images. Moreover, separating norms by gender is required when designing future studies. Fifth, study parameters were measured cross-sectionally and not longitudinally. Therefore, future studies should investigate whether changes attributed to age could be due to other factors (e.g. sex and race) that may vary between subjects. Six, because patients enrolled in our study were 91-100% Caucasian, results cannot be generalized to other racial populations. Despite these basic limitations, the data presented here reveal that it may be possible to differentiate MDR eyes from normal healthy eyes by analyzing the OCT signal using fractal analysis [37]. More comprehensive studies including investigations on larger subject populations

and longitudinal studies using advanced OCT technologies are needed to confirm our preliminary results.

In summary, we have shown that it may be possible to differentiate MDR eyes from normal healthy eyes by analyzing the OCT signal using fractal analysis. The highest AUROC values estimated for the fractal dimension were observed for the GCL + IPL complex in diabetic patients. Particularly, fractal dimension was smaller for this complex in diabetic eyes. A smaller value of this parameter in case of pathologic retinal deformation, or degradation due to apoptosis (cell death) is expected [26]. As cells undergo this apoptosis process, bodies within the cell, like the nucleus or mitochondria, go through structural changes. The use of fractal analysis for classification of diabetes-induced retinal damage in OCT clinical data could potentially provide additional diagnostic information for early detection and progression of DR.

Conclusions

A potential improvement in the clinical application of OCT to eye diseases is the quantification of the anatomic changes along with the dysfunction of the cellular layers of the neurosensory retina. Our preliminary results suggest that the fractal dimension of the intraretinal layers might provide useful information to differentiate MDR eyes, which are characterized by neurodegeneration at the early stages, from healthy eyes in addition to the structural information. Particularly, the differentiation between normal and abnormal retinal tissue may improve understanding on the sequence of events involved in the visual field defects and provide new insights of the clinical relevance of certain specific morphological features. Further research is warranted to determine how this approach may be used to improve diagnosis of diabetic retinopathy and retinal dysfunction in DR. Specifically, we will have to prove that the fractal dimension is able to discriminate pathological eyes from healthy eyes with higher sensitivity and specificity compared to standard thickness parameters. Thus, the future evaluation of this method using a larger set of data would ultimately lead to a more rational and effective approach to therapy and improved diagnosis. In addition, a more effective classification analysis could be implemented by incorporating other metrics such as depth-dependent attenuation rate and a three-dimensional fractal-based method for 3D OCT data [25,38-41]. It is important to mention that high resolution is vital for extracting information from OCT images affected by speckle noise. Therefore, the results in our study may be to some extent improved by the utilization of an ultrahigh resolution OCT device [42].

Abbreviations

OCT: Optical coherence tomography; ROC: Receiver operating characteristic; AUROC: Area under the ROC curve; MDR: Mild diabetic retinopathy; PLR: Positive likelihood ratio; OCTRIMA: OCT retinal image analysis;

RNFL: Retinal nerve fiber layer; GCL + IPL: Ganglion cell and inner plexiform layer complex; INL: Inner nuclear layer; OPL: Outer plexiform layer; ONL: Outer nuclear layer; ONL + IS: Inner photoreceptor segment; OS: Outer photoreceptor segment; RPE: Retinal pigment epithelium; FD: Fractal dimension; MS: Multiple sclerosis; SD: Standard deviation; SE: Standard error; 2D: Two-dimensional; 3D: Three-dimensional.

Competing interests

The University of Miami and Dr. Cabrera DeBuc hold a pending patent used in the study and have the potential for financial benefit from its future commercialization. All other authors of the paper report no disclosures.

Authors' contributions

GMS, ET, WES and AS collected clinical data. ET, RT, LL, BEV, VÖ analyzed the data. DCD interpreted data. ET performed the statistical analysis. RT, GMS and DCD drafted the manuscript. DCD and GMS designed and supervised this study. All authors read and approved the final manuscript.

Acknowledgements

This study was supported in part by a Juvenile Diabetes Research Foundation Grant, a NIH Grant No. NIH R01EY020607, a NIH Center Grant No. P30-EY014801, by an unrestricted grant to the University of Miami from Research to Prevent Blindness, Inc., by the Zsigmond Diabetes Fund of the Hungarian Academy of Sciences and by an Eötvös Scholarship by the Hungarian Scholarship Fund. Authors acknowledge Wei Gao, PhD for his contribution with data analysis to this study. The University of Miami and Dr. Cabrera DeBuc hold a pending patent used in the study and have the potential for financial benefit from its future commercialization.

Author details

¹Department of Ophthalmology, Faculty of Medicine Semmelweis University, Budapest, Hungary. ²Miller School of Medicine, Bascom Palmer Eye Institute, University of Miami, Miami, Florida 33136, USA. ³Faculty of Science, Department of Physics, University of Dschang, Dschang, Cameroon. ⁴2nd Department of Internal Medicine, Faculty of Medicine Semmelweis University, Budapest, Hungary.

Received: 21 March 2013 Accepted: 18 June 2014
Published: 1 September 2014

References

- Huang D, Swanson EA, Lin CP, Schuman JS, Stinson WG, Chang W, Hee MR, Flotte T, Gregory K, Puliafito CA, Fujimoto JG: **Optical coherence tomography.** *Science* 1991, **254**(5035):1178–1181.
- Cabrera DeBuc D: *A review of algorithms for segmentation of retinal image data using optical coherence tomography.* Pei-Gee Ho INTECH: In Image Segmentation. Edited by Dr; 2011. ISBN 978-953-307-228-9.
- Cabrera Fernandez D, Salinas HM, Puliafito CA: **Automated detection of retinal layer structures on optical coherence tomography images.** *Opt Express* 2005, **13**(25):10200–10216.
- Debuc DC, Somfai GM: **Early detection of retinal thickness changes in diabetes using optical coherence tomography.** *Med Sci Monitor* 2010, **16**(3):Mt15–Mt21.
- Tatrai E, Simo M, Iljicsov A, Nemeth J, Debuc DC, Somfai GM: **In vivo evaluation of retinal neurodegeneration in patients with multiple sclerosis.** *PLoS One* 2012, **7**(1):e30922.
- Gossage KW, Tkaczyk TS, Rodriguez JJ, Barton JK: **Texture analysis of optical coherence tomography images: feasibility for tissue classification.** *J Biomed Opt* 2003, **8**(3):570–575.
- Gao W, Tatrai E, Ölvedy V, Varga B, Laurik L, Somogyi A, Somfai GM, Debuc DC: **Investigation of changes in thickness and reflectivity from layered retinal structures of healthy and diabetic eyes with optical coherence tomography.** *Hemoglobin* 2011, **100**(100):91.
- Gao W, Tatrai E, Somfai GM, Debuc DC: **Assessing the performance of optical properties determination of intraretinal layers in healthy normal and type 1 diabetic eyes using optical coherence tomography.** *Invest Ophthalmol Vis Sci* 2011, **52**(6):3689.
- Gao W, Tatrai E, Somfai GM, Cabrera DeBuc D: **Evaluation of intraretinal scattering measurements in eyes of healthy and type 1 diabetic subjects using optical coherence tomography.** *Invest Ophthalmol Vis Sci* 2010, **51**(5):1786.
- Mandelbrot BB: *Fractals: form, chance, and dimension.* San Francisco: W. H. Freeman; 1977.
- Nailon WH: **Texture analysis methods for medical image characterisation.** In *Biomedical Imaging.* Edited by Mao Y. (Rijeka): InTec; 2010:75–100.
- Daxer A: **Characterisation of the neovascularisation process in diabetic retinopathy by means of fractal geometry: diagnostic implications.** *Albrecht Von Graefes Arch Klin Exp Ophthalmol* 1993, **231**(12):681–686.
- Daxer A: **The fractal geometry of proliferative diabetic retinopathy: implications for the diagnosis and the process of retinal vasculogenesis.** *Curr Eye Res* 1993, **12**(12):1103–1109.
- Avakian A, Kalina RE, Sage EH, Rambhia AH, Elliott KE, Chuang EL, Clark JI, Hwang JN, Parsons-Wingerter P: **Fractal analysis of region-based vascular change in the normal and non-proliferative diabetic retina.** *Curr Eye Res* 2002, **24**(4):274–280.
- Landini G, Murray PI, Misson GP: **Local connected fractal dimensions and lacunarity analyses of 60 degrees fluorescein angiograms.** *Invest Ophthalmol Vis Sci* 1995, **36**(13):2749–2755.
- Cabrera DeBuc D, Tchtngna R: *Photoreceptor rearrangement and vision restoration in eyes with outer retinopathy: Quantitative assessment by fractal analysis.* Pennsylvania: APS March Meeting Pittsburgh; 2009.
- Kim P, Iftekharuddin K, Gunvant P, Tóth M, Holló G, Essock E: **Efficacy of fractal analysis in identifying glaucomatous damage.** In *Pro. SPIE 7627, Medical Imaging. Image Perception, Observer Performance, and Technology Assessment, 76271G;* 2010. (February 23, 2010); doi:10.1117/12.848428.
- Schmoll T, Singh AS, Blatter C, Schriefl S, Ahlers C, Schmidt-Erfurth U, Leitgeb RA: **Imaging of the parafoveal capillary network and its integrity analysis using fractal dimension.** *Biomedical optics express* 2011, **2**(5):1159–1168.
- Diabetic Retinopathy Clinical Research N: **Diabetic retinopathy study. Report Number 7. A modification of the Airlie House classification of diabetic retinopathy.** *Invest Ophthalmol Vis Sci* 1981, **21**(Pt 2):210–226.
- Group ETRDRSR: **Photocoagulation for diabetic macular edema. Early Treatment Diabetic Retinopathy Study report number 1.** *Arch Ophthalmol* 1985, **103**(12):1796–1806.
- Wilkinson CP, Ferris FL 3rd, Klein RE, Lee PP, Agardh CD, Davis M, Dills D, Kampik A, Parajasegaram R, Verdaguer JT: **Proposed international clinical diabetic retinopathy and diabetic macular edema disease severity scales.** *Ophthalmology* 2003, **110**(9):1677–1682.
- Hasegawa M, Liu J, Okuda K, Nunobiki M: **Calculation of the fractal dimensions of machined surface profiles.** *Wear* 1996, **192**(1):40–45.
- Debuc DC, Salinas HM, Ranganathan S, Tatrai E, Gao W, Shen M, Wang J, Somfai GM, Puliafito CA: **Improving image segmentation performance and quantitative analysis via a computer-aided grading methodology for optical coherence tomography retinal image analysis.** *J Biomed Opt* 2010, **15**(4):046015.
- DeBuc DC, Somfai GM, Ranganathan S, Tatrai E, Ferencz M, Puliafito CA: **Reliability and reproducibility of macular segmentation using a custom-built optical coherence tomography retinal image analysis software.** *J Biomed Opt* 2009, **14**(6):064023.
- Cabrera DeBuc D, Gao W, Tatrai E, Laurik L, Varga B, Olvedy V, Smiddy W, Tchtngna R, Somogyi A, Somfai G: **Extracting Diagnostic Information from Optical Coherence Tomography Images of Diabetic Retinal Tissues Using Depth-dependent Attenuation Rate and Fractal Analysis.** In *Biomedical Optics. Proceedings of SPIE, Miami, Florida United States; 01/2012,* http://dx.doi.org/10.1364/BIOE.2012.BTu3A.74: Optical Society of America; 2012.
- Salinas HM, Fernandez DC: **Comparison of PDE-based nonlinear diffusion approaches for image enhancement and denoising optical coherence.** *IEEE transaction on medical imaging* 2007, **26**(6):761–771.
- Wehbe H, Ruggeri M, Jiao S, Gregori G, Puliafito CA, Zhao W: **Automatic retinal blood flow calculation using spectral domain optical coherence tomography.** *Opt Express* 2007, **15**(23):15193–15206.
- Castelli C, Losa GA: **Ultrastructural complexity of nuclear components during early apoptotic phases in breast cancer cells.** *Anal Cell Pathol* 2001, **23**(1):1–9.
- van Dijk HW, Verbraak FD, Kok PH, Garvin MK, Sonka M, Lee K, Devries JH, Michels RP, van Velthoven ME, Schlingemann RO, Abramoff MD: **Decreased retinal ganglion cell layer thickness in patients with type 1 diabetes.** *Invest Ophthalmol Vis Sci* 2010, **51**(7):3660–3665.
- van Dijk HW, Verbraak FD, Kok PH, Stehouwer M, Garvin MK, Sonka M, DeVries JH, Schlingemann RO, Abramoff MD: **Early neurodegeneration in**

- the retina of type 2 diabetic patients. *Invest Ophthalmol Vis Sci* 2012, **53**(6):2715–2719.
31. Lipsitz LA, Goldberger AL: **Loss of 'complexity' and aging.** *JAMA* 1992, **267**(13):1806–1809.
 32. Goldberger AL, Amaral LA, Hausdorff JM, Ivanov P, Peng CK, Stanley HE: **Fractal dynamics in physiology: alterations with disease and aging.** *Proc Natl Acad Sci U S A* 2002, **99**(Suppl 1):2466–2472.
 33. Family F, Masters BR, Platt DE: **Fractal pattern formation in human retinal vessels.** *Physica D: Nonlinear Phenomena* 1989, **38**(1):98–103.
 34. Masters BR: **Fractal analysis of the vascular tree in the human retina.** *Annu Rev Biomed Eng* 2004, **6**:427–452.
 35. Liew G, Wang JJ, Mitchell P, Wong TY: **Retinal vascular imaging: a new tool in microvascular disease research.** *Circulation Cardiovascular imaging* 2008, **1**(2):156–161.
 36. Azemin MZC, Kumar DK, Wong TY, Wang JJ, Mitchell P, Kawasaki R, Wu H: **Age-related rarefaction in the fractal dimension of retinal vessel.** *Neurobiol Aging* 2012, **33**(1):194. e191–194. e194.
 37. Somfai GM, Tátrai E, Laurik L, Varga B, Ölvedy V, Jiang H, Wang J, Smiddy WE, Somogyi A, DeBuc DC: **Automated classifiers for early detection and diagnosis of retinopathy in diabetic eyes.** *BMC Bioinformatics* 2014, **15**:106. doi:10.1186/1471-2105-15-106.
 38. Faber D, van der Meer F, Aalders M, van Leeuwen T: **Quantitative measurement of attenuation coefficients of weakly scattering media using optical coherence tomography.** *Opt Express* 2004, **12**(19):4353–4365.
 39. Popescu DP, Flueraru C, Mao Y, Chang S, Sowa MG: **Signal attenuation and box-counting fractal analysis of optical coherence tomography images of arterial tissue.** *Biomedical optics express* 2010, **1**(1):268–277.
 40. Kestener P, Arneodo A: **Three-dimensional wavelet-based multifractal method: the need for revisiting the multifractal description of turbulence dissipation data.** *Phys Rev Lett* 2003, **91**(19):194501.
 41. Veneziano D, Moglen G, Bras R: **Multifractal analysis: pitfalls of standard procedures and alternatives.** *Phys Rev E* 1993, **52**(2):1387–1398.
 42. Wang Y, Jiang H, Shen M, Lam BL, DeBuc DC, Ye Y, Li M, Tao A, Shao Y, Wang J: **Quantitative analysis of the intraretinal layers and optic nerve head using ultra-high resolution optical coherence tomography.** *J Biomed Opt* 2012, **17**(6):066013.

doi:10.1186/1471-2105-15-295

Cite this article as: Somfai et al.: Fractal-based analysis of optical coherence tomography data to quantify retinal tissue damage. *BMC Bioinformatics* 2014 **15**:295.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

